Diagnosis and Classification of Malignant Lymphoma and Related Entities in the Bone Marrow Trephine Biopsy

Falko Fend  Markus Kremer
Institute of Pathology, Technical University Munich, Munich, Germany

Abstract
The trephine bone marrow (BM) biopsy is an important diagnostic tool in patients with malignant lymphoma. BM examination can serve to establish or confirm a primary diagnosis of lymphoma or to determine the extent of disease dissemination for staging purposes. BM histology renders information which cannot be gained equally from aspirate material, such as spacial distribution and extent of infiltrates, BM cellularity and fibrosis. Furthermore, cytology including flow cytometric immunophenotyping can give false-negative results in BM involvement by lymphoma due to intraleSIONal fibrosis. In addition to morphological examination, the availability of a broad panel of antibodies suitable for paraffin-embedded tissues, in conjunction with less damaging decalcification procedures, nowadays enables us to perform complete immunophenotyping on BM trephines and allows for classification of lymphoma infiltrates according to established algorithms. Molecular determination of clonality and interphase fluorescent in situ hybridization can be employed selectively to resolve difficult cases. This review describes important diagnostic features of malignant lymphoma in the BM, relevant differential diagnoses, and the proper use of ancillary techniques.

Introduction
The trephine bone marrow (BM) biopsy is an integral part of the diagnostic work-up of patients with a diagnosis of malignant lymphoma [1–7]. In most instances, BM biopsy is performed for staging or follow-up purposes, but in a significant minority of cases, BM examination serves as a means for primary diagnosis, especially in disseminated low-grade B-cell non-Hodgkin’s lymphoma (B-NHL). The absence or presence of BM infiltration has a significant impact on prognosis and treatment options in a variety of NHL and Hodgkin’s disease.

In lymphoma patients, the trephine biopsy renders important diagnostic information which cannot be obtained from BM aspirates, namely on architectural features of the infiltrate and its topographic distribution as well as the amount of marrow space occupied by a malignant infiltrate [4, 6, 7]. Furthermore, assessment of marrow involvement by histology is not impeded by fibrosis, which is a frequent finding in lymphomatous infiltrates and may lead to false-negative results in BM aspirates due

Key Words
Bone marrow · Immunohistochemistry · Molecular pathology

This work was supported in part by DFG Grant FE597/1-2 to F.F. and M.K.

KARGER
Fax +41 61 306 12 34
E-Mail karger@karger.ch
www.karger.com

Falko Fend, MD
Institute of Pathology, Technical University Munich
Ismaningerstrasse 22
DE–81675 Munich (Germany)
Tel. +49 89 4140 4162, Fax +49 89 4140 4865, E-Mail fend@lrz.tum.de

© 2007 S. Karger AG, Basel
1015–2008/07/0742–0133$23.50/0
Accessible online at:
www.karger.com/pat

Copyright © 2007 S. Karger AG, Basel

Downloaded by: 54.70.40.11 - 11/19/2017 7:15:06 PM
to sampling error [8–10]. For these reasons, the performance of a unilateral (or bilateral) BM trephine of adequate size with a minimal length of 2 cm, preferably examined by step sectioning, is regarded mandatory in patients with malignant lymphoma both for the initial determination of BM involvement and response evaluation [11, 12]. However, trephine biopsy, aspiration cytology and flow cytometric immunophenotyping should be seen as integral parts of a combined approach ideally resulting in a joint diagnostic assessment [1, 3].

The development of a broad range of antibodies suitable for paraffin-embedded tissues, in conjunction with decalcification techniques, which are non-detrimental for most relevant antigens, has increased the diagnostic value of the BM biopsy further and has made it complementary, and in many instances superior to flow cytometric immunophenotyping of BM aspirates [6, 7, 13]. The present review gives an overview of the practical issues in the diagnostic evaluation of BM trephines in patients with malignant lymphoma and critically reviews the role of ancillary techniques.

