Simulating Sequences of the Human Genome with Rare Variants

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Abstract

Objective: Simulated samples have been widely used in the development of efficient statistical methods identifying genetic variants that predispose to human genetic diseases. Although it is well known that natural selection has a strong influence on the number and diversity of rare genetic variations in human populations, existing simulation methods are limited in their ability to simulate multi-locus selection models with realistic distributions of the random fitness effects of newly arising mutants. Methods: We developed a computer program to simulate large populations of gene sequences using a forward-time simulation approach. This program is capable of simulating several multi-locus fitness schemes with arbitrary diploid single-locus selection models with random or locus-specific fitness effects. Arbitrary quantitative trait or disease models can be applied to the simulated populations from which individual- or family-based samples can be drawn and analyzed. Results: Using realistic demographic and natural selection models estimated from empirical sequence data, datasets simulated using our method differ significantly in the number and diversity of rare variants from datasets simulated using existing methods that ignore natural selection. Our program thus provides a useful tool to simulate datasets with realistic distributions of rare genetic variants for the study of genetic diseases caused by such variants.

Introduction

Increasing evidence has suggested that rare and generally deleterious genetic variants might have a strong impact on the risk of not only rare Mendelian diseases, but also many common human diseases and related traits [1, 2]. Because genome-wide association studies are inefficient in identifying rare variants that predispose to common diseases, whole-genome and whole-exome sequencing in families or individuals with extreme traits have been proposed to identify the disease-causing variants of these diseases [3].

Simulated datasets have been used to explore the role of genetic variants in human genetic diseases and to evaluate the performance of statistical methods that are designed to detect these variants [4, 5]. Under the Common Disease/Many Rare Variants hypothesis [1, 4–8], a common disease might be caused by multiple rare variants that are under relatively strong selection pressures. A realistic selection model that reflects the distribution of random fitness effects among newly arising mutations is therefore important for simulating rare variants. Unfortunately, existing coalescent-based simulation programs can only simulate neutral alleles (e.g. ms [9]) or natural selection on a single locus (e.g. SelSim [10]), and existing forward-time simulation programs only focus on a fixed number of mutations that are under selection [11], simulate sequences as independent nucleotides that are under selection [5], or use simple selection models with random fitness effects [12–14].
We developed the computer program srv (Simulator of Rare Variants) to simulate large populations of gene sequences using an evolutionary process with realistic demographic, mutation and multi-locus selection models with random fitness effects. The simulated populations can be used to study the impact of demographic and natural selection models on the number and frequencies of mutants (i.e. derived alleles), and, more importantly, generate samples for the development of statistical gene mapping methods for detecting rare variants that predispose to human genetic diseases.

Methods

Our method simulates one or more chromosomal regions that represent genes on the human genome. During a long evolutionary process, single nucleotide mutants are introduced to these genes and cause changes in the fitness of individuals who carry these mutants. The number and frequency of these mutants in the simulated population are affected by mutation, natural selection, and population demography. A gene typically spans from 10,000 to 100,000 base pairs. Although genetic recombination is usually negligible in such short regions, recombination at a constant rate (per base pair per generation) can be used to recombine parental chromosomes before they are transmitted to offspring.

We assume a multi-stage demographic model where a population of size $N_i$ at stage $t$ expands exponentially or reduces instantly to size $N_i+1$ in $G_i$ generations ($t = 0, ..., m – 1$, where $m$ is number of stages). More complex demographic models can be approximated with finer stages (e.g. exact population size at each generation). Optionally, the population can be split into several subpopulations with given proportions at a specified generation. An island model is used to migrate individuals between the resulting subpopulations.

We use two dialleleic mutation models to mutate alleles at all nucleotides at a fixed mutation rate. A finite-sites model is used by default in which all mutations such as forward, recurrent and back mutations are allowed. A mutation event can happen at a locus by relocating a mutant if it hits a locus with existing mutants. An island model is used to migrate individuals between the resulting subpopulations.

We assume a multi-allele mutation model in which all mutations are allowed. A mutation event can happen at a locus by relocating a mutant if it hits a locus with existing mutants. The dominance parameter of 0.184 and a scale parameter of 0.320 were drawn from a gamma distribution with a shape parameter of 0.184 and a scale parameter of 0.320. Alternatively, a pseudo-infinite-sites model can be used to mimic an infinite-sites mutation model by relocating a mutant if it hits a locus with existing mutants.

