Progress and Prospects of Long Noncoding RNAs (IncRNAs) in Hepatocellular Carcinoma

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Abstract
Hepatocellular carcinoma (HCC) is one of the most frequently occurring cancers with poor prognosis, and novel diagnostic or prognostic biomarkers and therapeutic targets for HCC are urgently required. With the advance of high-resolution microarrays and massively parallel sequencing technology, IncRNAs are suggested to play critical roles in the tumorigenesis and development of human HCC. To date, dysregulation of many HCC-related IncRNAs such as HULC, HOTAIR, MALAT1, and H19 have been identified. From transcriptional “noise” to indispensable elements, IncRNAs may re-write the central dogma. Also, IncRNAs found in body fluids have demonstrated their utility as fluid-based noninvasive markers for clinical use and as therapeutic targets for HCC. Even though several IncRNAs have been characterized, the underlying mechanisms of their contribution to HCC remain unknown, and many important questions about IncRNAs need resolving. A better understanding of the molecular mechanism in HCC-related IncRNAs will provide a rationale for novel effective IncRNA-based targeted therapies. In this review, we highlight the emerging roles of IncRNAs in HCC, and discuss their potential clinical applications as biomarkers for the diagnosis, prognosis, monitoring and treatment of HCC.

Introduction
In the last decade, HCC has become one of the most frequently occurring cancers, and is considered to be highly lethal, accounting for approximately one-third of cancer-related
deaths worldwide [1, 2]. HBV or HCV infection, alcohol and tobacco use and liver cirrhosis are the major causes of HCC [2, 3]. Despite advances in the understanding of the molecular mechanisms underlying HCC and improved treatments for HCC, the overall survival time is still limited.

Based on the central dogma, the role of protein-coding genes in the pathogenesis of HCC has been the focal point of research. However, with the advance of technologies such as high-resolution microarray and massively parallel sequencing, non-coding transcripts including small transcripts (<200 nucleotides in length) and long non-coding RNAs (>200 nucleotides in length), which were previously regarded as transcriptional noise or garbage for their lack of coding ability, have been highlighted, and have been characterized and functionally annotated [4-6]. Based on previous experimental studies, it is well accepted that small ncRNAs including miRNAs play vital roles in the regulation of gene expression by transcriptional and post-transcriptional destabilization [7, 8]. More recently, while lncRNAs are among the least well-studied transcripts, functional studies have revealed that lncRNAs transcribed by RNA polymerase II are associated with the carcinogenesis of several cancers including HCC [9]. Recent studies indicated that HCC-related lncRNAs play critical regulatory roles in the development and progression of HCC, while their dysregulation is associated with diverse biological processes including proliferation, differentiation, apoptosis, invasion, and metastasis [10, 11] (Table 1).

Investigation of HCC-associated lncRNAs and their biological functions are important for understanding the development and progression of HCC, and hence enable the production of more efficient treatments. Here, we briefly outline lncRNAs and highlight the prospects of several major HCC-related lncRNAs.

Characteristics of lncRNAs

LncRNAs can be classified into five categories based on their location relative to neighboring protein-coding genes: (i) sense, lncRNA overlaps with the sense strand of a protein-coding gene; (ii) antisense, lncRNA overlaps one or more exons of a protein-coding gene on the opposite strand and initiate 3’ of a protein-coding gene; (iii) bidirectional, the expression of an lncRNA and a protein-coding gene on the opposite strand are initiated <1,000 base pairs away in close genomic proximity; (iv) intronic, lncRNA initiated completely within an intron of a protein-coding gene without overlapping exons; (v) intergenic (also termed large intervening non-coding RNAs or lincRNAs), lncRNA located near no other protein-coding loci [12-17] (Fig. 1).

Given that structural studies of lncRNAs are in their infancy, we describe the existing structural data for lncRNAs. Primary, secondary and tertiary structures are the three basic levels of lncRNA structure and sequence composition [17]. Insights into the structural architecture of lncRNAs provide a better understanding of the molecular mechanisms responsible for functional lncRNAs. As the fundamental components of functional lncRNAs for Watson-Crick base pairing and mediation of unpaired regions [18], secondary structures also have duplexes, bulges, hairpins, internal loops and junctions that can provide binding sites for proteins. Moreover, the high-level structure of lncRNAs provides interacting interfaces as well as maintaining lncRNA stability by the triple helix at the 3’ lncRNA end, which is used to stabilize poly(A) tail-lacking lncRNAs [19].

