A Prime Time for Trained Immunity: Innate Immune Memory in Newborns and Infants

Ofer Levy, James L. Wynn

Division of Infectious Diseases, Department of Medicine, Boston Children’s Hospital, and Harvard Medical School, Boston, Mass., and Division of Neonatology, Department of Pediatrics, Vanderbilt University, Nashville, Tenn., USA

Key Words
Newborn · Infant · Trained immunity · Innate immunity

Abstract
The newborn and infant periods of early life are associated with heightened vulnerability to infection. Limited antigen exposure and distinct adaptive immune function compared to the adult places a greater burden on innate immunity for host defense to microbial challenge during this time. Trained immunity describes the phenomenon of augmented innate immune function following a stimulus that is not specific to the original stimulus. We review the concept of trained immunity in the context of the newborn’s unique innate immune system function, the preclinical and clinical evidence that supports the tenet of innate immune memory in early life, and potential consequences of altered innate immune host responses.

Introduction
Unlike the highly specific and long-lived memory responses associated with an effective adaptive immune system, the mammalian innate immune system has classically been described to manifest a rapid, short-lived ‘nonspecific’ response to foreign antigens. The concept whereby a prior exposure to an immune stimulus might result in augmentation of innate immune function upon subsequent exposure to the same or a different stimulus was described in humans nearly 50 years ago [1]. Netea et al. [2] recently coined the term ‘trained immunity’ to specifically describe enhancement of innate immune function with reinfection; the development of innate immune memory. A trained innate immune response to a subsequent challenge has likely existed for millions of years in plants and invertebrates, both of which lack a classic adaptive immune system [3, 4].

In plants, the process by which protection against reinfection is mediated is termed systemic acquired resistance. Epigenetic reprogramming through specific histone acetylation (H3K9) is vital for this effect [5]. Because adaptive immunity is largely restricted to vertebrates, invertebrates represent excellent sources to look for the presence or absence of trained immunity [6]. Midgut barrier disruption by Plasmodium in Anopheles gambiae resulted in differentiation of hemocytes to an abundance of granulocytes associated with enhanced bacterial immunity that resulted in protection against reinfection with Plasmodium [3]. The mealworm beetle exhibited enhanced protection against antigenically unrelated secondary infection following lipopolysaccharide or bacterial priming [7]. These are but a few examples of many...
investigations in organisms from plants to corals to shrimp to water fleas that demonstrate that trained innate immunity is prevalent. Recently, in-depth studies have also been performed in adult mammals aimed at uncovering the mechanisms that result in trained immunity among innate immune cellular populations including NK cells, monocytes, macrophages, DCs, and microglia. Discussions of each study’s findings are beyond the scope of this mini review but the reader is directed to several recent outstanding reviews [2, 8, 9].

A more robust innate immune response on reinfection with the same pathogen may provide enhanced protection or have heterologous beneficial effects by providing protection against an unrelated pathogen. Trained immunity phenomena are demonstrable in both preclinical neonatal disease models and in human neonates (<28 days of life). Indeed, the development of trained immunity may be particularly important for host survival in early life and could potentially affect the risks of infection, allergy, and chronic inflammatory diseases later in life. In this mini review, we will review key functional distinctions of the newborn innate immune system, present evidence for early-life trained immunity in neonatal preclinical disease models and in neonatal humans, and discuss how trained immunity might be associated with negative effects on the host.

**Distinct Early-Life Innate Immune Function**

Early life represents a period of dramatic stimulation of the relatively naïve newborn immune system. Exposure and colonization with commensal organisms harboring trillions of nucleic acid, carbohydrate, and protein antigens occurs shortly after birth. Sentinel cells of the innate immune system represent the first responders to this massive immune system exposure. Innate immune responses in turn stimulate the adaptive immune system and the development of classic immune memory responses. Thus, the response of the innate immune system is critical for the initiation and maintenance of host defense through effective immune surveillance, successful discrimination between pathogens and commensals, and development of immunologic memory.

Multiple lines of evidence now support that newborn and early-life innate immune function is not simply immature, but is distinct from that seen in older more mature populations well into the first year of life [10–12]. Furthermore, the newborn has a significant dependence on innate immune function due to distinct adaptive immune capabilities in early life [13]. Once innate immune sentinel cells, including tissue macrophages, is the identification of invading pathogens. Pathogen-associated molecular patterns are sensed via several pattern-recognition receptors including the Toll-like receptors (TLRs), nucleotide oligomerization domain-like receptors, retinoic acid-inducible protein I-like receptors, integrins and C-type lectins.

