

# Clinical Use of Polihexanide on Acute and Chronic Wounds for Antisepsis and Decontamination

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## Key Words

Polihexanide · Antisepsis · Decontamination · Wound cleansing · Chronic wounds · Acute wounds

## Abstract

Polihexanide is an antimicrobial compound suitable for clinical use in critically colonized or infected acute and chronic wounds. Its beneficial characteristic is attributable particularly to its broad antimicrobial spectrum, good cell and tissue tolerability, ability to bind to the organic matrix, low risk of contact sensitization, and wound healing promoting effect. In addition, no development of microorganism resistance during polihexanide use has been detected to date, nor does this risk appear imminent. The aim of therapy using polihexanide is to reduce the pathogen burden in a critically colonized or infected acute or chronic wound. An increasing number of articles on the subject of wound antisepsis with polihexanide can be found in the medical literature. However, there is still little published information on the practical use of polihexanide-containing wound antiseptics. The purpose of this review article is to describe the handling and the different possibilities of use of polihexanide-containing preparations, including the currently approved indications, contraindications and reservations. The use of polihexanide is not the only therapeutic option in management of

wounds; therefore, priority is also given to prior surgical debridement and clarification of the cause of the underlying disease, including appropriate therapy.

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## Background

The term ‘antisepsis’ was first coined at the end of the 18th century by the English military physician John Pringle (1707–1782) to apply to substances that were able to prevent putrefaction [1]. With the continuation of antiseptic measures in surgery, Sir Joseph Lister (1827–1912) subsequently made the prevention of postoperative complications a central theme internationally in surgery [2].

Lister flushed surgical wounds with carbolic acid, and combined this procedure with dressings soaked in carbolic acid. Finally, he introduced the spray technique, in which the entire operation field was sprayed regularly with a carbolic acid spray. With the aid of these measures, the mortality rate after amputations and open bone fractures was reduced from 60% to less than 10% [3].

Later, however, the substance revealed its severe side effects resulting from systemic absorption with local tissue damage and general symptoms of toxicity. As a result, Billroth and Kocher formulated their famous warning

against the use of carbolic acid, although they questioned only the substance and not the prevention principle of antiseptics. In 1881, Kocher rehabilitated Lister, and described his ideas as an 'immortal service'. He accepted that prevention of wound infection could only have been established through Lister's pioneering work [4].

Thus, the preventive principle of wound antiseptics was born. However, the search continued for suitable substances that could be used safely. The procedure was also fundamentally improved technically. Alexis Carrel (1873–1944) extended simple wound irrigation and developed the specific, continuous or intermittent wound drainage [5].

In the following years, a wide variety of irrigation solutions were recommended, beginning with low-concentrated zinc-chlorine solutions (Kocher 1881), sublimate (Robert Koch 1883), iodoform (Billroth 1885), Dakin's (chlorine-containing) solution (Carrel 1917), azo dyes (Rivanol 1920s), and sulphonamides (1930s). This progress repeatedly bore fruit to some degree, but no truly satisfactory substance could be found. The main problems were not so much the antiseptic efficacy, but the side effects that occurred as a result of systemic absorption, and tissue toxicity that occurred repeatedly. For example, it was found in the case of the sulphonamides that formation of granulation tissue and thus wound healing overall is inhibited. With all the mentioned substances that were in use, the cases of postoperative wound infections were considerably reduced. Nevertheless, following the use of chlorine solutions, for example, up to 90% of the patients treated with these died after a few years [6].

The discovery of antibiotics led to the principles of wound antiseptics very quickly being forgotten. The previously constant search for new, suitable antiseptic substances was abruptly interrupted. The optimism induced by the possibilities of systemic antibiotic therapy led to rapidly increasing systemic and topical use of antibiotics. For some years now, we have been seeing the long-term consequence of this in the form of the increasing resistance of bacteria to antibiotics.

However, doubts were expressed at an early stage as to whether topical treatment with antibiotics could be justified. In 1961, the leading article in the *Journal of Clinical Pathology* stated: 'The administration of antibiotics, which often betrays a shocking lack of insight, has led to the occurrence of resistant bacteria among the banal bacteria. The return to the elementary principles of topical wound treatment and wound antiseptics is therefore what is called for' [7]. This assessment is still just as relevant today. As a result of these developments, we are now experiencing a comeback of antiseptics.