The fitness effect of a mutant at locus $i$ is modeled by fitness values $1/2$, $1 – h_i s_i$, and $1 – s_i$ for individuals with 0, 1 and 2 mutants at this locus, respectively. Although the selection coefficients $s_i$ are usually positive, zero or negative values can be used to simulate neutral loci or loci under positive selection. The dominance coefficient $h_i$ is frequently set to 0.5 for an additive model, or 0 for a recessive model. The selection and dominance coefficients at each locus are usually drawn from a random distribution but locus-specific coefficients can be specified using user-provided functions. This program incorporates multiple sets of selection parameters estimated from human genome data using different demographic models [5, 15–17]. For example, using a mixed gamma distribution, a mutant can have a selection coefficient of zero (neutral alleles) or a random number drawn from a gamma distribution ranging from 0.00001 to 0.1 [5].

Instead of simulating each locus separately [5], we assign an overall fitness value to an individual if he or she carries mutants at more than one locus. Either an exponential ($f = \exp \Sigma (1 – f_i)$, where $f_i$ is the fitness value at locus $i$, and $f$ is the overall fitness value), a multiplicative ($f = \Pi f_i$), or an additive ($f = \max [0, 1 – \Sigma (1 – f_i)]$) model can be used. Although these models are different from the models used in SFS_CODE where the fitness effect of multiple mutants (not genotypes) are combined either additively or multiplicatively [12], the differences between these models are small when $s_i$ is small and $h_i = 0.5$ for all loci.

Our program produces several output files, including a mutant file that contains mutants of all individuals, and a map file that contains the location, frequency, and selection and dominance coefficients of each mutant. Mutation events that happened during the evolutionary process can also be saved and used to trace the age of mutants. The simulated population can be post-processed to generate samples for particular studies. For example, a quantitative trait model can be applied to the simulated populations from which individuals with extreme trait values can be sampled. Pedigree samples can be generated by evolving the simulated population for one or more generations while keeping parental genotype information. Examples on how to generate such samples are provided in the program website.

Results

As a demonstration, we evolved initial populations of 8,100 individuals with 63,000 base pairs for 81,000 generations and expanded them to 900,000 individuals in 370 generations after a short bottleneck of 7,900 individuals. This model reflects the demographic models of the European population [5]. We used a neutral model and an exponential multi-locus selection model for nonsynonymous mutations, where fitness values of new mutants were drawn from a gamma distribution with a shape parameter of 0.184 and a scale parameter of 0.320 [16]. A finite-sites mutation model with a mutation rate of $1.8 \times 10^{-8}$ per generation per base pair was used. We drew 700 random individuals from the simulated population and obtained the number of SNPs for each mutant frequency class [a class $i$ SNP means there are $i$ copies of mutants (derived allele) on the site, $0 < i < 1,400$].

As a comparison, we also simulated sequences with the same neutral model described above using ms [9]. With 10,000 replicates, we compared the average numbers of SNPs for each mutant frequency class obtained from the three simulations. The results obtained with srv and ms under the neutral model were very similar, except the number of singletons of srv was slightly smaller than
that of ms (fig. 1). The difference may partly be due to the different models employed by the two methods. srv uses the default finite-sites model for mutations and simulates individuals in discrete time (generation), whereas ms uses an infinite-sites model for mutations and the continuous-time coalescent approximation for simulation. Compared with the neutral models, the mutants under the selection model showed dramatic reductions in the total number of mutants and a proportional concentration of extremely rare variants, as expected (fig. 1).

Although the distribution of allele frequencies of sequences with linked loci (nucleotides) is difficult to derive, a computer program (prfreq [16]) has been developed to estimate the distribution of allele frequencies of a nucleotide locus using numerical integration, effectively giving us a distribution of allele frequencies of unlinked loci under certain demographic and genetic assumptions. To validate our simulation method against theoretical estimates, we simulated a constant-size population of 8,100 individuals with 10,000 base pairs for 81,000 generations, using an infinite-sites mutation model with a mutation rate of $1.8 \times 10^{-8}$ per generation per base pair. We used a multiplicative multi-locus selection model for non-synonymous mutations, with fitness values of new mutants drawn from a gamma distribution with a shape parameter of 0.184 and a scale parameter of 0.320 [16].

We drew 10 random individuals from the simulated population and obtained the number of SNPs for each mutant frequency class. We compared the average number of each SNP class to the expectations obtained from the prfreq program with the same demographic model and selection coefficient distribution (fig. 2). Although prfreq obtains the expectations assuming independent nucleotide sites in contrast to our assumption of a non-recombined DNA strand, the means of the simulation results fit reasonably well to prfreq’s expectations, which suggests that the marginal distribution of each nucleotide in our simulation is similar to that of an independent site.
Discussion

Due to the stochastic nature of the evolutionary process, the number, location, and frequency of mutants vary from population to population. A random realization of this evolutionary process requires a long evolution time (for example a long period of constant population size). If multiple replicates of the same evolutionary scenario are simulated, a population that is under mutation and selection equilibrium can be simulated in advance and used as the founder population for replicate simulations. Similarities between populations simulated in this manner (e.g. sharing of mutants from the founder population) can be controlled by evolving the founder populations for additional generations.