Functions and pathways involving lncRNAs targets

To date, numerous oncogenic genes have been confirmed as targets for HCC-related lncRNAs (Table 1). For example, lncRNAs can suppress cancer cell metastasis by targeting related genes such as HBx, AKT1, GSK3B, Cdc25A, E2F1, hnRNP U, PCAF, GF, PCAF, DMR, ICR, IGF2 and P53 [15, 20-35]. Moreover, affecting apoptosis by DNMT, ABCG2 and TSA also
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contributes to lncRNA-mediated tumor growth [34, 36-38]. Regulation of some growth factor-related genes such as Axin1, PCNA, TGF-β1, ZAK, CTCF, cyclin D1, EZH2, WDR5, P18,
PRKACB, CREB and MLL has been identified in HCC [25, 35, 39-51]. In addition, there is evidence that lncRNAs have important roles in promoting metastasis by targeting factors such as HOXD, VEGF, MMP-9, PRC2, H3K27, RBM38, caspase-3/-8, Bax, Bcl-2, BclxL, ZEB, and IL-11[23, 33, 52-56]. Moreover, lncRNA can inhibit angiogenesis by targeting PGK1 [13, 57]. By regulating these related genes, HCC-related lncRNAs exert potent tumor-suppressive effects. However, the number of known target genes is growing quickly, indicating a complicated regulatory network for lncRNAs.

Generally, lncRNAs can influence almost every cellular behavior of the central-dogma, from transcription to translation by diverse mechanisms including epigenetic alterations, lncRNA–miRNA interactions, protein-lncRNA interactions and genetic variations [44, 58-61] (Fig. 2). LncRNA-XIST, which has a well-known essential role in X-chromosome inactivation (XCI), causes stable epigenetic silencing of numerous genes on the X chromosome during female development by recruiting the crucial chromatin-modifying complex polycomb repressive complex 2 (PRC2) [62]. Another mechanism includes lncRNAs-activated by TGF-β (lncRNA-ATB), which upregulates ZEB1 and ZEB2 by competitively binding the miR-200 family to induce the epithelial–mesenchymal transition (EMT) and invasion [54]. In addition, upregulation of LncRNA-HEIH in HCC plays an important role in G0/G1 arrest with cooperation of enhancer of zeste homolog 2 (EZH2), which also requires the repression of the EZH2 target gene [45]. Wang et al. confirmed that the hPVT1–NOP2 cell cycle gene pathway in which plasma-cytoma variant translocation 1 (hPVT1) first upregulates nucleolar protein homolog 2 (NOP2) proteins and then the function of hPVT1, which is dependent on the presence of NOP2 proteins and is involved in promoting cell cycling, cell proliferation and carcinogenesis in HCC cells [30]. High expression of upregulated in hepatocellular carcinoma (URHC) can promote cell proliferation and inhibit apoptosis by repressing sterile alpha motif and leucine zipper containing kinase AZK (ZAK) expression through inactivation of the extracellular signal-regulated kinase (ERK)/mitogen-activated protein kinases (MAPK) pathway [47]. Similarly,
by binding to heterogeneous nuclear ribonucleoprotein (hnRNP) U protein, and disrupting the hnRNP U-actin complex, H19 can prevent RNA polymerase II-mediated transcription, which first inhibits the phosphorylation of the RNA Pol II C-terminal domain (CTD) at Ser5 [27]. Surprisingly, the different scale mutations in lncRNAs were also recently revealed to be highly associated with HCC. In a study of 1344 HBV persistent carriers and 1300 HBV-positive HCC patients, Liu et al. found two single nucleotide polymorphisms, rs7763881 in HULC and rs619586 in MALAT1; the data showed that mutations of rs7763881 in HULC were associated with decreased HCC risk, while mutations of rs619586 in MALAT1 were associated with decreased HCC risk with borderline relevance [26].

Dysregulation of several key lncRNAs in HCC

As a new type of regulator of cellular processes including proliferation, apoptosis, and carcinogenesis, lncRNAs play irreplaceable roles in the progression of HCC. Dysregulation of lncRNAs in HCC marks the spectrum of disease, and has been proposed to be related to hepatocarcinogenesis. Even though the underlying mechanism of HCC-related lncRNAs remains unknown, understanding the differential expression and potential functional roles of lncRNAs in HCC is essential. Accordingly, here we highlight four comparatively known HCC-related lncRNAs: HULC, HOTAIR, MALAT1 and H19.

