Mini Review

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Anti-Müllerian Hormone and Sertoli Cell Function in Paediatric Male Hypogonadism

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Abstract
In the prepubertal male, Sertoli cells are the most active testicular cell population. Without stimulation tests, prepubertal hypogonadism can only be evidenced if Sertoli cell function is assessed. Anti-müllerian hormone (AMH) is a distinctive marker of the prepubertal Sertoli cell. Serum AMH is high from fetal life until puberty. In postnatal life, AMH testicular production is stimulated by FSH and potently inhibited by androgens. In anorchid patients, AMH is undetectable. In prepubertal males with fetal- or childhood-onset primary or central hypogonadism affecting the whole gonad, serum AMH is low. Conversely, when hypogonadism only affects Leydig cells (i.e., LH/human chorionic gonadotrophin receptor or steroidogenic enzyme defects), serum AMH is normal/high. AMH is also normal/high in patients with androgen insensitivity. In patients of pubertal age with central hypogonadism, AMH is low for Tanner stage – reflecting lack of FSH stimulus, – but high for age – reflecting lack of testosterone inhibitory effect. FSH treatment results in serum AMH rise, whereas human chorionic gonadotrophin treatment increases testosterone levels which inhibit AMH production. In conclusion, AMH determination is helpful in assessing gonadal function, without need for stimulation tests, and orientates the aetiological diagnosis of paediatric male hypogonadism. Furthermore, serum AMH is an excellent marker of FSH and androgen action in the testis.

The testes are composed by two different compartments: the seminiferous tubules, containing germ and Sertoli cells, and the interstitial tissue, containing Leydig cells. Testicular function is mainly regulated by pituitary gonadotrophins LH and FSH, whose secretion is induced by the hypothalamic gonadotrophin-releasing hormone (GnRH). The hypothalamic-pituitary-testicular axis shows specific maturational changes in each of its components from fetal life through adulthood. The biological aspects of Sertoli cell function have been recently reviewed [1]. Here, we will address the relevance of Sertoli cell physiology and pathophysiology in the understanding of prepubertal male hypogonadism.
Normal Development of the Hypothalamic-Pituitary-Testicular Axis

Fetal Life
Testis differentiation, by the end of the 5th embryonic week, is characterised by the aggregation of Sertoli and germ cells to form the seminiferous cords. Sertoli cells synthesise anti-müllerian hormone (AMH), responsible for the regression of müllerian ducts, the anlagen of the uterus and fallopian tubes. The initiation of AMH secretion is independent of pituitary function. Afterwards, fetal FSH regulates Sertoli cell activity [1]. Leydig cells differentiate a few days later and secrete testosterone [2] and INSL3 [3], stimulated by placental human chorionic gonadotrophin (hCG). Androgens are responsible, during the first trimester, for the differentiation of the wolffian duct into the epididymis, seminal vesicle, and vas deferens, as well as for the masculinisation of the external genitalia. During the third trimester, LH is the main stimulus for testosterone secretion, responsible for the second phase of testicular descent and for penile growth. INSL3, also regulated by hCG and LH, is the main responsible for the first phase of testicular descent during the second trimester [3].

Neonatal Period, Infancy and Childhood
During the whole prepubertal period – between birth and the onset of puberty – very few changes are observed in the hypothalamic-pituitary-gonadal axis. The seminiferous tubules show no lumen and immature Sertoli cells. Germ cells are represented solely by gonocytes and spermatogonia, which proliferate by mitosis. Typical Leydig cells persist for 3–6 months and then disappear from the interstitial tissue of the testis until puberty. The appearance of external genitalia does not show major changes.

