Impact of Nasal Continuous Positive Airway Pressure Therapy on Markers of Platelet Activation in Patients with Obstructive Sleep Apnea

Morohunfolu E. Akinnusi\textsuperscript{a} Linda L. Paasch\textsuperscript{a} Kristie R. Sarpa\textsuperscript{a} Paul K. Wallace\textsuperscript{b} Ali A. El Solh\textsuperscript{a, c}

\textsuperscript{a}Division of Pulmonary, Critical Care, and Sleep Medicine, Western New York Respiratory Research Center, Department of Medicine, State University of New York at Buffalo School of Medicine and Biomedical Sciences,\textsuperscript{b} Department of Flow and Image Cytometry, Roswell Park Cancer Institute, and \textsuperscript{c} Department of Social and Preventive Medicine, State University of New York at Buffalo School of Public Health and Health Professions, Buffalo, N.Y., USA

Abstract

Background: Considerable evidence implicates CD40 signaling in the pathogenesis of atheromas. Exposure to CD40 ligand induces platelet-leukocyte conjugation, a heightened expression of inflammatory cytokines, matrix-degrading enzymes, and procoagulant factors. Objectives: To investigate the association between plasma soluble CD40 ligand (sCD40L) and platelet-monocyte aggregates in patients with obstructive sleep apnea (OSA) and to determine whether treatment of OSA with nasal continuous positive airway pressure (nCPAP) alters this relationship. Methods: Twelve patients with OSA who were free of other diseases and 12 healthy controls matched for age, gender, and body mass index had blood drawn for sCD40L and platelet-monocyte aggregate measurements. A repeat assessment was obtained following 8 weeks of nCPAP therapy. Results: Subjects with OSA had significantly higher plasma sCD40L levels and exhibited elevated platelet-monocyte aggregates compared to nonapneic subjects (7.6 $\pm$ 4.3 versus 1.7 $\pm$ 1.1, $p = 0.004$; and 41.3 $\pm$ 23.7 versus 6.7 $\pm$ 4.9, $p = 0.001$, respectively). Both parameters correlated positively with the percentage of time spent with SpO$_2$ < 90% ($r = 0.69$, $p = 0.01$ and $r = 0.6$, $p = 0.03$, respectively). After 8 weeks of nCPAP treatment, sCD40 levels declined by 47% ($p = 0.003$) and platelet-monocyte aggregates by 42% ($p = 0.002$). None of the controls showed any changes in either sCD40L or platelet-monocyte aggregates after nCPAP therapy. Conclusions: OSA is associated with upregulation of circulating sCD40L and platelet-monocyte aggregation that may account for the increased incidence of cardiovascular events in this population. Treatment with nCPAP may alleviate this risk.

Introduction

Obstructive sleep apnea (OSA) is a complex disorder characterized by recurrent episodes of upper airway collapse during sleep leading to oxyhemoglobin desaturation and sleep fragmentation. The syndrome has been recognized as a risk factor for hypertension, acute cardiovascular events and metabolic disturbances [1]. Different mechanisms have been advanced to explain the relationship between OSA and cardiovascular disease, including increased sympathetic activity, oxidative...
stress, inflammation, and endothelial dysfunction [2–4]. Another purported connection between OSA and cardiovascular disease is enhanced coagulability, possibly mediated by higher levels of circulating apoptotic endothelial cells [5] and platelet activation [6–8]. Indeed, several studies showed that a high percentage of patients with OSA may have a condition of in vivo platelet activation as indicated by increased levels of plasma β-thromboglobulin, serum thromboxane B₂, and soluble P-selectin [8].

Recently our understanding of the interaction between inflammation and OSA has been augmented by the report of increased levels of soluble CD40 ligand (sCD40L) in apneic subjects [9]. CD40L is a transmembrane protein that belongs to the tumor necrosis factor (TNF) family [10, 11]. It was identified originally on CD4+ T cells but has also been found on mast cells, basophils, eosinophils and activated platelets [12, 13]. Following expression on the cell surface, CD40L is partly cleaved and released into the circulation as sCD40L, where it binds to CD40, a receptor constitutively expressed on endothelial cells [14]. The ligand-receptor interaction promotes atherogenesis through platelet aggregation [15], platelet-leukocyte conjugation [16], and generation of reactive oxygen and nitrogen species [17].