Frequency and Patterns of BM Infiltration in Malignant Lymphoma

The frequency of BM infiltration varies widely for different subtypes of malignant lymphoma and ranges from 5% in Hodgkin’s lymphoma [14, 15] to almost 100% for patients with small lymphocytic B-cell lymphoma/chronic lymphocytic leukemia [1]. BM infiltration by malignant lymphoma is not a random process but is influenced by characteristics of the neoplastic cells as well as micro-environmental features, leading to a characteristic pattern of marrow involvement for many lymphoma subtypes [4, 6, 7, 13]. These infiltration patterns, though not absolutely specific, may provide diagnostic clues both for the discrimination of benign and malignant infiltrates as well as for NHL subtyping. Five major patterns of BM infiltration can be discerned, namely a nodular intertrabecular, a nodular paratrabecular, an interstitial, a diffuse and an intrasinusoidal pattern. However, patterns can change during the course of the disease, and mixed patterns occur frequently. For neoplasms such as B-CLL, not only the presence, but the amount as well as the distribution of BM infiltrates are of clinical relevance, although the independent prognostic significance of BM pattern in B-CLL has recently been challenged [16–18].

The differential diagnosis of various lymphoma subtypes in the BM is based on a combination of architectural and cytological features, complemented by immunohistochemistry. Whereas most lymphomatous BM infiltrates are readily identifiable by morphology, early interstitial infiltrates and the intrasinusoidal pattern may easily be overlooked in standard histological stains and require immunohistochemistry for their detection [19–21]. The following summarizes important diagnostic features of major lymphoma subtypes.

Hodgkin’s Lymphoma

BM infiltration is rare both in classical Hodgkin’s lymphoma as well as in lymphocyte-predominant Hodgkin’s lymphoma (nodular paragranuloma), occurring in less than 5% in most large clinical series [14, 15]. For these reasons, some authors have advocated to forgo BM examination in patients lacking clinical risk factors such as B symptoms and involvement on both sides of the diaphragm. In contrast to Hodgkin’s lymphoma in immunocompetent patients, BM involvement in patients with HIV infection is found in 25–50% of cases, and a BM trephine may be the site of primary diagnosis [22–24].

BM infiltration by Hodgkin’s lymphoma is characterized by randomly distributed infiltrates consisting of a mixture of small lymphocytes, plasma cells, histiocytes and eosinophils and rare atypical cells, usually accompanied by significant fibrosis and occasional epithelioid cell granulomas [1]. In patients with a confirmed diagnosis of Hodgkin’s lymphoma at another site, the presence of such infiltrates is considered compatible with a diagnosis of BM involvement even in the absence of diagnostic Reed-Sternberg cells. Immunohistochemistry for CD30 and, with limitations due to the presence of granulocytes, CD15 is very helpful in confirming suspected BM infiltrates of classical Hodgkin’s lymphoma (fig. 1G). Infiltrates of some NHL subtypes, mainly peripheral T-cell lymphoma NOS and angioimmunoblastic T-cell lymphoma may simulate Hodgkin’s lymphoma in the BM and require either a lymph node biopsy or extensive immunophenotyping for a definitive diagnosis.

Non-Hodgkin’s Lymphoma

Small B-Cell NHL

The small B-cell NHL including B-CLL/SLL, lymphoplasmacytic lymphoma (LPL), follicular lymphoma (FL), mantle cell lymphoma (MCL), marginal zone B-cell lymphomas and hairy cell leukemia (HCL) frequently involve
the BM and make up for a large percentage of BM infiltrates in patients with malignant lymphoma [2, 4, 6, 7, 13]. The diagnostic criteria for the different small B-cell lymphoma entities are summarized in Table 1.