Gonadotrophin secretion, transiently low in the first week of life [4], increases in the second week and remains high during the first 3–6 months of life [5]. LH stimulates Leydig cell testosterone production, while FSH stimulates Sertoli cell proliferation and AMH and inhibin B secretion. Thereafter, gonadotrophins decrease again resulting in a fall of testicular testosterone secretion to very low or undetectable levels during infancy and childhood (fig. 1a). Childhood is therefore a period of physiologic hypogonadotrophic hypoandrogenism. Testicular volume, assessed clinically by palpation and comparison with Prader’s orchidometer – a method with low sensitivity to detect small changes in gonadal size, – also shows no major changes (1–2 ml). For these reasons, the prepubertal testis has been classically described as quiescent. However, when attention is driven to Sertoli cells, it becomes clear that there is an intense activity. The use of accurate methods of evaluation has shown that testis size increases from ap-
proximately 0.5 cm³ in the first year of life to 1.5 cm³ at 10 years [6], mainly due to Sertoli cell proliferation [7] (fig. 1b). AMH, a conspicuous marker of Sertoli cell function, remains high during the whole prepubertal period.

Puberty

The reactivation of the hypothalamic-gonadotrophin axis induces Leydig cells functional activity again. Intratesticular androgen concentration increases before serum testosterone does [8], and provokes Sertoli cell maturation. Sertoli cells stop proliferating, AMH production is down-regulated [9] (fig. 2) and inhibin B secretion increases [10]. Pubertal spermatogenesis is triggered: spermatogonia differentiate into primary spermatocytes, which undergo meiosis and form spermatids that subsequently mature to spermatozoa.

Clinically, the onset of puberty is defined by a testicular volume $\geq 4$ ml. The subsequent increase of gonadal size to a final volume of 15–25 ml is dependent on germ cell proliferation (fig. 1b), which depends on intratesticular androgen levels and normal androgen receptor function in Sertoli cells.

AMH and Sertoli Cell Physiology

AMH Ontogeny

AMH is a distinctive marker of the immature Sertoli cell. Except for a transient decline in the perinatal period, AMH levels remain high from the end of the 7th fetal week until the onset of puberty (table 1) [4, 11, 12]. No extragonadal AMH source has been found. AMH is also produced by the ovaries [12, 13], at very low levels (table 1).

AMH Regulation

Up-Regulation of AMH

Onset of AMH Expression in Fetal Life Is Gonadotrophin Independent. Binding of SOX9 to the AMH promoter is essential for the initiation of AMH expression in...

Fig. 2. Relationship between serum testosterone, androgen receptor (AR) expression in Sertoli cells and serum AMH from birth to puberty. AR expression in Sertoli cells is represented as the percentage of Sertoli cells with AR immunolabelling [data from 28, 45].

Fig. 3. FSH regulation of testicular AMH secretion. FSH binds to the FSH receptor (FSH-R) present on Sertoli cell membrane, and triggers protein G-mediated activation of adenylyl cyclase (AC), resulting in an increase of cyclic AMP (cAMP) levels and activation of protein kinase A (PKA). PKA induces Sertoli cell proliferation and also nuclear translocation of transcription factors AP2 and NFκB, which bind to specific response elements on the distal AMH promoter, enhancing AMH expression [data from 22].
early fetal development [14, 15], independently from FSH action. SF1 [14, 16, 17], GATA4 [17, 18] and WT1 [19] further enhance AMH transcription.

**FSH Increases Basal AMH Testicular Production.** Although basal AMH expression is FSH independent, serum AMH is low in patients with congenital hypogonadotropic hypogonadism, and increases in response to exogenous FSH administration [20, 21], indicating that serum AMH is a useful marker of FSH action on the prepubertal testis. Experimental models have shown that lack of FSH stimulation during fetal and neonatal life results in low AMH owing to a decrease in both Sertoli cell number and AMH expression. Low serum AMH correlated with small testes [22]. Both in mice [23] and humans [20], FSH administration results in an increased testis volume and in AMH transcriptional activity through the classical FSH receptor transduction pathway involving Gsα protein, adenylyl cyclase and protein kinase A (fig. 3), with subsequent nuclear translocation of transcription factors NFκB and AP2, which bind to specific response elements in the AMH promoter [22].