In contrast to atherothrombosis, where the role of sCD40L has been systematically investigated [18, 19], the association between sCD40L and platelet-monocyte aggregates in the biology of OSA remains obscure. Because platelet-monocytes aggregates are known to contribute to ongoing injury at atheromatous sites and in plaque disruption [20], the aims of the study were to investigate the relationship between sCD40L expression and platelet-monocyte aggregates in patients with OSA and to determine whether treatment of OSA with nasal continuous positive airway pressure (nCPAP) alters this relationship.

**Methods**

**Subjects and Controls**

After receiving approval from the Institutional Review Board at the State University of New York at Buffalo, all consecutive adult patients referred to the Sleep Disorders Clinic at the Erie County Medical Center for suspected OSA between June 2006 and February 2008 were considered for participation in the study provided the following criteria were met: (1) a new diagnosis of OSA by polysomnography; (2) agreement to participate in the investigation by signing an informed consent, and (3) absence of coexisting cardiovascular diseases, diabetes mellitus, renal diseases, or active cigarette smoking. Subjects with a history of neoplastic diseases, or hematologic disorders, or who were regularly using medications were excluded. Control subjects free of comorbidities and sleep-disordered breathing, matched for age, gender, and body mass index (BMI), were enrolled.

**Polysomnography**

The diagnosis of OSA was established by standard overnight polysomnography using a computerized recording system (Alice® 5 Diagnostic Sleep System; Respironics Inc., Murrysville, Pa., USA). Electroencephalography electrodes were applied at C4-A1, C3-A2, O1-A2, O2-A1. Electrooculography, submental electromyography, appropriate leg and electrocardiography electrodes were applied as previously described. Oxyhemoglobin saturation was recorded by pulse oximetry, respiratory effort by piezoelectric belts, and nasal/oral airflow by pressure sensor. Physiologic signals were digitized (Embla Recording Systems; Medcare, Buffalo, N.Y., USA) for offline analysis of sleep and breathing patterns. Sleep stage scoring was performed on 30-second epochs according to standard criteria [21]. Apnea was defined as complete cessation of airflow for at least 10 s. Hypopnea was defined as any reduction in airflow for more than 10 s associated with electroencephalographic arousals or 4% drop in oxygen saturation. The apnea-hypopnea index (AHI) was defined as the total number of apneas and hypopneas per hour of sleep. All sleep scorers had interrater reliability indexes (κ) >0.86 for staging, arousal, and respiratory parameters. An AHI ≥ 5 was considered diagnosis of OSA. An AHI ≥ 5 and <15 indicated mild OSA, ≥ 15 to <30 indicated moderate OSA, and ≥ 30 indicated severe OSA. All patients completed the Epworth Sleepiness Scale (ESS) [22] at their initial visit.

**Flow Cytometry**

Peripheral venous blood was drawn from an antecubital vein through a 21-gauge needle into a sodium citrate Vacutainer (Becton Dickinson) after the first 2 ml of blood had been discarded. Samples were immediately fixed for 10 min with 1.1% formaldehyde (Polysciences) in 1:4 Hanks balanced saline solution (InVitrogen), then diluted 4.6-fold with distilled water to lyse the erythrocytes. Aliquots (500 µl) of the fixed/lysed blood samples were concentrated by centrifugation (400 g for 5 min), and the resuspended pellet was incubated at 22 °C for 10 min with saturating concentrations of the monoclonal antibodies Y2/S1-FITC (GPIIIa-specific) and MoP9-PE (CD14-specific). Isotype-matched mouse IgG-FITC (Dako) and preinfusion samples containing no biotinylated platelets served as negative controls. The samples were placed on ice until analysis within 4 h by flow cytometry (with low-flow setting) on a FACSCalibur flow cytometer (Becton Dickinson) equipped with a 488-nm argon ion and 635-nm diode lasers, standard four-color filter configuration and CELLQuest cell analysis software (Becton Dickinson). A minimum of 2,000 monocytes was counted per test. Using WinList (Werity Software House, Topsham, Me., USA), monocytes were identified and gated by their brightly positive CD14 expression on a two-parameter dot plot displaying linear orthogonal light scatter vs. MoP9-PE (FL2). The threshold was set on forward angle light scatter to include all leukocytes and exclude debris and uncomplexed platelets. The percentage of platelet-monocyte aggregates was identified in single-parameter histograms of Y2/S1-FITC (FL1) fluorescence displaying events from the monocyte gate (fig. 1). The positive analysis region was determined using an IgG-FITC conjugated isotypic control.
Measurement of Soluble CD40 Ligand Levels