**Table 1. Differential diagnosis of small B-NHL**

<table>
<thead>
<tr>
<th>Diagnosis</th>
<th>Common infiltration patterns</th>
<th>CD20</th>
<th>CD5</th>
<th>CD23</th>
<th>Cyclin D1</th>
<th>Additional markers</th>
<th>Comments</th>
</tr>
</thead>
<tbody>
<tr>
<td>B-CLL</td>
<td>nod/i, int, diffuse</td>
<td>+ (w)</td>
<td>+</td>
<td>-</td>
<td></td>
<td>CD43+, ZAP70+/−</td>
<td>occasional proliferation centers</td>
</tr>
<tr>
<td>MCL</td>
<td>nod/p+i, int</td>
<td>+</td>
<td>+</td>
<td>−</td>
<td>+</td>
<td>CD43+, p27−</td>
<td></td>
</tr>
<tr>
<td>MZL</td>
<td>nod/i</td>
<td>+</td>
<td>−</td>
<td>−</td>
<td></td>
<td>CD43−/+</td>
<td></td>
</tr>
<tr>
<td>LPL</td>
<td>nod/i</td>
<td>+</td>
<td>−/+</td>
<td>−</td>
<td></td>
<td>clg+</td>
<td></td>
</tr>
<tr>
<td>SMZL</td>
<td>nod/i, sin</td>
<td>+</td>
<td>−</td>
<td>−</td>
<td></td>
<td>IgD+</td>
<td></td>
</tr>
<tr>
<td>FL</td>
<td>nod/p</td>
<td>+</td>
<td>−</td>
<td>−</td>
<td>−</td>
<td>CD10+, bcl-6+, CD43−</td>
<td></td>
</tr>
<tr>
<td>HCL</td>
<td>int, diffuse</td>
<td>+</td>
<td>−</td>
<td>−</td>
<td>+/−</td>
<td>DBA44+, CD25+, TRAP+, CD11c+, Annexin A1+</td>
<td></td>
</tr>
<tr>
<td>MM</td>
<td>random</td>
<td>−/(+)</td>
<td>−</td>
<td>−</td>
<td>−/−</td>
<td>Ig light chains, CD138+, CD38+, VS38c+, CD56+/−</td>
<td></td>
</tr>
</tbody>
</table>

MCL = Mantle cell lymphoma; MZL = marginal cell lymphoma; LPL = lymphoplasmacytic lymphoma; SMZL = splenic MZL; FL = follicular lymphoma; HCL = hairy cell leukemia; MM = multiple myeloma; nod/i = nodular intertrabecular; nod/p = nodular paratrabecular; int = interstitial; sin = sinusoidal; w = weak.

invariably associated with unmutated immunoglobulin variable genes [29, 30], can also be detected in routinely processed BM biopsies. ZAP-70 positivity is more frequently found in cases with diffuse BM infiltration, a pattern previously known to be associated with higher stage and poor prognosis [25]. However, ZAP-70 expression is not limited to B-CLL, but may be found in a variety of other NHL [31–33].

B-prolymphocytic leukemia is a rare disease characterized by high leukocyte counts, splenomegaly and BM infiltrates by cells with wider cytoplasm and enlarged nuclei with prominent nucleoli. It usually lacks CD5 expression. The precise classification of these cases is currently under discussion. Cases of B-PLL carrying a t(11;14) as described in the older literature will currently be regarded as leukemic MCL by most authors [34].