TLRs are key elements in the innate immune system’s ability to recognize and respond to pathogens and are critically important for early-life host immune responses [14]. Present on multiple cell types, TLRs recognize extracellular and intracellular pathogens by their respective microbial products. TLR agonist-receptor-binding results in downstream production of cytokines and chemokines as well as antimicrobial effector mechanisms [15]. There are 10 known TLRs in humans, 12 in mice, and each receptor has a specific molecular activation trigger [15–17]. Microorganisms may stimulate multiple TLRs simultaneously akin to a ‘molecular piano’ playing ‘chords’ ultimately signaling the presence of particular types of pathogens [16, 18]. Following PRR stimulation, production of cytokines and chemokines results in amplification of the innate response directed at the invading organisms.

Although multiple factors contribute to the altered innate immune response profile of the newborn as compared to the adult, key developmental age-related differences in TLR-mediated cytokine production as compared to adults have been recently reviewed [14]. Basal expression and cellular distribution of TLRs on term newborn monocytes is broadly similar to that of adult monocytes [19]. In contrast, TLR4 expression of preterm monocytes is reduced and increases with gestational age [20]. Interestingly, post-natal monocyte TLR expression increases early in life [21]. Despite similar basal TLR expression, the functional consequences of TLR engagement in neonates are distinct. For example, preterm infant mononuclear cells demonstrate robust interleukin (IL)-10 production but diminished production of pro-inflammatory cytokines [14]. In contrast, mononuclear cells of term infants demonstrate high levels of IL-6 and IL-23 production supporting Th17 differentiation [14]. Diminished production of TNF-α, IFN-γ, IL-1β and IL-12p70 relative to adults is present for several weeks after birth and likely contribute to increased susceptibility to intracellular infection [14, 19, 20, 22, 23]. The decreased pro-inflammatory cytokine production is due in part to decreased...
production of important intracellular mediators of TLR signaling including myeloid differentiation factor 88 (MyD88), interferon regulatory factor 5, and p38, which exhibit gestational age-specific diminution [20]. Recently, neonatal CD71+ erythroid precursor cells (nucleated red blood cells) have been implicated as mediating arginase-dependent suppression of DC TNF-α production, impaired host resistance to Listeria monocytogenes, and inhibition of postnatal intestinal leukocyte TNF production [24]. Specifically, soluble newborn cord blood plasma factors, including high concentrations of adenosine, reduce monocytic production of TNF-α with preservation of IL-6 synthesis [25–28]. As IL-6 has pro-resolution properties, including inhibition of neutrophil migration [29], this polarization may serve to reduce the risk of excessive pro-inflammatory/Th1 response during the initial colonization of the skin and intestinal tract. Other potential teleologic explanations for the pattern of cytokine production in the newborn period include (1) the prevention preterm birth secondary to in utero inflammatory responses, (2) a reduction in the likelihood of fetal rejection by the mother, and (3) developing fetal immunologic tolerance [30].

**Laboratory and Preclinical Evidence of Trained Immunity in Neonates**

Our laboratories and others have reported on the effects of innate immune priming on subsequent cellular function in both ex vivo human and preclinical animal models of disease. For example, TLR4 is upregulated on monocytes and neutrophils from term neonates after labor and neutrophil migration is enhanced after exposure to a TLR4 agonist [31–33]. IL-8 priming of neutrophils that occurs during labor significantly improves neutrophil chemotaxis over that seen with Caesarian delivery and even adult controls [34]. Zhang et al. [35] showed newborns amplify the TLR2/MyD88 pathway in Gram-positive bacterial infection and the TLR4/MD2/MyD88 pathway in Gram-negative bacterial infection, suggesting infection-specific changes in innate immune signaling. Rather than demonstrate a tolerance phenomenon, these examples suggest the potential for enhanced innate immune function in human neonates following a stimulus.

Neonatal mice pretreated with low-dose specific TLR agonists such as lipopolysaccharide (TLR4) or the imidazoquinoline R-848 (TLR7/8) demonstrated a significant survival advantage over saline pretreated animals when later (24 h after pretreatment) challenged with polymicrobial sepsis [36]. TLR-mediated immune priming induced multiple enhancements in subsequent innate immune function such as altered cytokine production, reactive oxygen species production, neutrophil phagocytosis, and improved bacterial clearance; all associated with improved survival. Importantly, the TLR-mediated survival enhancement was independent of the adaptive immune system (recombinase-activating gene 1−/−) and supports the premise of trained immunity in the neonate. Similar survival enhancements were seen in TLR-primed murine neonates with subsequent Listeria or neurotropic Tacaribe arenavirus challenge [37, 38]. Cord blood monocytes harvested from fetal lambs exposed to a single dose of intra-amniotic endotoxin 7 and 14 days prior to delivery demonstrated augmented innate responses (IL-6 and hydrogen peroxide) to subsequent in vitro endotoxin stimulation [39, 40].