**Inhibition of AMH**

**Androgens Are Potent Inhibitors of AMH Production in Sertoli Cells.** AMH decreases during puberty, mainly between Tanner stages II and III [24], in coincidence with the increase of intratesticular testosterone concentration. This negative correlation between testosterone and AMH is observed also in central precocious puberty and in gonadotrophin-independent precocious puberty (testotoxicosis), suggesting that androgens are responsible for AMH down-regulation independently of age and of gonadotrophin levels [24]. Furthermore, in patients with constitutional delay of puberty [25] or with defective androgen production or sensitivity [26, 27], AMH levels remain at high prepubertal levels. The inhibitory effect of androgens prevails over the positive effect of FSH on AMH secretion in the pubertal testis (fig. 4).

The surprising coexistence of high levels of androgens and AMH during fetal life and the first months after birth is explained by the physiological androgen insensitivity of Sertoli cells in these periods, owing to the lack of androgen receptor expression in Sertoli cell nuclei (fig. 2, 4) [23, 28–30].

**Meiosis Is Also Associated with AMH Down-Regulation.** During human puberty, AMH expression is significantly down-regulated in the seminiferous tubules after meiotic onset [31]. Experimental observations in XXsxr<sup>4</sup> male mice, a model for the human XX male, indicate that meiotic germ cells may play a role in the inhibition of AMH expression [23].

<table>
<thead>
<tr>
<th>Age group</th>
<th>Serum AMH</th>
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<tbody>
<tr>
<td></td>
<td>pmol/l</td>
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<tr>
<td></td>
<td>males</td>
</tr>
<tr>
<td>2 days</td>
<td>371 ± 168</td>
</tr>
<tr>
<td>7 days</td>
<td>397 ± 152</td>
</tr>
<tr>
<td>10 days</td>
<td>427 ± 127</td>
</tr>
<tr>
<td>20 days</td>
<td>523 ± 156</td>
</tr>
<tr>
<td>1 month–0.9 years</td>
<td>794 ± 264</td>
</tr>
<tr>
<td>1–4.9 years</td>
<td>699 ± 245</td>
</tr>
<tr>
<td>5–12 years:</td>
<td>516 ± 275</td>
</tr>
<tr>
<td>Tanner I &gt;10 years:</td>
<td>249 ± 126</td>
</tr>
<tr>
<td>Tanner II III</td>
<td>98 ± 65</td>
</tr>
<tr>
<td>IV–V/adults</td>
<td>42 ± 38</td>
</tr>
</tbody>
</table>

Table 1. Serum AMH in normal males and females, using the AMH/MIS ELISA from Beckman-Coulter-Immunotech [data from 13, 45]
In the prepubertal male, Sertoli cells are the most active testicular cell population, while testosterone and sperm production are physiologically inexistent. Therefore, male hypogonadism in the boy can only be discovered if Sertoli cell function is assessed. A more comprehensive clinical definition of hypogonadism includes decreased testicular function involving a disorder of gamete production or function, Leydig cell hormone production (androgens and INSL3) and/or Sertoli cell hormone production (AMH and inhibin B).

**Classification of Hypogonadism (table 2)**

**Testicular (Primary), Hypothalamic-Pituitary (Central) or Combined**

Male hypogonadism can be classified according to the level of the hypothalamic-pituitary-gonadal axis that is primarily affected. In central hypogonadism, gonadal failure results from a malfunction of the hypothalamic GnRH pulse generator or the pituitary gonadotroph to secrete LH and FSH. In primary hypogonad-
### Table 2. Classification of hypogonadism