MCL involves the BM in 55–90% of cases and can show a variety of patterns, including predominantly paratrabecular infiltrates. Demonstration of the characteristic CD5+/CD23− immunophenotype and of cyclin D1 overexpression facilitates the diagnosis [27]. Cyclin D1 expression, especially when investigated with the new monoclonal rabbit antibody (SP-4, DCS, Germany) and complemented by CD20 staining, is very sensitive for the detection of residual disease (fig. 1F). Blastoïd and pleomorphic variants of MCL have to be distinguished from lymphoblastic lymphoma/acute leukemia and large cell lymphoma, respectively. Another characteristic feature of MCL is the lack of the CDK inhibitor p27, which is strongly expressed by other small B-cell neoplasms with the exception of HCL [35].
Fig. 1. Patterns of BM infiltration by malignant lymphoma.
A CD20 staining of a characteristic paratrabeular FL infiltrate. Note the characteristic ‘wallpaper-like’ infiltration pattern.
B, C Two microphotographs representing a DLBCL with a ‘pseudo-discordant’ BM infiltration. In the HE staining (B), a densely packed lymphocytic infiltrate is noticed, with only a few blasts. The CD20 immunostain (C) highlights the scattered blasts within the background of ‘reactive’ T cells simulating a ‘discordant’ infiltration.
D, E A typical case of HCL with a loose interstitial infiltrate of hairy cells (D). The tumor cells are strongly positive for CD20 (E).
F Cyclin D1 staining of a MCL, with a strong immunoreactivity of neoplastic lymphocytes, intermingled with cyclin D1-negative T cells.
G CD30-positive Hodgkin and Reed-Sternberg cells in a BM infiltrate of classical HL.
H, I A case of a periph-
FL, especially of lower grades, shows BM involvement in 50–60% of cases. FL classically shows strictly paratrabeicular, rather loose infiltrates accompanied by significant fibrosis (fig. 1A) [1, 4, 6]. True neoplastic follicles are rare, occurring in less than 5% of cases, and can be distinguished from reactive lymphoid follicles by their bcl-2 positivity [36]. Due to the characteristic distribution of the neoplastic infiltrates, immunohistochemistry is of minor importance in cases with an established diagnosis of FL, but occasionally can detect small linear accumulations of neoplastic B cells lining the bony trabecules [10, 37]. The neoplastic infiltrates in FL usually contain a high number of reactive T cells, which should not distract from a diagnosis if the typical morphological features are present. Of note, CD10 expression can be absent in BM infiltrates and is difficult to evaluate due to positive staining of other marrow elements. A relatively frequent phenomenon of BM infiltration in FL and also diffuse large B-cell lymphoma (DLBCL) is the presence of predominantly small cell infiltrates in cases with a diagnosis of higher-grade FL or DLBCL at another site. These so-called ‘discordant’ infiltrates can be evidence of either disease transformation at an extramedullary location, the presence of a second, unrelated lymphoid neoplasm in the BM or might even be due to the influence of the BM microenvironment [38–40]. True discordant infiltrates need to be discerned from cases of high-grade lymphoma in which the neoplastic large cells are obscured by large numbers of small T cells, simulating a lymphomatous infiltrate of lower grade. Immunohistochemistry for CD20 and CD3 helps to resolve these cases (fig. 1B, C).

LPL (corresponding to the lymphoplasmacytic immunocytoma of the Kiel classification) is most often encountered in the presence of the clinical syndrome of Waldenström’s macroglobulinemia with the presence of an IgM paraprotein and BM involvement [41, 42]. Histologically, the BM shows variably distributed, often interstitial and diffuse infiltrates of small lymphocytes, lymphoplasmacytoid cells and plasma cells with variable fibrosis and a characteristic admixture of mast cells. Dutcher bodies may be frequent. Demonstration of cytoplasmic immunoglobulin expression and the frequent, though not obligatory absence of CD5 expression can help to confirm the diagnosis.

Nodal and extranodal marginal zone B-cell lymphomas infiltrate the BM less frequently than other small B-cell NHL, in approximately 10–40% of cases [43–45], possibly with a higher frequency for nodal MZL [46]. They usually show interstitial, nodular infiltrates, which may contain reactive germinal centers. Their phenotype (CD20+, CD10–, CD43+/−, CD5−) with absence of characteristic markers, makes differentiation from reactive nodules and other NHL subtypes difficult. The rare CD5+ extranodal MZL cases seem to show a higher tendency for BM involvement [47].

Splenic marginal zone B-cell lymphoma (SMZL) [20, 48–50], which encompasses splenic lymphoma of villous lymphocytes, shows a much higher frequency of BM infiltration than the other marginal zone B-NHLs. In addition to nodular and interstitial infiltrates, SMZL characteristically shows an intrasinusoidal pattern [19, 21, 51]. In cases of pure intrasinusoidal spread, the infiltrates are very hard to detect by conventional morphology, and CD20 immunohistochemistry is required for suspected cases of SMZL. An important pitfall is persistent polyclonal lymphocytosis with binucleated lymphocytes, a benign lymphoproliferation usually found in female smokers, which can closely simulate the sinusoidal pattern of BM involvement of SMZL [52].