HULC (highly upregulated in liver cancer)

Panzitt et al. [40] first reported HULC, which is located on chromosome 6p24.3, as a novel mRNA-like non-coding RNA that is dramatically upregulated in HCC compared with normal liver tissues. In a study by Du et al. [41], the upregulation of HULC mediated by HBx promoted the proliferation of HCC through downregulation of the tumor suppressor gene CDKN2C (p18) at the mRNA and protein level. As a tumor suppressor gene located nearby, CDKN2C was reported to regulate cell cycle and to function within signaling pathways including ATM/ATR and p53 [41]. Similarly, Wang et al. pointed out that phospho-CREB stimulates HULC expression by interacting with the HULC promoter via a binding site located between -67 and -53 nt, and the upregulated lncRNA-HULC expression acts as an endogenous ‘sponge’ by interacting with miR-372 [63]. In the cell cycle, inhibition of miR-372 reduced translational repression of the target gene PRKACB, which in turn induced phosphorylation of CREB. The inactive holoenzyme of PKA is composed of two regulatory and two catalytic subunits, which, following splicing of cAMP, joins a dimer of regulatory subunits bound to four cAMP and two free monomeric catalytic subunits [64]. Finally, as the catalytic subunit, PRKACB, through its miR-372 binding site in its 3′ UTR, phosphorylates CREB [63]. Based on these results, Wang et al. then speculated that activated CREB protein could recruit histone acetyltransferases including P300 and CBP to the core promoter, which leads to acetylation of the histone tail and open chromatin structure. Ultimately, this kind of mechanism occurring at the proximal promoter provides chromatin accessibility to polymerase II and initiates the transcription of HULC [63]. These findings indicate that the expression of HULC in plasma can be used as a non-invasive promising novel biomarker for the diagnosis and prognosis of HCC.

HOTAIR (HOX transcript antisense RNA)

HOTAIR is a lincRNA in the HOXC locus located on chromosome 12q13.13, which binds to and targets the PRC2 complex to the HOXD locus and can regulate gene expression by promoting genomic relocalization of PRC2 and H3K27 trimethylation [65, 66]. PRC2 is a histone H3 lysine 27 methylase related to developmental gene silencing and cancer progression [67]. Previous studies on breast cancer metastasis revealed that HOTAIR
trimethylates H3K27 to repress expression of specific suppressor genes based on PRC2 and polycomb group (PcG) [66]. H3K27 trimethylation can be regulated by the binding of the 5’domain of HOTAIR to PRC2 and the 3’terminus of HOTAIR to the LSD1/CoREST/REST complex [68]. Notably, HCC patients with high HOTAIR expression had significantly poorer prognoses than those without HOTAIR expression [69]. Inspired by the observation of HOTAIR dysregulation in breast cancer progression, Yang et al. investigated the biology of HOTAIR in HCC progression for the first time and demonstrated that high expression levels of HOTAIR in HCC compared with noncancerous tissues could be used as a candidate biomarker for HCC recurrence and shorter survival [39]. In terms of the specific molecular mechanism of HOTAIR in promoting HCC cell migration and invasion, the finding of Ding et al. suggested that HOTAIR plays a critical role in HCC progression by repressing RNA binding motif protein 38 (RBM38) [70, 71]. This previous study described that RBM38 could be targeted by P53 and induce cell cycle arrest in G1 by stabilizing the CDK inhibitor p21 [72, 73]. Similar to breast cancer, to decrease cell viability and invasion, and increase the sensitivity of cancer cells to doxorubicin and cisplatin, inhibition of HOTAIR by siRNA is potential therapy in a liver cancer cell line.

