Nonsteroidal Anti-Inflammatory Drugs Alter the Human Mesothelial Pleural Permeability via Ion Cellular Transportation by Inhibiting Prostaglandin Synthesis

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\textbf{Key Words}
Human \cdot Ion transportation \cdot Mesothelial \cdot Nonsteroidal anti-inflammatory drugs \cdot Permeability \cdot Pleura \cdot Prostaglandin

\textbf{Abstract}
\textbf{Background:} Nonsteroidal anti-inflammatory drugs (NSAIDs) are used in clinical practice as analgesics or anti-inflammatory drugs. Studies have implicated them in participating in permeability throughout various tissues such as the kidneys and lungs. \textbf{Objective:} The effect of NSAIDs on the pleural permeability and the underlying mechanisms whereby this effect is mediated were investigated. \textbf{Methods:} Parietal pleural specimens were obtained from patients subjected to thoracic surgery and were mounted in Ussing chambers. Solutions containing paracetamol, acetylsalicylic acid, diclofenac, lornoxicam, parecoxib and ibuprofen were added in the chambers facing the pleural and the outer-pleural surface. Prostaglandin E$_2$ was similarly used to investigate prostaglandin synthesis involvement at low and high doses. Amiloride- and ouabain-pretreated specimens were used in order to investigate ion transportation involvement. Trans-mesothelial resistance ($R_{TM}$) was determined as a permeability indicator. \textbf{Results:} Paracetamol, acetylsalicylic acid, diclofenac, lornoxicam and ibuprofen increased $R_{TM}$ on the pleural and outer-pleural surface, inhibited by amiloride and ouabain. Parecoxib had no effect on the $R_{TM}$. Prostaglandin decreased $R_{TM}$ on the pleural and outer-pleural surface inhibited by amiloride, ouabain and ibuprofen. \textbf{Conclusion:} NSAIDs, except parecoxib, induce a rapid decrease of the pleural permeability by inhibiting cellular transportation, an effect that is mediated by prostaglandin synthesis inhibition.

\textbf{Introduction}

The nonsteroidal anti-inflammatory drugs (NSAIDs), apart from anti-inflammatory and analgesic properties, have been implicated in causing permeability alterations in various tissues such as the kidneys and lungs, interfering with ion transportation across epithelial tissues [1, 2]. In most cases these alterations are being mediated via interference with prostaglandin production [1–3].
In pleura, the anti-inflammatory drugs, and more specifically NSAIDs, are thought to play an important role in exudate progression [4] while they are shown to interfere with the pleurodesis quality [5, 6]. Furthermore, histamine, an inflammatory hormone, was shown to cause permeability alterations in human parietal pleura mainly by interacting with the H1 histaminic receptors and eventually interfering with the normal process of transmucosal ion transportation [7].

The aim of the present study is to investigate the effect of NSAIDs on the electrophysiology of the human parietal pleura. Additionally, we investigate the mechanisms whereby these drugs mediate their effect such as the inhibition of prostaglandin synthesis and the involvement of ion cellular transporters.

**Materials and Methods**

**Study Subjects**

Intact sheets of human parietal pleura were obtained from 12 patients who underwent thoracic surgery for lung cancer (via thoracotomy or thoracoscopy). The pleural specimen was obtained from an area distant to the lung tumor. A piece from each dissected specimen was sent for histopathological examination. All specimens included in the study were proven to be free of any disease as per histopathological reports. Patients with pleural effusion preoperatively were excluded from the study. After its dissection, each remaining specimen was placed in Krebs solution, pre-oxygenated at 95% O2-5% CO2 and cooled at 4 °C, and transferred to the laboratory within 30 min.

The study was approved by the Local Ethics Committee and informed signed consent was obtained from all patients participating in the study.

**Study Design**

Experiments were conducted with different NSAID solutions added towards the pleural and outer-pleural surfaces of stripped pleural specimens. Amiloride- and ouabain-pretreated specimens were used to investigate ion transportation involvement in the NSAID effect on pleura. Experiments with prostaglandin E2 were also performed with amiloride, ouabain and ibuprofen in order to investigate prostaglandin synthesis involvement in this effect.