HCL is clinically characterized by splenomegaly, frequent cytopenia with monocytopenia, presence of circulating hairy cells and a dry tap in BM examination. Morphologically, the characteristic infiltrate shows interstitial, in early cases rather inconspicuous aggregates of medium-sized lymphoid cells with clear cytoplasm and oval to bean-shaped nuclei, accompanied by reticulin fibrosis (fig. 1D) [3]. The cells show strong expression of CD20, frequently giving them a ‘woolly’ or ‘hairy’ appearance (fig. 1E). In addition, they are usually positive for the monoclonal antibody DBA.44, tartarate-resistant acid phosphatase, CD25 (IL-2 receptor) and in the majority of cases for cyclin D1 [35, 53–55]. Hairy cells lack CD5 and CD23. A highly specific new marker for primary diagnosis is annexin-1, which however is not suitable for residual disease detection due to its expression in granulocytes [56]. This unique immunoprofile helps to differentiate HCL from other B-NHL in difficult cases and aids in the detection of minimal residual disease.
Diffuse large cell B-cell lymphoma has a rather low frequency of BM infiltration, ranging from 10 to 35% [3]. The patterns of infiltration are highly variable, ranging from subtle, focal infiltrates to a ‘packed marrow’ with complete replacement of hematopoiesis. As mentioned above, so-called discordant infiltrates in patients with DLBCL at another site consist of small neoplastic cells and carry a better prognosis than concordant (i.e. large cell) BM infiltration [57]. In addition to other lymphoma subtypes such as peripheral T-NHL or plasmablastic MCL, non-lymphoid neoplasms such as acute myeloid leukemia or even carcinomas may mimic DLBCL in the BM, and immunohistochemistry is important for establishing the diagnosis. A special problem is the diagnosis of T-cell-rich/histiocyte-rich large B-cell lymphoma (TCRBCL) infiltrating the BM, since it can be confused with Hodgkin’s lymphoma [58, 59]. The CD20+, EMA+, CD30–, CD15– phenotype of the neoplastic large cells helps to separate TCRBCL from classic Hodgkin’s lymphoma, but separation from nodular lymphocyte-predominant HL is virtually impossible on the BM trephine. However, the latter involves the BM rather infrequently, whereas BM infiltrates are common in TCRBCL. BM involvement by Burkitt lymphoma ranges from scattered interstitial foci to ‘packed marrow’ (fig. 1L). Burkitt lymphoma shows a mature B-cell phenotype with coexpression of CD20, CD10 and BCL-6, but lacks terminal deoxyribonucleotidyl transferase (TdT).

Plasma Cell Disorders

Multiple myeloma (MM) accounts for approximately 10% of all hematologic neoplasms. BM infiltration by MM is highly variable and does not follow a recurrent pattern. In typical cases, diagnosis of MM is straightforward by morphology alone [60]. Grading of MM based on cytological features as well as infiltration volume is of prognostic importance [61]. In addition to neoplastic infiltrates, the presence and extent of bone resorption and remodeling should be noted in the report.

In daily practice, distinction from monoclonal gammapathy of unknown significance (MGUS) and occasionally separation from other neoplasms are the most commonly encountered diagnostic problems. In addition to the percentage of plasma cells (above 10% are considered suspicious), their localization in sheets or clusters without association with vessels, cytologic atypia, a clear-cut light chain restriction and expression of CD56 are criteria supporting a diagnosis of MM rather than MGUS (fig. 1L). MM usually lacks markers of mature B cells such as CD20, but expresses the plasma cell markers CD138, CD38, VS38c, frequently CD56 and cytoplasmic immunoglobulin [60, 62–64]. This phenotype helps to differentiate MM from B-NHL with plasmacytic differentiation, such as LPL or marginal zone B-cell lymphoma. Of note, the lymphoplasmacytic or small cell variant of MM frequently coexpresses CD20 in addition to plasma cell markers and commonly shows cyclin D1 reactivity due to the presence of a (11;14) translocation [7, 65, 66]. This MM subtype needs to be distinguished from MCL. In patients without morphologically obvious neoplastic infiltrates, such as in early stage disease or after treatment, immunohistochemistry should be performed on a routine basis, since subtle interstitial infiltrates are easily overlooked, especially in small cell and well-differentiated cases. Anaplastic MM and purely plasmablastic cases need to be differentiated from large cell lymphoma and even non-hematologic neoplasms.