**Evidence Supporting Trained Immunity in Human Neonates following Early-Life Exposures**

An early-life immune stimulus may transform the host defense status of the preterm infant from a relative state of tolerance, as required to prevent maternal rejection, to a state of immunocompetence to provide effective defense against the many microbes in the extraterine environment. In line with this hypothesis, Strunk et al. [41] demonstrated that histologic chorioamnionitis exposure (inflammation of the placental chorionic disk and the extraplacental membranes) reduced the risk of late-onset sepsis (LOS) in preterm neonates. Analyses of whole blood genome-wide expression profiling revealed >2-fold upregulation of C5aR, C-LEC7A/12A, IL8RA/B, TLR4, TREM1, SIRPB1 and TNFAI6 in those with histologic chorioamnionitis exposure but without the development of early-life infection as compared to control preterm infants without histologic chorioamnionitis exposure/early-life infection [42]. Retrospective analyses of two large independent cohorts of very low birth weight (<1,500 g at birth) infants (n = 136,713) showed that early (≤3 days after birth) blood culture-positive sepsis in preterm infants was not associated with an increased risk of subsequent infection during the hospitalization (late sepsis) [43] and was associated with a reduced risk of late sepsis in the smallest most immature infants [44].

Additional precedents for potentially beneficial trained immunity exist in neonatal humans in the form of heterologous vaccination benefits that may lead to a reduction in subsequent infection-related mortality [45, 46].
birth weight neonates given Mycobacterium bovis bacillus Calmette-Guérin vaccination (has TLR2/4/8/9 agonist activity [47]) at birth experienced a heterologous (so-called ‘nonspecific’) reduction in neonatal mortality over neonates who did not receive bacillus Calmette-Guérin [48, 49]. In line with the hygiene hypothesis based on epidemiologic studies of those exposed to microbes and microbial products early in life, the risk of asthma/atopy may be reduced with early-life immune stimulation via infection [50] or breast feeding [51]. Much remains to be learned regarding both the mechanisms and ontogeny of trained immunity in humans, including whether this phenomenon demonstrates distinct features in preterm neonates.

**Potential Negative Effects of Trained Immunity**

There may also be potential negative effects of early-life trained immunity. Preterm neonates are at high risk of inflammatory sequelae of prematurity including retinopathy, chronic lung disease, and white matter injury [52]. There are clear relationships between chorioamnionitis exposure and a variety of untoward neonatal morbidities including chronic lung disease, cystic periventricular leukomalacia, intraventricular hemorrhage and cerebral palsy [53–55]. Early immune exposures might result in enhanced innate immune responses that contribute negatively to these and other sequelae of preterm birth [56].

Though beyond the scope of this review, preterm and/or low birth weight infants demonstrate an increased risk for adult cardiovascular and renal disease [57], raising the possibility that trained immunity may potentially contribute to the risks of developing chronic inflammatory conditions in later life. Others have suggested that neonatal exposure may eventually lead to development of disease as an adult [58], including Barker’s fetal origins of adult disease [59]. Early-life innate immune system exposures such as severe infection or enhanced innate immune responses in young adulthood following innate immune training with low-grade chronic inflammatory conditions such as gingivitis or bacterial vaginosis could potentially contribute to the greater risk of preterm labor, stroke, diabetes, or myocardial infarction associated with these conditions. This phenomenon, if relevant to preterm birth, would shed light on why treatment of these conditions has not modified preterm birth risk [60, 61], may partially explain racial disparities in preterm birth risk [62], and may lead to the development and testing of immunomodulation strategies. Indeed, recent global surveys suggest a worldwide incidence of preterm birth of ~11% [63]. In a recent report, it was stated ‘preterm births will remain a major public health issue, from which no country in the world is immune’ [64]. Perhaps what is needed is a retraining of innate immunity.

**Conclusion**

Trained immunity is apparent in the newborn and may be an important and necessary process to protect the vulnerable newborn while adaptive responses are limited. At this early stage of our understanding of trained immunity in newborns, there are far more questions than answers. What are the limits of the triggers and duration of the innate immune augmentation effect? Are the effects permanent? Are the modifications transmitted to offspring as they can be in plants? How are innate immune modifications mediated? Do posttranslational epigenetic changes such as histone protein modification including methylation, acetylation, phosphorylation, ubiquitination, and sumoylation play a role? Much remains to be learned about the immunology behind the transition from intrauterine to extrauterine life, the factors that modify this transition, and the duration and clinical consequences of the modifications that occur.

