Evaluation of the Different Antiphospholipid Antibodies for the Diagnosis of Antiphospholipid Syndrome

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Key Words
Antiphospholipid antibody syndrome • Anticardiolipin antibodies • Anti-\(\beta_2\)-glycoprotein 1 antibodies • Hughes syndrome • Laboratory evaluation

Abstract
Objective: Antiphospholipid syndrome (APS) or Hughes syndrome is a frequently seen condition in Kuwait that is characterised by the presence of significant titres of a variety of antiphospholipid antibodies (aPL). The main clinical features of this syndrome include thrombosis (both venous and arterial), recurrent fetal wastage and thrombocytopenia. The aim of the present study was to find out the antibodies with a high predictive value for the diagnosis of APS. Methodology: The study included a total of 38 subjects. Nine of them were patients with proven primary APS, 10 with secondary APS and 19 were patients (controls) with weak clinical evidence of APS. After a complete clinical examination and routine investigations, the following categories of aPL were estimated by standard laboratory techniques: anticardiolipin antibodies of IgG and IgM isotypes (GPL and MPL, respectively), anti-\(\beta_2\)-glycoprotein-I antibodies (anti-\(\beta_2\)-GPI) of IgG and IgM isotypes and activated partial thromboplastin time as a surrogate for lupus anticoagulant (LAC). The tests were considered positive if the titres were more than a standard cut-off value provided by the manufacturer of the kits, and in one instance, the normal range was established in our laboratory (LAC). Results: The highest sensitivity (78.9%) and diagnostic accuracy (71.1%) were obtained by the estimation of GPL. However, excellent specificity (100%) and positive predictive value (100%) with almost as good a diagnostic accuracy (68.4%) were also obtained with anti-\(\beta_2\)-GPI of IgG isotype.
All the other tests performed poorly when compared with these two. There was no difference in the performance of these tests between primary and secondary APS. **Conclusion:** Of the various aPL estimated, GPL and anti-β2-GPI of IgG isotype were statistically sensitive and specific investigations for confirming the diagnosis of APS.

**Introduction**

Since its original description by Hughes in 1983 as a syndrome of thrombosis, recurrent fetal loss and thrombocytopenia associated with antibodies reactive with anionic phospholipids [1], this syndrome is now purported to be the commonest acquired hypercoagulable state [2]. The condition has been labelled ‘antiphospholipid antibody syndrome’ (APS) [3]. However, because of the recent discovery that antibodies seen in this syndrome may primarily be reactive against several cofactors (proteins) that are naturally associated with anionic phospholipids in the body, the term APS may be inappropriate. Therefore, to overcome the nomenclature problems and to give credit to the worker who first described the syndrome, a strong case has been made to use the eponym ‘Hughes syndrome’ for this condition [4, 5].

An earlier study from this institution had reported that Hughes syndrome was a commonly seen condition in hospitals in Kuwait [6]. Since then, clinical colleagues have frequently approached the rheumatology service for guidance in requisitioning appropriate laboratory investigations for the confirmation of diagnosis of APS and interpretation of their results.

The specific laboratory investigation that has been an integral part of the definition of this syndrome is the presence and persistence of significant titres of antiphospholipid antibodies (aPL) [7, 8]. However, aPL are a heterogeneous ‘family’ of auto-antibodies reactive against different anionic phospholipids and certain additional cofactors that are essential factors in prothrombin activation and other sequences in the cascade of blood coagulation [9]. The wide spectrum of clinical events and the heterogeneous nature of the aPL, therefore, confound the diagnosis of APS [10]. In this regard one of the more perplexing issues is that many infections, certain drugs and other underlying illnesses such as liver disease, malignancy and other acute-phase reactions may also be associated with elevated levels of aPL [2, 7]. However, unlike the aPL associated with systemic lupus erythematosus (SLE) or other auto-immune diseases, infection and drug-associated aPL are usually not associated with clinical features of APS [2, 10]. Thus there remains no uniform laboratory test to identify pathogenic aPL that have clinical relevance [10].