**Methods**

The Krebs solution used throughout the whole study was balanced at pH 7.45 and contained 117.5 mM NaCl, 1.15 mM NaH2PO4, 24.99 mM NaHCO3, 5.65 mM KCl, 1.18 mM MgSO4, 2.52 mM CaCl2, and 5.55 mM glucose.

The pleural surface (which faces the pleural cavity in vivo) will be referred to as the mesothelial surface, whereas the outer-pleural surface (which faces the chest wall) will be referred to as the interstitial surface.

The pleural specimens were mounted as planar sheets of tissue in Ussing-type chambers (model DVC-1000; World Precision Instruments, Sarasota, Fla., USA), were bathed in Krebs solution on both sides and perfused continuously with 95% O2-5% CO2 gas mixture heated to 37°C in order to ensure tissue viability [8–10].

Following mounting, the pleural specimens were equilibrated for 30 min. Transmucosal potential difference (PD) was then measured for 30 min, with or without the application of a current of variable intensity (0 ± 400 μA) [8–10], constituting the control potential difference. Fifty control experiments were performed.

Following the equilibration period and control measurements, solutions containing different NSAIDs (paracetamol 1 g, acetylsalicylic acid 325 mg, ibuprofen 300 mg, lornoxicam 8 mg, diclofenac 75 mg or parecoxib 100 mg) were added sequentially on both surfaces in order to investigate their effects on pleural tissue (n = 7 experiments for each drug, n = 7 experiments for each side). In other experiments, in order to investigate whether prostaglandins interfere in permeability alterations caused by NSAIDs, prostaglandin E2 (Sigma Chemical Co., St. Louis, Mo., USA) was used in the same manner at one low concentration (10–6 M) and one high concentration (10–4 M) (n = 7 experiments for each concentration, n = 7 experiments for each side). In another group of experiments, specimens were pretreated with the Na+ channel blocker amiloride (10 μM; Sigma Chemical Co.) [9, 10] and the Na+/K+ pump inhibitor ouabain (10 μM; Sigma Chemical Co.) [9, 10] for at least 30 min before the addition of each NSAID solution, in order to clarify whether ion cellular transporters take part in their effect on human pleura (n = 7 experiments for each drug, n = 7 for each surface). Experiments with tissues pretreated with amiloride and ouabain before the addition of prostaglandins at high doses (based on observations made before) were also used in order to clarify the involvement of cellular transporters in prostaglandin effect (n = 7 experiments for each drug, n = 7 for each surface). Finally, given that ibuprofen is widely used as a prostaglandin inhibitor [11], experiments with the coaddition of ibuprofen and prostaglandin were also conducted (n = 7 experiments for each drug, n = 7 for each surface). In total, 126 experiments were conducted.

In order to ensure that the recorded results were due to drug action and not due to mechanical perturbation while emptying and refilling the chambers, the experiments were conducted using solely Krebs solution (data not shown as no electrical changes in PD were observed). The PD following electrical stimulation with the application of a current (0 ± 400 μA) was measured 1, 5, 10, 30 and 60 min following the addition of each solution. All solutions were freshly prepared before every experiment, heated to 37°C and continuously perfused with 95% O2-5% CO2 gas mixture.

R74 was calculated from the PD according to Ohm’s law [8–10].