In order to simulate samples that resemble real-world genetic data, our program provides an option to import alleles from an existing sample (e.g. the HapMap dataset [18]) during evolution. These alleles are usually introduced before rapid population expansion. Because common alleles will most likely remain common during rapid population expansion [19], manually inserted common variants will remain common in the simulated population, but at the same time blend nicely with new mutants that are introduced during the population expansion stage. Such datasets provide an ideal tool to study the power of genome-wide association studies of diseases that are caused by rare and unobserved variants [4].

srv allows the use of a pseudo-infinite-sites mutation model in which mutations only happen at nucleotide loci without existing mutants. Whereas a mutant will be considered new in a coalescent-based simulation if it does not appear in individuals in the coalescent tree, our model requires that no mutant at the same locus exists in the whole population. Because large populations of short sequences may become saturated so that every locus has existing mutants, an infinite-sites model should not be used if the number of segregation sites in the simulated population is close to the length of sequences.

If the underlying evolutionary process can be sufficiently approximated by a Wright-Fisher model, a scaling technique can be used to speed up a forward-time simulation [20, 21]. Compared to a regular simulation that evolves a population of size $N$ for $t$ generations, a scaled simulation with a scaling factor $\lambda$ evolves a smaller population of size $N/\lambda$ for $t/\lambda$ generations with magnified (multiplied by $\lambda$) mutation, recombination, and selection forces. However, because this scaling technique might not be applicable to all supported selection models, and will result in a final population of size $N/\lambda$ instead of $N$, this technique should be used with caution.

A number of computer programs are available to simulate sequences for genetic epidemiological studies. Table 1 lists some of the popular coalescent-based programs and forward-time simulation programs that support natural selection on multiple loci. More complete surveys of such software are provided by Liu et al. [22] and Carvajal-Rodriguez [23]. srv differs from these programs in its ability to simulate diploid and site-specific selection models, to use an infinite-sites in addition to a finite-sites mutation model, and to introduce alleles from empirical data.

<table>
<thead>
<tr>
<th>Program</th>
<th>Method</th>
<th>Selection</th>
<th>Recombination</th>
<th>Mutation models</th>
<th>Disease model</th>
<th>Ref.</th>
</tr>
</thead>
<tbody>
<tr>
<td>ms</td>
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<td>no</td>
<td>uniform</td>
<td>infinite sites</td>
<td>no</td>
<td>9</td>
</tr>
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<td>GENOME</td>
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<td>infinite sites</td>
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<tr>
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<tr>
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<td>diallelic</td>
<td>yes</td>
<td>26</td>
</tr>
<tr>
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<td>no</td>
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</tr>
<tr>
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<tr>
<td>SFS_CODE</td>
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<td>varying</td>
<td>finite sites, diallelic</td>
<td>no</td>
<td>12</td>
</tr>
</tbody>
</table>

1 User can define multiple consecutive fragments of the sequence, the recombination only occurs between fragments but not within the fragments.
2 Multiplicative multi-locus model with selection coefficients drawn from a gamma distribution.
3 Additive multi-locus selection model with selection and dominance coefficients drawn from a mixture of two Gaussian distributions.
4 Additive or multiplicative multi-mutant selection model with scaled selection coefficients drawn from a random distribution.
Perhaps the most distinguishing feature of srv is that it is implemented using a general-purpose forward-time population genetics simulation environment (simuPOP [24]) and can take full advantage of the processing power of this software. For example, a single python script can be used to simulate multiple populations using srv, apply different disease or quantitative trait models, draw population- or pedigree-based samples, and use different statistical methods to analyze them. Because of the scripting language design, it is relatively easy to modify srv to reveal details of the evolutionary process or use alternative demographic or genetic models. srv can be executed from a graphical user interface, a command line in batch mode, or called as a function from another script. It takes around 10 min to simulate 90,000 sequences of 63,000 base pairs on a reasonably configured PC using a demographic model with a long constant population size followed by rapid population expansion, and an exponential multi-locus selection model with selection coefficients drawn from a gamma distribution. srv is freely available at the 'Complete Script’s section of the simuPOP online cookbook at http://simupop.sourceforge.net/cookbook. Several examples are provided to demonstrate features of this program.

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References


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