<table>
<thead>
<tr>
<th>Classification</th>
<th>Whole testicular dysfunction</th>
<th>Cell-specific testicular dysfunction</th>
</tr>
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<tbody>
<tr>
<td><strong>Central hypogonadism</strong>&lt;br&gt;Fetal-onset (1st trimester)</td>
<td>Inexistent</td>
<td>Inexistent</td>
</tr>
<tr>
<td>Fetal-onset (2nd–3rd trimesters)</td>
<td>Isolated hypogonadotrophic hypogonadism&lt;br&gt;Gene mutations: GnRH-R, KAL1, FGFR1, GRP54, Leptin/Leptin-R&lt;br&gt;Multiple pituitary hormone deficiency&lt;br&gt;Gene mutations: HESX1, LHX3, PROP1, OTX2&lt;br&gt;Other causes of septo-optic dysplasia and defects of mid-line</td>
<td>Leydig cells&lt;br&gt;Isolated LH deficiency&lt;br&gt;(LHβ-subunit gene mutation)&lt;br&gt;Sertoli cells&lt;br&gt;Isolated FSH deficiency&lt;br&gt;(FSHβ-subunit gene mutation)</td>
</tr>
<tr>
<td>Childhood-onset</td>
<td>Isolated hypogonadotrophic hypogonadism&lt;br&gt;Endocrine disorders: hypothyroidism, hyperprolactinemia&lt;br&gt;Multiple pituitary hormone deficiency&lt;br&gt;Central nervous system tumours, Langerhans’ histiocytosis</td>
<td>Inexistent</td>
</tr>
<tr>
<td>Peripubertal-onset</td>
<td>Isolated hypogonadotrophic hypogonadism&lt;br&gt;Functional central hypogonadism: anorexia nervosa&lt;br&gt;Endocrine disorders: hypothyroidism, hyperprolactinemia&lt;br&gt;Multiple pituitary hormone deficiency&lt;br&gt;Central nervous system tumours, Langerhans’ histiocytosis</td>
<td>Inexistent</td>
</tr>
<tr>
<td><strong>Primary hypogonadism</strong>&lt;br&gt;Fetal-onset (1st trimester)</td>
<td>Gonadal dysgenesis&lt;br&gt;Gene mutations: SRY, SOX9, ATRX, WT1&lt;br&gt;Chromosomal abnormalities: 9p deletion, DSS (Xq) duplication, 45,X/46,XY</td>
<td>Leydig cell&lt;br&gt;Leydig cell aplasia/hypoplasia: LH/hCG-R mutation&lt;br&gt;Steroidogenic protein defects: STAR, P450sc, P450c17, 17β-HSD, 17β-HSD&lt;br&gt;Sertoli cell&lt;br&gt;AIS: AR mutation&lt;br&gt;PMDS: AMH mutation</td>
</tr>
<tr>
<td>Fetal-onset (2nd–3rd trimesters)</td>
<td>Testicular dysgenesis syndrome&lt;br&gt;Endocrine disruptors&lt;br&gt;Testicular regression syndrome&lt;br&gt;Testicular torsion&lt;br&gt;Mild gonadal dysgenesis</td>
<td>Sertoli cell&lt;br&gt;FSH-R mutation</td>
</tr>
<tr>
<td>Childhood-onset</td>
<td>Cryptorchidism&lt;br&gt;Testicular torsion&lt;br&gt;Orchitis&lt;br&gt;Chemotherapy/radiotherapy&lt;br&gt;Klinefelter syndrome (germ cell depletion)</td>
<td>Inexistent</td>
</tr>
<tr>
<td>Peripubertal-onset</td>
<td>Cryptorchidism&lt;br&gt;Testicular torsion&lt;br&gt;Orchitis&lt;br&gt;Chemotherapy/radiotherapy&lt;br&gt;Klinefelter syndrome (Sertoli and Leydig cell dysfunction)</td>
<td>Germ cells&lt;br&gt;XX male</td>
</tr>
<tr>
<td><strong>Combined hypogonadism</strong>&lt;br&gt;Fetal-onset (1st trimester)</td>
<td>Gene mutations: DAX1, SF1</td>
<td>Inexistent</td>
</tr>
<tr>
<td>Fetal-onset (2nd–3rd trimesters)</td>
<td>Prader-Willi syndrome</td>
<td>Inexistent</td>
</tr>
<tr>
<td>Childhood-onset</td>
<td>Total body irradiation</td>
<td>Inexistent</td>
</tr>
<tr>
<td>Peripubertal-onset</td>
<td>Total body irradiation</td>
<td>Inexistent</td>
</tr>
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</table>
ism, a primary testicular failure exists. Certain disorders may impair both the hypothalamic-pituitary axis and the testis concomitantly, and provoke a combined hypogonadism.