**Analysis**

Statistical analysis was performed using the statistical package SPSS ver. 10.00 for Windows (Statistical Package for the Social Sciences; SPSS Inc., Chicago, Ill., USA). Data are expressed as mean R74 (Ω cm2) ± standard error of mean (SE) within the first minute. Statistical significance was determined by paired t test. Comparison among groups was performed by one-way ANOVA (Bonferroni’s post hoc test). p values <0.05 were considered significant.
Results

Effect of Anti-Inflammatory Agents on the Electrophysiology of the Human Parietal Pleura

The addition of paracetamol significantly increased $R_{TM}$ on both the mesothelial ($p = 0.032$, fig. 1a) and the interstitial surfaces ($p = 0.03$, fig. 1b) of the pleura within the first minute after addition. Amiloride inhibited this effect on both the mesothelial ($p = 0.932$, fig. 2a) and interstitial surfaces ($p = 0.993$, fig. 2b). Ouabain also inhibited paracetamol’s effect on the mesothelial ($p = 0.892$, fig. 3a) and interstitial surfaces ($p = 0.923$, fig. 3b).

The addition of acetylsalicylic acid acutely and significantly increased $R_{TM}$ on both the mesothelial ($p = 0.012$, fig. 1a) and interstitial surfaces ($p = 0.058$, fig. 1b) of the pleura. Amiloride inhibited this effect on both the meso-
theleial (p = 0.934, fig. 2a) and interstitial surfaces (p = 0.993, fig. 2b). Ouabain also inhibited the aforementioned effect on the mesothelial (p = 0.762, fig. 3a) and interstitial (p = 0.954, fig. 3b) surfaces.

Diclofenac also induced a rapid and significant increase of $R_{TM}$ when added on both the mesothelial (p = 0.012, fig. 1a) and interstitial surfaces (p = 0.008, fig. 1b) of the pleural specimens. Again amiloride inhibited this effect on both the mesothelial (p = 0.863, fig. 2a) and interstitial surfaces (p = 0.883, fig. 2b) and ouabain had a similar effect on the mesothelial (p = 0.892, fig. 3a) and interstitial surfaces (p = 0.658, fig. 3b).

A similar effect was induced by lornoxicam within the first minute after addition, i.e. $R_{TM}$ significantly increased on both the mesothelial (p = 0.028, fig. 1a) and interstitial surfaces (p = 0.033, fig. 1b). Amiloride inhib-
ited this effect on both the mesothelial (p = 0.928, fig. 2a) and interstitial surfaces (p = 0.228, fig. 2b). Oubain also inhibited lornoxicam’s effect on both the mesothelial (p = 0.438, fig. 3a) and interstitial (p = 0.588, fig. 3b) surfaces.

Parecoxib had a weak effect on the $R_{TM}$ of the pleural specimens when added on the mesothelial (p = 0.078, fig. 1a) and interstitial (p = 0.248, fig. 1b) surfaces, which were not totally inhibited by amiloride (p = 0.08, fig. 2a for the mesothelial and p = 0.198, fig. 2b for the interstitial surface) and ouabain (p = 0.09, fig. 3a for the mesothelial and p = 0.128, fig. 3b for the interstitial surface).

The addition of ibuprofen significantly increased $R_{TM}$ on both the mesothelial (p = 0.001, fig. 1a) and interstitial (p = 0.004, fig. 1b) surfaces of the pleura within the first minute after addition. Amiloride inhibited this effect on the mesothelial (p = 0.932, fig. 2a) and interstitial surfaces (p = 0.893, fig. 2b). Oubain also inhibited ibuprofen’s effect on the mesothelial (p = 0.942, fig. 3a) and interstitial surfaces (p = 0.923, fig. 3b).

**Comparison of the Effects Induced by Inflammatory Drugs on the Electrophysiology of the Human Parietal Pleura**

The comparison of the effect induced by the above-investigated drugs revealed no difference among them except the comparison of the parecoxib effect on the mesothelial surface which was significantly different from the effect induced by paracetamol (p = 0.048) and ibuprofen (p = 0.042), and on the interstitial surface when compared to the effect produced by paracetamol (p = 0.028) and ibuprofen (p < 0.01). Ibuprofen induced a higher effect on the interstitial surface in comparison with the other NSAIDs (vs. paracetamol (p = 0.032, acetylsalicylic acid p = 0.018, lornoxicam p = 0.018 and parecoxib p < 0.01).