In recent years, the detection of non-random chromosomal abnormalities by interphase fluorescent in situ hybridization (FISH) in large series of MM has given significant insights into the biology of the disease and has provided us with important prognostic markers. Recurrent chromosomal translocations involving the immunoglobulin heavy chain locus at 14q32 and numerical aberrations such as deletions of chromosome 13 or hyperdiploid MM allow stratification into groups with widely differing survival and clinical features [67–69]. Interphase FISH can also be applied successfully to BM trephines, thus allowing for the detection of recurrent cytogenetic alterations in MM and other lymphoid neoplasms [70, 71].
megaly and features of immune dysregulation, is characterized by an interstitial and intrasinusoidal spread of CD3+, usually CD8+ T cells with expression of cytotoxic granule proteins TIA-1 and granzyme B (fig. 1J, K) [75]. Of note, the BM in T-LGL may contain reactive lymphoid nodules which contain a mixture of B cells as well as CD4+ and CD8+ T cells, a potential source of confusion. In contrast to T-LGL, T-PLL commonly shows a marked diffuse, less frequently nodular infiltrate with replacement of hematopoiisis and fat cells. In most cases of T-PLL, the neoplastic cells express CD4, together with other mature T-cell markers. Hepatosplenic T-cell lymphoma, an aggressive disease with poor prognosis, is frequently negative for CD4, CD5 and CD8 and perforin, but often expresses CD56 and is TIA-1-positive [76].

Nodal (systemic) anaplastic large cell lymphoma shows overt BM involvement in approximately 10% of cases, but is detected in about 30% of cases studied by immunohistochemistry [77]. Immunostains for CD30 or the ALK-1 protein, which is expressed in 70–90% of cases depending on age group, highlight singly dispersed neoplastic cells in the marrow.

Peripheral T-NHL, NOS shows great cytological and immunophenotypical variability. BM infiltration is relatively common and usually characterized by irregular, nodular infiltrates with a high content of reactive cells, making immunophenotypical characterization of the infiltrate difficult (fig. 1H, I) [72–74]. Angioimmunoblastic T-NHL (AITL) infiltrates the BM in the majority of cases and shows similar diagnostic features as in lymph node biopsies. Of note, CD10 reactivity, a characteristic feature of AITL, is often absent in the BM [78]. The presence of large, transformed, EBV-positive B cells in the infiltrate may point to a diagnosis of AITL, but can also lead to confusion with T-cell-rich large B-cell lymphoma or Hodgkin’s disease. BM involvement is of less importance in the extranodal cytotoxic T-NHL and rare in cutaneous T-cell lymphomas including mycosis fungoides. An important paraneoplastic change accompanying some cases of peripheral T-NHL, mainly of cytotoxic origin, is hemophagocytic syndrome [79]. This syndrome frequently shows a dramatic clinical presentation with fever, cytopenia and hepatosplenomegaly, and may also be found in association with viral and bacterial infections. The BM shows a reduction of hematopoietic elements, variable fibrosis and pronounced infiltrates of histiocytes with ingestion of red cells and other hematopoietic elements [3].

Acute lymphoblastic leukemia/lymphoblastic lymphoma of both B- and T-cell origin is usually characterized by a massive, diffuse infiltrate of monomorphic blasts with strongly reduced hematopoiesis. The neoplastic cells are monomorphic to moderately pleomorphic, with fine chromatin and small to medium-sized nucleoli. Immunohistochemistry for TdT, which is usually strongly positive in the nuclei of the blasts, CD10, CD79a, Pax-5 and T-cell markers helps to determine the cell lineage and to discriminate it from other neoplasms, mainly AML [80]. Of note, aberrant cross-lineage expression of B, T or myeloid antigens is relatively frequent in acute leukemias, as is positivity for CD34, and some cases may not be resolved satisfactorily by paraffin section immunophenotyping. For example, expression of B-cell markers Pax-5 or CD79a can be observed in subsets of AML [81, 82]. CD99 (MIC2) expression is common in ALL/LBL and may lead to confusion with other small blue round cell tumors, such as Ewing’s sarcoma [83].