Willenegger and Good recognized the fundamental possibility of use of the surface disinfectant polihexanide, known in the industry since the 1960s, and characterized by its good bactericidal efficacy. They hit on the idea of adding polyethylene glycol to a biguanide (polihexanide) that was already known for surface disinfection and had a good bactericidal efficacy and no known toxicity [8]. The antimicrobial mechanism of action of polihexanide was clarified [9, 10; Kaehn et al., this supplement issue]. The work published by Werner [11] finally led to the recognition of this substance as a wound antiseptic in Central Europe. Thus, a surface disinfectant and swimming pool disinfectant, unsuitable for use in a wound, became a polihexanide-containing solution for wound antiseptics. This topical anti-infective agent exhibited a hitherto unknown therapeutic margin [12], and in 1994 Switzerland was the first country in the world to approve it as a medicinal product.

The characteristics of polihexanide are comprehensively described in another article in this supplement [Hübner and Kramer, this supplement issue]. The first randomized controlled trial was conducted by Schmit-Neuerburg et al. [13, 14]. They studied the effect of gauze compresses soaked in 0.04% polihexanide in comparison with Ringer solution in a randomized, controlled, double-blind study in 85 patients. The group treated with polihexanide showed improved wound healing with a significantly more rapid reduction of Gram-positive organisms and also better tissue compatibility than was the case in the control group.

By far the most extensive collection of data on the use of polihexanide-containing external preparations is presented by Roth et al. [15]. These data from a retrospective, open-label, controlled, multicentre, randomized cohort study involving 7,862 patients, mainly with agricultural, severe and contaminated soft-tissue injuries, compare the rate of postoperative wound infections with polihexanide, PVP-iodine, Ringer solution and hydrogen peroxide after only a single antiseptic irrigation with prior thorough surgical debridement (irrespective of whether the wound was primarily closed or had to be left partially open or completely open). With the use of polihexanide, the infection rate could be significantly reduced in comparison to PVP-iodine, Ringer solution and hydrogen peroxide.

Although there are an increasing number of articles on the subject of wound antiseptics with polihexanide in the medical literature, there is little detailed information on the use of polihexanide-containing wound antiseptics in practice. The purpose of this review article is to describe the handling and the different possibilities of use

**Table 1.** BI for selected antiseptic substances after 30 min contact in MEM cell culture medium in the presence of 10% fetal bovine serum [30]

Substance	BI	
	L929 cells/ < <i>E. coli</i>	L929 cells/ <i>S. aureus</i>
Octenidine dihydrochloride	1.8	1.5
Polihexanide	1.5	1.3
PVP-iodine (referring to I <sub>2</sub> )	1.0	0.9
Chlorhexidine digluconate	0.8	0.7
Silver protein (referring to Ag)	0.13	–
Silver sulphadiazine	<<0.004	–
AgNO <sub>3</sub>	<<0.002	–

BI = Biocompatibility index.

of polihexanide-containing preparations, including the currently approved indications, contraindications and reservations.

#### *Properties of Polihexanide-Containing Preparations Relevant for the Treatment of Wounds*

Depending on the concentration and application form, there are various advantages for polihexanide that in view of efficacy and tolerability predestine the substance for use in critically colonized as well as locally infected acute and chronic wounds. In addition to its simplicity of use, these advantages also include the following:

- broad antimicrobial spectrum [13, 14, 16–19, 21–24, 29];
- sustained and post-antiseptic effect [19, 20];
- concentration-dependent promotion of wound healing in vitro und in vivo [21–27];
- specific mechanism of action against acidic lipids of the bacterial membrane with an only slight effect on the neutral lipids in human cell membranes [28];
- favourable biocompatibility index (BI) >1 [29]. For comparison of the tolerability of wound antiseptics, the BI is suitable [30]. A condition of this is that the testing for microbicides and cytotoxicity must be carried out under identical test conditions. Cell culture media with a protein content of 6–7 g/l serum albumin and a physiological electrolyte concentration are largely equivalent in composition to the protein and electrolyte contents of wound fluids. The BI is obtained from the quotient of IC<sub>50</sub>, i.e. the molar concentration at which 50% of the test cells in the cytotoxicity test are no longer vital, and the molar concentration that in the

quantitative suspension test against bacterial test microorganisms results in a reduction of at least 3 log steps. The BI is thus a dimensionless figure and permits a comparison of tolerability. A value >1 describes good tolerability, a value <1 poor microbicidal efficacy, combined with high cytotoxicity. In table 1, BIs for selected antiseptic substances for the system mouse fibroblasts (L929 cells, ATCC CCL1)/*Escherichia coli* (ATCC 11229) and *Staphylococcus aureus* (ATCC 6538) are given;

- no known toxic and absorptive risks [19];
- to date, no known development of resistance [19, 31];
- reduction of biofilm [32–34] and fibrin formation [35–37];
- good clinical tolerability [17, 38–41];
- additional anti-inflammatory properties [42];
- low risk of contact sensitizations as well as of type I sensitizations [19, 43–46].