The comparison of the effect induced by all drugs on the mesothelial with the effect induced on the interstitial surface revealed no difference.

**Effect of Prostaglandins on the Electrophysiology of the Human Parietal Pleura**

Prostaglandin at high concentration induced a rapid decrease of the $R_{TM}$ within the first minute after addition when added on the mesothelial surface (p = 0.021, fig. 4a). On the interstitial surface the decrease had the tendency to be statistically different (p = 0.059, fig. 4b).

Amiloride and ouabain also inhibited the effect of prostaglandins on the electrophysiology of the human parietal pleura on both surfaces (p = 0.886, fig. 4a for mesothelial and p = 0.799, fig. 4b for interstitial surface for amiloride, and p = 0.936, fig. 4a for mesothelial and p = 0.249, fig. 4b for interstitial surface for ouabain).

Prostaglandin, when added with ibuprofen, had no effect on the electrophysiology of the human parietal pleura on both surfaces (p = 0.971, fig. 4a for mesothelial and p = 0.987, fig. 4b for interstitial surface).

**Discussion**

The main finding of the present study is that NSAIDs, except parecoxib, induce a rapid effect on the electrophysiology of the human parietal pleura by increasing the $R_{TM}$ and consequently decreasing the overall permeability. This effect is mediated via the inhibition of the Na$^+$ channels and Na$^+/K^+$ pumps normally operating in the pleura, since it is blocked by amiloride and ouabain. Prostaglandin synthesis seems to be an intermediate event between the interaction of NSAIDs with the pleura and the inhibition of ion transportation, as prostaglandin $E_2$ induced an opposite effect to the NSAIDs effect by rapidly decreasing the $R_{TM}$ and therefore increasing the pleural permeability, an effect again inhibited by the ion transporter blockers amiloride and ouabain.

NSAIDs have been implicated in altering the endothelial permeability and the cellular transportation via ion channels in various tissues [12] such as the kidneys, where urinary sodium excretion was decreased and sodium absorption was increased by ibuprofen [1]. Indomethacin decreased the permeability increase induced by lung adenocarcinoma cells [2]. Ibuprofen also inhibited the current when basolaterally applied in Xenopus kidney A6 cell lines [12] while it suppressed the upregulation of Na$^+$ channels in the small dorsal root ganglion [13], an effect also observed with celecoxib [14]. The inhibition of various ion channels by NSAIDs, such as celecoxib, has been demonstrated in retinal neurons as well [15]. Ion cellular transporters, such as amiloride-sensitive Na$^+$ channels, ouabain-sensitive Na$^+/K^+$ pumps, glibenclamide-sensitive K$^+$ channels and others were previously shown to be present in the pleura, producing a pleural fluid recycling potential operating under normal conditions [8–10]. This study suggests that these ion transporters operating in the pleura are inhibited by NSAIDs, decreasing its permeability and therefore hindering part of the pleural fluid recycling process.

It has been debated whether specific cyclooxygenase-2 (COX-2) inhibitors induce less and weaker side effects than the other nonselective NSAIDs [16]. These selective

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inhibitors were considered to offer a better inflammatory pleural response in pleurodesis [17], thereby overcoming the decreased quality of pleural adhesions during pleurodesis induced by the nonselective NSAIDs [1, 6, 18]. Celecoxib, on the other hand, is considered to inhibit Na+ channels in dorsal root ganglion neurons [14] or in rat retinal neurons [15]. COX-2 expression stimulated by prostaglandins was not investigated. Another limitation of the study is that it was performed on normal tissues and not on inflammatory tissues which express different types of cyclooxygenases (COX-2 more specifically), an event that could partially explain the fact that parecoxib did not alter pleural permeability.

In conclusion, NSAIDs induce a rapid decrease of the pleural permeability by inhibiting cellular transportation, an effect that is mediated by prostaglandin synthesis inhibition. Parecoxib was the only NSAID that induced a nonsubstantial effect. Thoracic surgeons should consider this information when managing postoperative pain.

References


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