**Whole or Cell-Specific Testicular Dysfunction**

Primary, central or combined hypogonadism may reflect the concomitant impairment of all testicular cell populations; in this case, there is a ‘whole testicular dysfunction or hypogonadism’. Alternatively, only one of the testicular cell populations (germ, Sertoli or Leydig) may be primarily affected; in this case, there is a ‘dissociated or cell-specific testicular failure or hypogonadism’.

**Fetal-, Childhood-, Puberty-, Adulthood-, or Late-Onset Hypogonadism**

The clinical consequences of male hypogonadism depend on the period of life in which it is established. Fetal-onset hypogonadism in the first trimester results in disorders of sex development (DSD) presenting with ambiguous or female genitalia, owing to the lack or to insufficient levels of testis hormone secretion during the critical window of male sex differentiation. It should be noted that in a patient carrying a Y chromosome, DSD due to hypogonadism are always primary gonadal dysfunctions: gonadal dysgenesis represents a whole gonadal dysfunction, whereas Leydig cell aplasia/hypoplasia and steroidogenic defects are cell-specific gonadal forms of hypogonadism. Central hypogonadism cannot result in genital ambiguity, since Leydig cell function in the first trimester of fetal life is dependent on hCG and not on fetal pituitary LH. If established during the second or third trimester, fetal hypogonadism – which may be primary, central or combined – results in micropenis and/or cryptorchidism. When established during infancy or childhood, hypogonadism may remain undiagnosed until the age of puberty – unless Sertoli cell function is assessed. At pubertal age, hypogonadism can provoke lack or arrest of pubertal development. During adulthood and senescence, hypogonadism leads to infertility and/or impaired androgen-regulated physiological functions (impaired sexual activity, body composition, bone mineral density, etc.).

**AMH Levels in Prepubertal Male Hypogonadism**

Because gonadotrophin and testosterone levels normally decrease to very low values until the onset of puberty, and only Sertoli cells remain active during childhood, the evaluation of gonadal function in the prepubertal male relies mainly on the assessment of Sertoli cell products [32–35]. In the male, AMH is secreted exclusively by Sertoli cells; therefore, serum AMH reliably reflects the presence and function of testes in prepubertal boys, without the need for any stimulation tests [33, 36, 37].

In this section, we shall address – based on an aetiological approach – how the different disorders causing hypogonadism affect AMH testicular production.

**Central Hypogonadism with ‘Whole Testicular Dysfunction’**

**Isolated Gonadotrophin Deficiency**

Low serum AMH levels have been reported in infants with congenital central hypogonadism. Early postnatal treatment with recombinant human FSH (rhFSH) mimicked the physiological development occurring after birth, resulting in an increase of testicular size – presumably by FSH-induced Sertoli cell proliferation – and an elevation in serum AMH [20].

In untreated patients of pubertal age with congenital hypogonadotrophic hypogonadism, serum AMH is elevated for age – because serum testosterone remains at prepubertal levels and does not down-regulate AMH – but lower than expected for patient’s Tanner stage and serum testosterone [21, 38] – reflecting the lack of FSH stimulus (fig. 4; table 3). Treatment with rhFSH increases serum AMH; further treatment with hCG, resulting in androgen production locally by the testes, mimics the physiological inhibition of AMH observed at puberty (fig. 5). Conversely, inhibition of AMH is less significant when patients are supplemented with exogenous testosterone, which probably indicates that intratesticular testosterone reaches much lower levels [21].