A second venous blood was collected into ethylenediaminetetraacetic acid (EDTA) tubes and centrifuged at 1,500 \( g \) for 15 min at \( 4^\circ C \). The platelet-poor plasma component was stored at \(-80^\circ C\). Plasma sCD40L levels were detected using a commercially available enzyme-linked immunosorbent assay (ELISA) based on the sandwich principle, according to the manufacturer's instructions (sCD40L ELISA test kit; R&D Systems, Minneapolis, Minn., USA). The assay is highly specific for the quantitative determination of sCD40L in humans with a detection limit of 0.095 ng/ml and intra- and interassay coefficients of variation of 6.8 and 14.2%.

nCPAP Therapy

All patients with OSA were treated with nasal autoCPAP (REMstar; Respironics, Inc.). nCPAP compliance was directly measured using compliance software provided by the manufacturer. Good compliance was defined as nCPAP use for at least 4 h per night during 5 days of the week with a reduction of obstructive episodes to \( \text{AHI} < 5 \) and elimination of snoring. At 8 weeks of nCPAP therapy, samples of venous blood were obtained and processed for flow cytometry and for sCD40L quantitation as previously described.

Sample Size

Based on a previous study that showed approximately 50% reduction in sCD40L levels following nCPAP therapy [9], a minimum of 8 pairs of OSA subjects before and after nCPAP therapy was required to achieve a power of 0.8 and an \( \alpha \) of 0.05.

Statistical Analyses

Data are presented as mean (\( \pm \)SD). The distribution of variables was examined by the Kolmogorov-Smirnov Goodness of Fit test. The paired and unpaired Student’s t test and Pearson’s correlation coefficient were used for normally distributed variables; otherwise, the Wilcoxon-Mann-Whitney U test and Spearman’s rank correlation test were applied. Cook’s distance was used to assess for potential outliers. All tests performed were two-sided and \( p < 0.05 \) was considered to indicate statistical significance. Data were analyzed using statistical software (NCSS 2000; Salt Lake City, Utah, USA).

Results

Twelve patients with polysomnographically confirmed OSA and 12 control subjects were included in the study. Table 1 shows the clinical profile of the study population. Six patients had mild to moderate OSA and 6 patients had severe OSA. There were no significant differences in baseline characteristics between OSA patients and healthy controls.
Effect of Sleep Apnea on sCD40L and Platelet-Monocyte Aggregates

Plasma sCD40L levels were significantly higher in patients with OSA compared to nonapneic subjects (7.6 ± 4.3 versus 1.7 ± 1.1; p = 0.004) (fig. 2). Similarly, platelet-monocyte aggregates were increased in subjects who had OSA compared to controls (41.3 ± 23.7 versus 6.7 ± 4.9; p = 0.001) (fig. 3).

Table 2 depicts Spearman’s correlation coefficients between plasma levels of sCD40L, platelet-monocyte aggregates and PSG variables, metabolic profile, and ESS in patients with OSA. There were no correlations between sCD40L levels or platelet-monocyte aggregates and age, gender, BMI, severity of OSA, and baseline objective sleepiness. Nor did glucose and total cholesterol show any relationship with either sCD40L or platelet-monocyte aggregates. Only the percentage of time spent with SpO2
<90% correlated positively with plasma sCD40L levels (r = 0.69; p = 0.01) and platelet-monocyte aggregates (r = 0.64; p = 0.03), respectively. Moreover, levels of sCD40L displayed a significant correlation with platelet-monocyte aggregates (r = 0.67, p = 0.02) (fig. 4).