**Acknowledgements**

O.L.’s laboratory is supported by Global Health (OPPGH5284) and Grand Challenges Explorations (OPP1035192) awards from the Bill & Melinda Gates Foundation and by NIH grant 1R01AI100135-01. J.L.W.’s laboratory is supported by awards from the Thrasher Research Fund, the Gerber Foundation, Vanderbilt Department of Pediatrics (Turner-Hazinski Research Award), and NIH grant K08GM106143.

**References**

19 Levy O, Zarember KA, Roy RM, Cywes C, Go- 

5 Slaughter A, Daniel X, Flors V, Luna E, Hohn 

20 Sadeghi K, Berger A, Langmartner M, Prus- 

8 Perry VH, Nicoll JA, Holmes C: Microglia in 

10 persist throughout the first month of life. 

11 Belderbos ME, van Bleek GM, Levy O, Blanko 

21 Shen CM, Lin SC, Niou DM, Kou YR: Develop-

7 Moret Y, Siva-Jothy MT: Adaptive innate im-

12 Kollmann TR, Crabtree J, Rein-Weston A, 

13 Wynn JL, Levy O: Role of innate host defens-

15 Kumagai Y, Takeuchi O, Akira S: Pathogen 

9 Benn CS, Netea MG, Selin LK, Aaby P: A 

22 Hotchkiss RS, Karl IE: The pathophysiology 

10 Strunk T, Currie A, Richmond P, Simmer K, 

14 Kollmann TR, Levy O, Montgomery RR, 

17 Kawai T, Akira S: Toll-like receptors and their 

18 Krumbiegel D, Zepp F, Meyer CU: Combined 

25 Belderbos ME, Levy O, Meyaard L, Bont L: 

26 Belderbos ME, Levy O, Stalpers F, Kimpen JL, 

16 Trinchieri G, Sher A: Cooperation of Toll-like 

17 Wynn JL, Levy O: Role of innate host defens-

19 Levy O, Zarember KA, Roy RM, Cywes C, Go-

186–192.

33 Shen CM, Lin SC, Niou DM, Kou YR: Labour 

34 Yektaei-Karin E, Moskhgah A, Lundahl J, 

35 Zhang JP, Yang Y, Levy O, Chen C: Human 

36 Wynn JL, Scumpia PO, Winfield RD, Delano 

37 Pedras-Vasconcelos JA, Goucher D, Puig M, 

38 Pedras-Vasconcelos JA, Goucher D, Puig M, 

39 Kramer BW, Ikemagi M, Moss TJ, Nitos T, 

40 Kramer BW, Joshi SN, Moss TJ, Newnham JP, 

41 Strunk T, Doherty D, Jacques A, Simmer K, 

42 Pedras-Vasconcelos JA, Goucher D, Puig M, 

43 Lin CB, Hornik CP, Clark R, Cotten CM, Ben-

44 Wynn JL, Hansen NI, Das A, Cotten CM, 

45 Lin CB, Hornik CP, Clark R, Cotten CM, Ben-

46 Wynn JL, Lopez MC, Ungaro R, Baker HV, 

47 Wynn JL, Lopez MC, Ungaro R, Baker HV, 

48 Wynn JL, Lopez MC, Ungaro R, Baker HV, 

49 Kramer BW, Ikemagi M, Moss TJ, Nitos T, 

50 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

51 Kramer BW, Ikemagi M, Moss TJ, Nitos T, 

52 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

53 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

54 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

55 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

56 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

57 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

58 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

59 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

60 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

61 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

62 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

63 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

64 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

65 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

66 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

67 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

68 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

69 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

70 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

71 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

72 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

73 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

74 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

75 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

76 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

77 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

78 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

79 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

80 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

81 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

82 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

83 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

84 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

85 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

86 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

87 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

88 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

89 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

90 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

91 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

92 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

93 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

94 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

95 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

96 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

97 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

98 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

99 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

100 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

101 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

102 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

103 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

104 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

105 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

106 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

107 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

108 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

109 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

110 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

111 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

112 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

113 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

114 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

115 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

116 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

117 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

118 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

119 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

120 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

121 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

122 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

123 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

124 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

125 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

126 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

127 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

128 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

129 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

130 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

131 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

132 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

133 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

134 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

135 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

136 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

137 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

138 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

139 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

140 Kramer BW, Ikemagi M, Moss TJ, Newnham JP, 

Levy/Wynn