**Multiple Pituitary Hormone Deficiency**

Although there are no published studies on AMH levels in patients with multiple pituitary hormone deficiency, serum AMH might be lower than in patients with isolated hypogonadotrophic hypogonadism. In fact, in experimental conditions GH regulates Sertoli cell function [39] and IGF-I and IGF-II regulate testicular interstitial cell steroidogenesis, proliferation and apoptosis [40].
Central Hypogonadism with ‘Cell-Specific Testicular Dysfunction’

**Central Hypogonadism with Sertoli Cell-Specific Dysfunction**

Although not yet reported, low serum AMH could be expected in patients with isolated FSH deficiency due to FSHβ-subunit gene mutations [41]. Transgenic mice carrying a deletion of the FSHβ-subunit gene have low serum AMH, in correlation with reduced Sertoli cell number and testicular volume. All parameters resumed normal values after FSH administration [22].

**Central Hypogonadism with Leydig Cell-Specific Dysfunction**

Isolated LH deficiency, due to mutations of the LHβ-subunit gene, results in failure of Leydig cell differentiation and testosterone secretion in late fetal life and pubertal age [42]. In mice, FSH levels are usually elevated and Sertoli cells remain immature, producing high AMH levels [43].

**Primary Hypogonadism with ‘Whole Testicular Dysfunction’**

**Complete Gonadal Dysgenesis, Vanishing Testes and Anorchia**

When due to early-onset complete gonadal dysgenesis, the absence of testicular tissue results in a DSD with female external genitalia, whereas vanishing or regression of testicular tissue occurring later in fetal life is associated with male genitalia, micropenis and hypoplastic scrotum. In all cases, the absence of gonadal tissue results in undetectable serum AMH [27, 36, 37].

**Partial Testicular Dysgenisis**

Early-onset partial testicular dysgenesis in XY patients usually results in DSD with ambiguous genitalia. Both tubular and interstitial compartments of the testes are affected producing insufficient AMH and testosterone [27, 44]. Serum AMH is low and usually correlates with the amount of functional Sertoli cells [31]. In most boys with

Table 3. Expected levels of gonadotrophins, testosterone and AMH in normal male development and in disorders affecting AMH production

<table>
<thead>
<tr>
<th></th>
<th>FSH</th>
<th>LH</th>
<th>AMH</th>
<th>Testosterone</th>
</tr>
</thead>
<tbody>
<tr>
<td>Normal fetus and postnatal 0–6 months</td>
<td>high</td>
<td>high</td>
<td>high</td>
<td>high</td>
</tr>
<tr>
<td>Normal childhood</td>
<td>low</td>
<td>low</td>
<td>high</td>
<td>low</td>
</tr>
<tr>
<td>Normal puberty</td>
<td>high</td>
<td>high</td>
<td>very low</td>
<td>high</td>
</tr>
<tr>
<td>AIS (puberty)</td>
<td>high</td>
<td>high</td>
<td>high</td>
<td>high</td>
</tr>
<tr>
<td>Central hypogonadism</td>
<td>very low</td>
<td>very low</td>
<td>low</td>
<td>very low</td>
</tr>
<tr>
<td>Leydig cell aplasia/T-synthesis defects</td>
<td>high</td>
<td>high</td>
<td>high</td>
<td>very low</td>
</tr>
<tr>
<td>Precocious puberty</td>
<td>high</td>
<td>high</td>
<td>very low</td>
<td>high</td>
</tr>
</tbody>
</table>