A variety of tests are available for the screening of aPL [11]. These include (i) anticardiolipin antibodies (aCL) and their immunoglobulin isotypes [IgG, IgM (GPL and MPL) and IgA]; (ii) assays for ‘lupus anticoagulant’ (LAC), and (iii) anti-β2-glycoprotein I antibodies (anti-β2-GPI) and their immunoglobulin isotypes [11]. The obvious question with regard to these antibodies is to identify the clinically relevant antibody/antibodies that correlate(s) best with the clinical features of APS [2, 10]. Therefore, in the present study assays for several of these antibodies were carried out in clinically definite cases with primary or secondary APS and the results were compared with those in ‘controls’ with weak clinical evidence of APS. The objective of the study was to find the test(s) with high diagnostic accuracy for the diagnosis of APS.
Patients and Methods

Patient Selection

For this study the following categories of patients were selected from the rheumatology service of the Mubarak Al-Kabeer Hospital, one of the two main teaching hospitals of this institution:

(1) patients with proven diagnosis of primary APS or clinical features highly suggestive of primary APS [7, 8, 12]; this group included 9 patients; (2) patients with a proven diagnosis of SLE: (a) who could be classified as cases of APS with SLE [13]; there were 10 such patients; (b) who were without any clinical evidence of APS; there were 7 such patients; (3) patients with clinical features that are not typical of, but occasionally reported in APS (clinical features that may or may not be a manifestation of this syndrome); there were 12 such patients.

The minimal diagnostic criteria for the diagnosis of APS were as follows [14]: clinical characteristics: venous thrombosis, arterial thrombosis, recurrent fetal loss, thrombocytopenia; laboratory characteristics: IgG aCL (moderate/high levels), IgM aCL (moderate/high levels), positive LAC test. Conditions: patients with the syndrome should have at least 1 clinical + 1 laboratory finding during their disease. The aPL test must be positive on at least 2 occasions more than 3 months apart.

Data Collection

Serum samples were collected over a period of 8 months. All the follow-up patients who attended the rheumatology service and new patients who could be categorised in any of the above 3 categories were selected for this study. Sera were separated on the day of the blood collection and frozen at –70°C till tested (maximum storage time 6 months). Information on the clinical characteristics of the patients, reports of their routine investigations including complete blood counts, serum biochemistry, coagulation profile and specialised investigations that were required to confirm certain features of APS (e.g. duplex Doppler study and/or venography for deep-vein thrombosis, ventilation-perfusion scan for the diagnosis of pulmonary embolism, computed tomography and/or magnetic resonance imaging studies for neurological lesions) were collected.

The following categories of aPL were investigated: (1) aCL: GPL and MPL; (2) anti-ß2-GPI of IgG and IgM isotypes; (3) for the screening of LAC, kaolin clotting time or activated partial thromboplastin time and diluted Russell viper venom time.

Methodology

Testing for aCL. Both GPL and MPL were tested using commercially available enzyme-linked immunosorbent (ELISA) assay kits from Sanofi Diagnostics Pasteur Inc., USA (cat. No. 30969). The instructions provided by the manufacturer were followed in exact details. Results were expressed in GPL and MPL units for IgG and IgM isotypes of aCL, respectively.

Testing for Anti-ß2-GPI. Both the IgG as well as the IgM isotypes of anti-ß2-GPI were tested using commercially available ELISA kits from Varelisa, USA (cat. No. 19096 for the ‘screening’ including all the 3 major immunoglobulin isotypes; cat. No. 18796 for anti-ß2-GPI of IgG isotype and cat. No. 18896 for IgM isotype). The instructions provided by the manufacturer were followed in exact details. Results were expressed in international units for IgG and IgM isotypes of anti-ß2-GPI, respectively.

It is to be noted that these kits conform to the standards laid down in the international workshops on the determination of different aPL [5].

LAC Methodology. The guidelines on testing for LAC as proposed by the SSC subcommittee for standardisation of LAC were followed for the detection of phospholipid-dependent inhibitors of blood coagulation [15]. These included kaolin clotting time or activated partial thromboplastin time with diluted plasma [16] and the diluted Russell viper venom time [17].

Data Analysis

The results were analysed using the Statistical Package for Social Sciences. The reliability of diagnostic tests was assessed using the diagnostic accuracy indices (sensitivity, specificity, positive and negative predictive values, and overall diagnostic accuracy). These indices reflect the extent of false-positive and false-negative results [18].