**MALAT1 (metastasis-associated lung adenocarcinoma transcript 1)**

MALAT1, which is expressed in both human and mouse tissues, is also known as nuclear-enriched abundant transcript 2 (NEAT2) and is transcribed from chromosome 11q13. It is upregulated in many solid carcinomas and is associated with cell proliferation and migration through the modulation of caspase-3, caspase-8, Bax, Bcl-2, and Bcl-xL [71]. Tripathi and colleagues demonstrated that MALAT1 localizes to nuclear speckles and interacts with several pre-mRNA splicing factors, acting as a “molecular sponge” and playing a critical role in the balance of the ratios of phosphorylated to dephosphorylated SR proteins [74]. SR proteins are a class of RNA-binding proteins that function in constitutive splicing and alternative splicing (AS), which is a key step in the regulation and diversification of gene function through their sequence-specific recognition of cis-acting exonic splicing enhancers and subsequent recruitment of other splicing factors to facilitate the assembly of the spliceosome [75-77]. At present, it is not clear how MALAT1 alters the ratios of phosphorylated to dephosphorylated SR proteins. A study by Lai et al. [55] was the first to examine the role of lncRNA-MALAT1 in HCC prognosis; following siRNA knockdown of MALAT1, decreased cell proliferation, inhibited migration and invasion as well as multi-stimuli-induced apoptosis were observed, indicating that silencing MALAT1 activity in HCC might be a vital anticancer therapy. However, MALAT1 regulates endogenous target genes by interacting with a serine/arginine family of nuclear phosphoproteins; as a result, other splicing factors such as C3 and SF2/ASF antigen are influenced and the cellular levels of phosphorylated forms of SR proteins are changed. Then finally, pre-mRNAs such as CTHRC1 and several other motility related genes are modulated by alternative splicing [13].

**H19**

As an abundantly rich and conserved transcript, H19 is a paternally imprinted oncofetal gene and is located at chromosome 11p15.5, from where it is expressed in embryonic and extra-embryonic cell lineages [23, 28]. Most reports show that dysregulation of H19 is involved in HCC deterioration. Based on the orthotopic xenograft experiments of Zhang et al. [26], decreased H19 expression in HCC specimens had more regressive and metastatic properties. Moreover, their analysis also indicated that by altering the epigenetic activation of miR-200 with cooperation of the protein complex hnRNP U/PCA/F/RNA Pol II, H19 could suppress HCC metastasis and cause EMT. Similarly, based on the conclusion of Jun et al. [78], H19, working together with miR-675, promotes migration and invasion of HCC through the
AKT/GSK-3β/Cdc25A signaling pathway, providing evidence that this type of IncRNA could be a potent diagnostic biomarker and therapeutic target for HCC.

Even though the functional and regulatory mechanism of HCC-related H19 remains elusive, united epigenetic dysregulation of insulin-like growth factor 2 (IGF2) and H19 is most likely. In most normal adult tissues, only one imprinted gene is expressed; either the paternal allele of IGF2 or the maternal allele of H19 [31]. During HCC, increased expression of IGF2 leads to the expression of H19 and the loss of adult-type promoter (P1); as a result, the fetal-type promoters (P2-P4) are re-imprinted [23]. Furthermore, multiple systems are involved in transcriptional regulation of the two genes, including an IGF2–H19 endodermal enhancer, a differentially methylated region (DMR) and an imprinted control region (ICR) [20, 30]. Epigenetic and genetic abnormality at the IGF2 and H19 loci are reported in many studies. Together with these previous studies, Takeda et al. [79] suggested that perturbations of IGF2 and H19 imprinting status occur as tumor-type specific events in the development of HCC [53]. Kim et al. revealed the biallelic expression of H19 and IGF2 in HCC could play a causal role in the epigenetic mechanism, and loss of imprinting (LOI) of IGF2 in HCC was associated with co-expression for H19 and IGF2 [54]. Li et al. reported that HCC-related H19 and IGF2 were regulated in parallel with variable expression levels [55]. Also, Sohda et al. found that the two genes were coordinately overexpressed in 37% of HCCs, and the almost identical cellular localization and spatiotemporal distribution suggested the presence of a reciprocal relation among the two genes [56]. In conclusion, since the altered transcription of IGF2 and H19 contributes to progression of HCC, the two imprinted genes may potentially be used as monitoring or diagnostic markers for HCC.

**LncRNAs as novel biomarkers and therapeutic targets for HCC**

Surgery, chemotherapy and biologics are relatively effective therapies for HCC. However, once it has become metastatic, the majority of HCC remains incurable and has a poor prognosis [80]. Molecular-based tumor predictors are essential for individualized HCC. In the past, cancer-specific miRNAs are widely detectable in the blood, urine, sputum and other biological fluids of patients, thus indicating their suitability as potential biomarkers for diagnosis and prognosis of cancers. Similarly, IncRNAs that are found in body fluids by next generation technologies, including RNA immunoprecipitation, sequencing techniques, microarray, and quantitative real-time PCR, have demonstrated the utility to be used as fluid-based noninvasive markers for clinical use.