#### *Contraindications*

With the present level of knowledge, polihexanide may not be used in the following indications or situations [19, 47]:

- as an irrigation solution in the peritoneal cavity;
- as an antiseptic joint irrigation solution in the use concentration recommended by the manufacturer (hyaline cartilage toxicity);
- in the entire central nervous system including the meninges and intraluminally;
- in the middle ear or inner ear or intraocularly;
- in the first 4 months of a pregnancy (later only after critical evaluation of the benefit/risk ratio);
- in cases of allergy to polihexanide.

#### *Overview of Available Polihexanide Application Forms*

Worldwide there are now a large number of polihexanide-containing products on the market. These include antiseptic solutions, wound irrigation solutions, hydrogels and wound dressings. Tables 2–5 give an overview of polihexanide application forms currently available on the world market, although they are subject to considerable national differences.

As a result of regulatory intricacies that can only be understood by the legal profession, there is currently a marketing authorization situation that is in some cases confusing, so that even identical products are classified differently, e.g. Lavasorb® (in Switzerland, Germany and Austria: a class IIb medical device) and Lavasept® (in Switzerland: a medicinal product).

**Table 2.** Wound antiseptics containing polihexanide and given medicinal product status

Product	Polihexanide concentration, %	Excipients	Manufacturer	Available in
Lavasept® concentrate	20.0	Macrogol 4000, Ringer's solution	B. Braun	Germany, Russia, Switzerland
Lavasept® solution	0.04	Macrogol 4000, Ringer's solution	B. Braun	Switzerland
Serasept® solution 1	0.02	Macrogol 4000, Ringer's solution	Serag-Wiesner	Germany
Serasept® solution 2	0.04			

**Table 3.** Wound irrigation solutions/wound cleansers containing polihexanide and given medical device status

Product	Polihexanide concentration, %	Excipients	Manufacturer	Available in
Lavasorb® solution	0.04	Macrogol 4000, Ringer's solution	Fresenius	EU
Lavanid® solution 1	0.02	Macrogol 4000, Ringer's solution	Serag-Wiesner	EU
Lavanid® solution 2	0.04			
Prontosan® Wound Irrigation Solution	0.1	Undecylenamidopropyl betaine (surfactant)	B. Braun	Australia, Brazil, Canada, Chile, Croatia, EU, Hong Kong, Israel, Malaysia, Norway, Peru, Philippines, Russia, Serbia, Switzerland, Thailand, Turkey, USA

**Table 4.** Hydrogels containing polihexanide for wound cleansing (medical devices)

Product	Polihexanide concentration, %	Excipients	Manufacturer	Available in
Lavanid® wound gel	0.04	Macrogol, glycerin, hydroxyethyl celluloses, Ringer's solution	Serag-Wiesner	EU
Prontosan® wound gel	0.1	Undecylenamidopropyl betaine (surfactant), glycerol, hydroxyethyl celluloses	B. Braun	Australia, Brazil, Canada, Chile, Croatia, EU, Hong Kong, Israel, Malaysia, Norway, Peru, Russia, Serbia, South Africa, Switzerland, Thailand, Turkey, USA

In clinical practice, the usually applied concentrations of polihexanide solutions for wound antiseptics are 0.01, 0.02 and 0.04%. In addition, wound irrigation solutions or polihexanide-containing dressings show concentrations of 0.1–0.5%. In the case of a new biocellulose moist dressing with a concentration of 0.3% polihexanide in the fibres, in *in vitro* studies the release of active substance in steady state is approximately 0.13% [48]. The products are only to be used topically, for example in the form of irrigations (lavages), irrigation-suction drains as well as moist wound coverings. As polihexanide has a relatively

slow onset of effect, and the microorganisms react with different sensitivity as a function of time, it must be ensured that with full coverage of the wound base a minimum contact time of 10–15 min is adhered to [19, 49]. Its use may be based on therapeutic indication, but also for prophylactic reasons.