Differential Diagnosis of Reactive Lymphoid Infiltrates

A common diagnostic problem in BM trephines is the distinction of reactive nodular lymphoid infiltrates from nodular infiltrates of low-grade lymphoma [1, 3, 4, 6, 84, 85]. These reactive infiltrates can be found in a variety of disorders, including autoimmune disease, infections, and chronic myeloproliferative syndromes. A more mixed cellular composition, well-defined borders, intertrabecular location, low number of infiltrates and absence of reticulin fibrosis are morphological features found more often in benign lymphoid nodules [84]. Germinal centers are relatively rare also in benign infiltrates. Immunophenotyping usually shows a predominance of T cells and a lack of an aberrant marker profile or immunoglobulin light chain restriction. Nevertheless, some cases may remain unresolved with standard diagnostic procedures and may require additional molecular studies (see below).

Interpretation of BM Trephines Post-Therapy

Description of the vast array of changes which can be encountered after chemotherapy is beyond the scope of this review, but some more common problems in follow-up biopsies of patients treated for malignant lymphoma are discussed below. The detection of minimal residual disease is easier for lymphoid neoplasms with a well-defined immunophenotype and/or morphology, such as anaplastic large T-cell lymphoma, HCL or MCL, than for predominantly small cell NHL lacking such features. For
some diseases such as HCL, criteria for defining remission by histological and immunohistochemical staining of BM trephines have been defined [86]. In treated acute lymphoblastic leukemia, combination of topography and immunophenotype helps to discern regenerating benign hematogones, immature B-cell precursors, from residual leukemic blasts, since the presence of small clusters of CD34- or TdT-positive cells is predictive of impending relapse [87, 88].

A common problem is the presence of residual nodular lymphoid infiltrates after therapy for small B-cell lymphoma. The determination of their benign or malignant nature frequently requires immunohistochemical examination. Residual nodules may consist entirely of reactive T cells, thus allowing a diagnosis of complete remission despite morphological features suggestive of persistent disease. In patients having received rituximab (anti-CD20 antibody) therapy, CD79a or CD22 should be included in the diagnostic panel, because downregulation of CD20 expression by the lymphoma cells may lead to a false-negative diagnosis [89, 90].

Role of Molecular Examination

Molecular examination of BM trephines plays only a minor role in routine diagnostic examination. Diagnostic molecular studies are usually performed on lymph node biopsies or on BM aspirates rather than trephines for reasons of easier handling and better DNA or RNA quality obtainable from fresh rather than fixed deca
calified BM samples. Molecular detection of minimal residual disease in NHL patients usually is also performed on aspirates, although the role of potential sampling error due to leisional fibrosis has to be seen critically.

Nevertheless, several groups have shown the feasibility of both DNA- and RNA-based molecular examinations of fixed and decalcified, paraffin-embedded BM trephines [reviewed in 91]. Assessment of B- or T-cell clonality is the most frequent indication for molecular studies [92–94]. In addition to the discrimination of benign and malignant lymphoid infiltrates, determination of the clonal relationship of medullary infiltrates with lymphoma at another site may help to resolve cases of discordant small cell infiltrates in patients with large cell lymphoma mentioned above [39]. Microdissection of lymphoid nodules can help to increase the sensitivity of PCR-based determination of clonality, but requires careful interpretation of results [85]. Since fixation and decalcification decreases the quality of nucleic acids and can lead to false-positive results due to preferential amplification of rare templates, rigorous quality controls and performance of diagnostic PCR assays in duplicate are mandatory. In addition to clonality determination, a range of applications for DNA- as well as RNA-based molecular studies have been described for diagnosis of lymphoma in BM trephines [91, 95, 96].

References

Malignant Lymphoma and Related Entities in the BM Trephine Biopsy


Malignant Lymphoma and Related Entities in the BM Trephine Biopsy