For example, further studies indicated that HEIH as an oncogenic IncRNA was significantly associated with the repression of EZH2 and could act as an independent prognostic factor for HCC [81]. A study by Tu et al. proved for the first time that GAS5 (growth arrest-specific transcript 5) expression was decreased in HCC specimens compared with normal matched tissues and could be used as a potential and independent valuable biomarker for predicting the clinical outcome of patients with HCC [25]. Additionally, Takahashi et al. revealed that linc-ROR was related to the modulation of cellular responses to chemotherapy [63]. Based on this conclusion, molecular-targeted therapy can be used to enhance sensitivity to HCC treatment modalities and improve responses to effective therapeutic agents. As an intercellular signaling mediator and extracellular vesicle-mediated transfer, TUC339 emphasizes the potential for modulating target cell behaviors, which means such an extracellular vesicle IncRNA can be used as a potential marker for HCC [42].

Since IncRNAs can target multiple genes/pathways similarly to miRNAs, re-establishing the expression of a single IncRNA to that of the non-diseased tissue will produce a more pervasive therapeutic effect compared with drugs that obey the one-drug-one-target paradigm [82]. IncRNA replacement therapy will be particularly attractive in the field of oncology, as many IncRNAs exhibit reduced expression in cancer target messenger RNAs of genes that are oncogenic. Further, toxicity may be minimized as IncRNA replacement therapy donates gene products that are already present in the normal tissue. In addition, IncRNAs
can influence the sensitivity of HCC to chemo- or radiation therapy. Thus, in the future, a better understanding of the molecular mechanism in HCC-related IncRNAs will provide a rationale for novel effective IncRNA-based targeted therapies.

Conclusions and future perspectives

In recent years, there has been exponential growth in research on the biological functions of IncRNAs in various cancers, including HCC. As one of the most common worldwide diseases, with highly aggressive malignancy and poor prognosis, early diagnosis and discovery of therapeutic targets for HCC is necessary. With application of next generation sequencing and high-resolution of microarray techniques, many studies suggest HCC-related IncRNAs can influence the initiation, progression and treatment of HCC. IncRNA are defined as transcripts that are longer than 200 nucleotides without obvious protein coding functions. As abovementioned, HCC-related IncRNAs play critical regulatory roles in the progression of HCC while their dysregulation is associated with diverse biological processes [10]. Dysregulation of HULC, HOTAIR, MALAT1, H19 and others has been identified.

Lack of targetable oncogenic driver and the heterogeneous nature of HCC limit the effectiveness of therapies. Molecular-based tumor predictors are essential for individualized HCC. Cancer-specific miRNAs have been widely regarded as potential biomarkers for the diagnosis and prognosis of cancers as they are easily detectable in the blood, urine, sputum and other biological fluids of patients. Similarly, IncRNAs that are found in body fluids have demonstrated the utility to be used as fluid-based non-invasive markers for clinical use. Even though some IncRNAs have been characterized, the underlying mechanism contributing to HCC remains unclear; and it is well accepted that IncRNAs can bring tremendous novel insights into the diagnosis and treatments of HCC. Accordingly, it is foreseeable that formidable efforts will be put in the development of IncRNA-based therapy, but there is a long way to go before they can be used in the prevention and treatment of HCC in the clinic.

In summary, with the rapid developments in genomics, proteomics and bioinformatics, an increasing number of IncRNAs are emerging as novel biomarkers for early diagnosis, better prognostic evaluation and efficient therapeutic targets for HCC in future clinical applications.

Abbreviations

IncRNAs (long noncoding RNAs); HCC (hepatocellular carcinoma); XCI (X-chromosome inactivation); PRC2 (polycomb repressive complex 2); IncRNA-ATB (IncRNA-activated by TGFB); EMT (epithelial-mesenchymal transition); EZH2 (enhancer of zeste homolog 2); hPVT1 (plasma-cytoma variant translocation); NOP2 (nucleolar protein homolog 2); URHC (upregulated in hepatocellular carcinoma); ZAK (sterile alpha motif and leucine zipper containing kinase AZK); ERK (extracellular signal-regulated kinase); MAPK (mitogen-activated protein kinases); hnRNP (heterogeneous nuclear ribonucleoprotein); CTD (C-terminal domain); HULC (highly upregulated in liver cancer); HOTAIR (HOX transcript antisense RNA); PcG (polycomb group); MALAT1 (metastasis-associated lung adenocarcinoma transcript 1); NEAT2 (nuclear-enriched abundant transcript 2); IGF2 (insulin-like growth factor 2); DMR (differentially methylated region); ICR (imprinted control region); LOI (loss of imprinting); AFP (elevated alpha-fetoprotein); CEA (carcinoembryonic antigen); GAS5 (growth arrest-specific transcript 5).

Disclosure Statement

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