#### *Special Aspects of Practical Use*

##### *Starting Point of Therapy*

Precisely with regard to the microbial condition, acute and chronic wounds can change dramatically within a

**Table 5.** Wound dressings containing polihexanide (medical devices)

Product	Polihexanide concentration, %	Dressing material	Manufacturer	Available in
Telfa® A.M.D.	0.2	dry cotton pad with polyester film	Covidien	EU, USA
Excilon® A.M.D.	0.2	dry cotton fleece compress	Covidien	EU, USA
Kerlix® A.M.D.	0.2	dry cotton gauze/pad	Covidien	EU, USA
Kendall® A.M.D.	0.5	polyurethane foam dressing	Covidien	EU, USA
Suprasorb® X+PHMB	0.3	antimicrobial moist cellulose dressing ca. 4% bacterial produced cellulose in water ca. 96%	Lohmann & Rauscher	EU
XCell			Xylos	USA

**Table 6.** Procedure of the practical application of intra-operative irrigation

- (1) Positioning of drain with infection or risk for infection after surgical cleansing
- (2) Beginning of irrigation
- (3) Flushing of still putrid exudate from the drain
- (4) Completion of the irrigation with clean conditions
- (5) Application of the dressing

short time. In such a case, it is crucial for the success of treatment that adequate therapy is initiated without delay. Therapy is indicated from the stage of critical colonization. Critical colonization is a state of transition from colonization (frequent in the case of secondarily healing wounds) to infection. By means of specific topical antimicrobial therapy the transition into a wound infection can often be prevented [50]. Antisepsis can only support wound cleansing, it can never replace it.

#### Duration of Treatment and Endpoint of Antiseptic Therapy and Prophylaxis

The prime objective of the treatment is to eliminate the clinical signs of infection. The duration of topical antiseptic treatment is usually 2–5 days, and in general should not exceed 14–21 days [51, 52]. If the signs of infection do not recede, the efficacy of the measures used must be checked, and if necessary new measures must be initiated. If therapy is successful, treatment must be continued according to the general principles of moist wound

treatment, but without the addition of an antimicrobial preparation.

This approach is based on the consideration that the use of previous antiseptics whose antimicrobial efficacy is necessarily always accompanied by some degree of cytotoxicity is only justified if a microbial burden that is affecting the course of wound healing is present. In this case, on the basis of the risk/benefit consideration, the risk of an infection must be judged higher than the risk of cytotoxicity. With polihexanide, however, we have a wound antiseptic that is favourable with regard to both antimicrobial effect and cytotoxicity and with a correspondingly high BI, thus permitting a different evaluation of the benefit/risk ratio. Although, purely theoretically, consideration should therefore be given as to whether, in the case of use of polihexanide-containing products, preventive continuation of the antiseptic treatment would indeed be possible under certain circumstances, a definitive statement cannot be made without the existence of further experimental and clinical data. In any case, financial considerations support the discontinuation of antiseptic therapy when the infection has been controlled. The exceptions to the continuation of antiseptic treatment after the completion of therapy are patients in whom, on account of their health or sociohygienic situation, recontamination with pathogenic organisms is very likely. In this case, the antiseptic therapy passes seamlessly into preventive antiseptic treatment.

Polihexanide is also suitable for primary preventive use. At the present time, however, there are few data available [15]. Whether preventive use is indicated must always be evaluated critically in the individual benefit/risk analysis and from the therapeutic and financial aspects,

and must be repeatedly reviewed. Non-specific, unjustified and permanent use must be refused.

### Practical Application

In wound treatment, classic moist dressings and wet compresses during dressing changes are good clinical practice. In this context, depending on the indication, various polihexanide-containing preparations or dressings may be used.

Application may take place in the form of use of an antiseptic, a wound irrigation solution or gel, or also in the form of a wound covering. The example of a typical procedure for such a dressing is described in table 6 [53].

### Conclusion

Polihexanide is an antimicrobial substance that is highly suitable for use in critically colonized or infected acute and chronic wounds. Its positive evaluation is attributable particularly to its broad antimicrobial spectrum, good cell and tissue tolerability, ability to bind to the organic matrix, low risk of contact sensitization, and

wound healing promoting effect. In addition, no development of microorganism resistance during polihexanide use has been detected to date, nor does this risk appear imminent.

The aim of therapy with polihexanide is to reduce the pathogen burden in a critically colonized or infected acute or chronic wound. The use of polihexanide is not the only therapy option; priority is given to prior surgical debridement and clarification of the cause of the underlying disease, including appropriate therapy.

### Disclosure Statement

The authors declare that they have no patents or patents pending for any of the medical devices, and no stocks or stocks options related to any company manufacturing commercially available products on the basis of polihexanide as the active antimicrobial ingredient. Dr. Thomas Eberlein declares having received financial support as a clinical investigator and speaker for B. Braun, Lohmann & Rauscher, and Prontomed. Professor Ojan Assadian received speaker's honoraria for public medical education related to clinical use of antiseptics, including polihexanide, in the past from B. Braun and Lohmann & Rauscher.

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