**Treatment Effect on sCD40L and Platelet-Monocyte Aggregates**

The use of nCPAP ranged from 4 to 7 h per night with an average of 5.3 ± 1.1 h per night. After 8 weeks of autoCPAP therapy, there was no significant change in anthropometric and metabolic profile compared to baseline (table 3). ESS and AH1 were reduced to 3.8 ± 1.4 and 2.3 ± 0.7 per hour, respectively, on successful nCPAP (p < 0.001). The percentage of time spent with SpO2 < 90% also decreased to 2.6 ± 2.3 (p < 0.001). When compared to the data obtained before nCPAP initiation, sCD40L was reduced by 47% (to 4.0 ± 2.9 ng/ml, p = 0.003) and platelet-monocyte aggregates by 42% (to 23.9 ± 20.1%, p = 0.002) (fig. 5). In 7 control subjects without sleep apnea, the use of nasal autoCPAP did not alter the plasma levels of sCD40L or the platelet-monocyte aggregates.

**Discussion**

In the current study, we have demonstrated that increased plasma sCD40L expression occurs in tandem with in vivo platelet activation among subjects with OSA. These derangements were correlated with hypoxic stress.

The 8-week treatment with nCPAP reduced, but failed to normalize, plasma levels of sCD40L and platelet-monocyte aggregates.

Recent interest has focused on the measurement of sCD40L for risk stratification of patients with coronary artery disease or at risk of developing coronary artery disease. Pilot studies have found that individuals with hypercholesterolemia [23] and diabetes [24] have elevated sCD40L levels and that very high levels of sCD40L may identify apparently healthy women at increased risk of having a first adverse cardiovascular event [25]. While a causal relationship is still lacking, the findings of the current study provide further evidence of the association between OSA and cardiovascular diseases by demonstrating higher levels of sCD40L in apneic patients than in healthy controls. The elevated sCD40L levels correlated positively with platelet-monocyte aggregates and were directly related to hypoxic stress in these patients. Similar observations were reported in two recent clinical investi-
Sustained release of platelet sCD40L in subjects with sleep apnea provides further insight into the mechanisms by which sCD40L may contribute to atherothrombotic events in this population. Treatment with nCPAP may help to alleviate this risk.

Consistent with previous studies [9, 26], correction of nocturnal hypoxemia with nCPAP therapy resulted in a reduction in the levels of sCD40L and platelet-monocyte aggregates. However, the mean plasma sCD40L levels following nCPAP therapy did not match those of healthy controls at baseline. Although the sleep indices have improved compared to pre-nCPAP levels, persistent hypoxic events expressed as percent time spent at <90% oxygen saturation were noted in few cases, which could have contributed to higher sCD40L than in normal controls. It is plausible also that the 8-week therapy with nCPAP might be too short to restore normal sCD40L levels in these patients. Alternatively, the higher residual sCD40L levels among subjects with OSA may indicate the presence of an additional sleep-related signal activation of CD40 independent of hypoxic stress. We should point out that platelet-poor EDTA plasma was used for measurement of in vivo sCD40L in our patients. Sample-processing methods can profoundly affect measured levels of sCD40L, with serum yielding levels 3- to 5-fold higher than plasma [40]. Moreover, platelet count influences serum levels of sCD40L, a confounding factor avoided by use of platelet-poor plasma in the present study. Finally, EDTA inhibits the release of CD40L from the platelet surface when administered before platelet activation [41, 42], thus preventing ex vivo release of sCD40L.

A major limitation of the present study is that the effects of nCPAP on plasma sCD40L levels and platelet-monocyte aggregates were not examined with a randomized, placebo-controlled design because of the difficulties of placebo nCPAP treatment. Second, because this is an observational study, we could not establish a causal relationship between OSA and increased levels of platelet-monocyte aggregates or sCD40L. We have attempted, however, to minimize confounding variables by matching the groups for age, gender and BMI.

In conclusion, we have demonstrated increased plasma sCD40L levels with a concomitant rise in platelet-monocyte aggregates in OSA patients. These findings provide insight into the mechanisms by which sCD40L may contribute to atherothrombotic events in this population. Treatment with nCPAP may help to alleviate this risk.

References

CD40 Ligand and Sleep Apnea

Respiration 2009;77:25–31