Results and Observation

Patients

The study included a total of 38 patients: 9 subjects in group 1, 17 in group 2 (10 in group 2a and 7 in group 2b) and 12 in group 3. The activated partial thromboplastin time result was not available for 1 patient from group 3 who had some features suggestive but not confirmatory of APS. Demographic details including median age, age
Table 1. Demographic and clinical features of 38 patients included in the study

<table>
<thead>
<tr>
<th>Patient groups</th>
<th>Age, years</th>
<th>Gender F:M</th>
<th>Clinical manifestations of APS</th>
<th>Haematological</th>
<th>DVT</th>
<th>Recurrent fetal loss</th>
<th>Arterial thrombosis</th>
<th>CNS</th>
<th>VHD</th>
<th>CAPS</th>
</tr>
</thead>
<tbody>
<tr>
<td>Typical primary APS (n = 9)</td>
<td>33.5</td>
<td>7:2</td>
<td>4</td>
<td>1</td>
<td>2</td>
<td>1</td>
<td>3</td>
<td>nil</td>
<td>nil</td>
<td>1</td>
</tr>
<tr>
<td>Systemic lupus erythematosus (n = 17)</td>
<td>33.5</td>
<td>8:4</td>
<td>2</td>
<td>0</td>
<td>nil</td>
<td>nil</td>
<td>nil</td>
<td>nil</td>
<td>nil</td>
<td>nil</td>
</tr>
<tr>
<td>With secondary APS (n = 10)</td>
<td>27.5</td>
<td>17:0</td>
<td>5</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>3</td>
<td>1</td>
<td>5</td>
<td></td>
</tr>
<tr>
<td>Without secondary APS (n = 7)</td>
<td>38</td>
<td>5</td>
<td>nil</td>
<td>nil</td>
<td>nil</td>
<td>nil</td>
<td>nil</td>
<td>nil</td>
<td>nil</td>
<td>nil</td>
</tr>
<tr>
<td>Miscellaneous group with weak evidence of APS (n = 12)</td>
<td>33.5</td>
<td>8:4</td>
<td>2</td>
<td>0</td>
<td>nil</td>
<td>nil</td>
<td>nil</td>
<td>nil</td>
<td>nil</td>
<td>nil</td>
</tr>
</tbody>
</table>

Haematological manifestations = Auto-immune haemolytic anaemia/thrombocytopenia; DVT = deep-vein thrombosis; CNS manifestations = cerebrovascular accidents/ headaches; VHD = valvular heart disease; CAPS = catastrophic APS.

Table 2. Correlation of different tests for aPL with clinical manifestations of APS

<table>
<thead>
<tr>
<th></th>
<th>APTT1 prolonged</th>
<th>Normal</th>
<th>GPL2</th>
<th>MPL3</th>
<th>Anti-ß2-GPI-GPL (IgG)4</th>
<th>Anti-ß2-GPI-MPL (IgM)5</th>
</tr>
</thead>
<tbody>
<tr>
<td>APS present</td>
<td>5</td>
<td>13</td>
<td>15</td>
<td>4</td>
<td>7</td>
<td>12</td>
</tr>
<tr>
<td>APS not present</td>
<td>1</td>
<td>18</td>
<td>7</td>
<td>12</td>
<td>2</td>
<td>17</td>
</tr>
<tr>
<td>Sensitivity, %</td>
<td>27.8</td>
<td>78.9</td>
<td>36.8</td>
<td>36.8</td>
<td>100.0</td>
<td>89.5</td>
</tr>
<tr>
<td>Specificity, %</td>
<td>94.7</td>
<td>63.2</td>
<td>89.5</td>
<td>100.0</td>
<td>77.8</td>
<td>71.4</td>
</tr>
<tr>
<td>PPV, %</td>
<td>83.3</td>
<td>68.2</td>
<td>77.8</td>
<td>100.0</td>
<td>61.3</td>
<td>54.8</td>
</tr>
<tr>
<td>NPV, %</td>
<td>58.1</td>
<td>75.0</td>
<td>58.6</td>
<td>61.3</td>
<td>68.4</td>
<td>57.9</td>
</tr>
<tr>
<td>DA, %</td>
<td>60.5</td>
<td>71.1</td>
<td>63.2</td>
<td>68.4</td>
<td>57.9</td>
<td></td>
</tr>
</tbody>
</table>

PPV = Positive predictive value; NPV = negative predictive value; DA = diagnostic accuracy.

1 Activated partial thromboplastin time as a surrogate for LAC.
2 Anticardiolipin antibody of IgG isotype, in GPL units; considered negative if ≤ 23 GPL units.
3 Anticardiolipin antibody of IgM isotype, in MPL units; considered negative if ≤ 11 MPL units.
4 Anti-ß2-GPI of IgG isotype, in international units; considered negative if ≤ 10 units.
5 Anti-ß2-GPI of IgM isotype, in international units; considered negative if ≤ 10 units.

range, female-to-male ratio and clinical features pertinent to subjects with primary and secondary APS are given in table 1. Of the 12 subjects in the miscellaneous category (group 3, i.e. those with clinically 'weak' evidence for APS) the diagnoses were demyelinating diseases of the central nervous system (2 patients), transverse myelitis (1 patient), cerebrovascular accident in a young woman (1 patient), systemic necrotising vasculitis (2 patients), cutaneous vasculitic syndrome (1 patient), syndrome of haemolytic anaemia.
with elevated liver enzymes and low platelets (1 patient), Sjögren’s syndrome with anaemia suspected to be haemolytic (1 patient), chronic inflammatory arthritides with deep venous thrombosis (2 patients) and Behcet’s disease with deep-vein thrombosis (1 patient). As can be appreciated, each of these patients had at least one clinical feature at the outset that could be considered a manifestation of APS. However, the subsequent course and investigations pointed out alternative diagnoses other than APS.

Analysis of the Results on aPL Estimations
The analysis of the results is given in table 2. As can be seen, the highest sensitivity and diagnostic accuracy were obtained by the estimation of GPL, i.e. 78.9 and 71.1%, respectively. However, the specificity and positive predictive value were relatively lower than those of other tests (63.2 and 68.2%, respectively). The highest specificity and positive predictive value were observed with the estimation of anti-ß2-GPI of IgG isotype. All the other tests performed poorly when compared with these 2 tests.

Discussion
Despite the availability of a number of laboratory tests for the screening of aPL, several workers have emphasised the difficulties in the interpretation of their clinical relevance/significance [2, 5, 19–27]. These and other studies, however, appear to reach a consensus that although generally GPL and LAC show a good correlation among themselves and with the thrombotic manifestations of APS, mainly thrombosis, LAC correlates much more with venous thrombosis [27, 28]. Some studies have shown a correlation of the IgA isotype of aCL with thrombocytopenia [29] while the IgM isotype of aCL has been shown to correlate with auto-immune haemolytic anaemia [30]. Some studies have suggested that aCL correlates better with recurrent fetal wastage, LAC correlates better with thrombotic events [31–33]. The present study showed a good correlation between GPL and LAC (Spearman correlation coefficient 0.6258, 2-tailed significance 0.000), and their good correlation with clinical manifestations of APS. This finding is in agreement with some of the earlier studies [31–33]. Although LAC showed a high positive predictive value for the presence of clinical features of APS, it was less sensitive in comparison with the IgG isotype of aCL. The present study also showed the highest clinical accuracy of GPL that is in agreement with several earlier studies [28, 30, 34–36].

In recent years a large number of studies have shown a high positive predictive value of anti-ß2-GPI for APS [19, 36–42]. The present study showed that of the various aPL available GPL closely followed by the IgG isotype of anti-ß2-GPI has the highest diagnostic accuracy. Other aPL also performed well but did not reach the same high levels of diagnostic accuracy. However, a look at the results would make it obvious that patients with clinically definite APS may have any one but not always both of these antibodies. Therefore, in practical terms, it may be necessary to carry out the tests for both of these antibodies in clinical situations where APS is a clinical possibility. However, it could be recommended that until more specific and more sensitive diagnostic test(s) become(s) available for the laboratory confirmation of APS, it would be prudent to screen for the presence of IgG isotypes of both aCL as well as anti-ß2-GPI. Possibly this would provide a highly satisfactory level of diagnostic accuracy. Other workers have found mainly the IgG isotype of anti-ß2-GPI to be more accurate for the clinical diagnosis of APS than GPL [19, 37, 40]. The difference between the clinical accuracies of

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these 2 antibodies was only marginal in the present study. It is therefore possible that in a large series anti-β₂-GPI may show higher clinical accuracy.

It is to be noted that the field of detection of ‘pathogenic’ versus ‘non-pathogenic’ aPL is still wide open with a number of additional antibodies being claimed as the ‘true’ marker of the clinical APS [25, 43, 44]. It would probably require much more observation and many more studies to reach the final conclusion regarding this perplexing issue. In the meantime, guidelines provided by Stratta et al. [45] may be found extremely helpful in situations where discrepancy is noted between the clinical setting and the laboratory results.

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