Fig. 5. Effect of recombinant human FSH (rhFSH) and hCG treatment on testicular AMH production by the testis in patients with previously untreated central hypogonadism. Initial treatment with rhFSH during 30 days resulted in an elevation of serum AMH in all 8 patients, while testosterone (T) remained at prepubertal levels. The shaded area represents normal AMH for Tanner I stage, according to T levels observed in these patients. Subsequent addition of hCG treatment resulted in an elevation of T which provoked a decline in serum AMH. The shaded area represents AMH values for Tanner IV–V stages, according to T levels observed in the treated patients. *, ** and *** indicate p < 0.05, p < 0.01 and p < 0.001, respectively [data from 21].
isolated hypospadias, AMH and testosterone are normal, indicating that there is no testicular dysfunction [45].

Late-onset testicular dysgenesis is a typical feature of Klinefelter syndrome. No overt signs of hypogonadism are evident before puberty: AMH levels are normal, in correlation with normal inhibin B and FSH. At puberty, androgens provoke a physiological decrease in serum AMH. However, Sertoli cell function deteriorates progressively from mid-puberty, resulting in extremely low or undetectable AMH, in coincidence with undetectable inhibin B, very high FSH levels and small testis volume [46].

Cryptorchidism

Cryptorchidism is a clinical sign with many possible aetiologies. It may be one component of the 'testicular dysgenesis syndrome', a primary hypogonadal disorder with 'whole testicular dysfunction' also presenting with hypospadias, as well as reduced semen quality and increased risk of testicular cancer in adults [47]. Alternatively, cryptorchidism may result from anatomical defects of the inguinal region or the abdominal wall (i.e., not associated with hypogonadism), from hypogonadotrophic hypogonadism or from rare mutations of INSL3 or its receptor [3]. According to its aetiology, cryptorchidism may be associated with normal or impaired Sertoli cell function. In boys with bilateral cryptorchidism, AMH is low in approximately 75% of those with non-palpable gonads and 35% of those with inguinal gonads, indicating Sertoli cell dysfunction [48]. AMH is expected to be normal, unless longstanding abnormal testis position occurs, in patients with abdominal wall or inguinal region defects or with impaired INSL3 signalling. As already discussed, AMH is low in patients with cryptorchidism due to congenital hypogonadotrophic hypogonadism [20, 21, 38].

Chemotherapy/Radiotherapy

Chemotherapy and radiotherapy affect primarily germ cells [49], but Sertoli cells may also be affected as shown by studies reporting low inhibin B/FSH ratios [50]. No studies have reported AMH values.

Primary Hypogonadism with 'Cell-Specific Testicular Dysfunction'

Primary Hypogonadism with Sertoli Cell-Specific Dysfunction

Inactivating mutations of the FSH receptor provoke Sertoli cell dysfunction resulting in low Sertoli and germ cell numbers leading to reduced testis volume, moderately elevated FSH and low inhibin B [51]. Low AMH could be expected in paediatric patients, but no reports exist in the literature. Since basal AMH secretion in fetal life is independent from FSH, müllerian ducts normally regress in these patients.

Inactivating mutations of the AMH gene, responsible for the AMH-negative form of the persistent müllerian duct syndrome (PMDS), result in extremely low or most often undetectable levels of serum AMH. These cases should be distinguished from the AMH-positive form of PMDS due to AMH receptor mutations, where serum AMH is normal, indicating that no testicular dysfunction exists [reviewed in 52].

In the androgen insensitivity syndrome (AIS) due to mutations in the androgen receptor gene, attention is usually driven to the lack of virilisation of genitalia. Nevertheless, testicular function is also partially affected. Indeed, Sertoli cells are unable to respond to androgens normally. In the complete form of AIS, FSH does not increase in the first months after birth and AMH remains in the normal male levels [53]. At pubertal age, if the patient has not yet been gonadectomised, FSH increases leading to very high levels of AMH, which cannot be inhibited by testosterone [26, 27] (fig. 4; table 3). In the partial form of AIS, the early postnatal FSH surge occurs normally driving to higher AMH levels [53]. At pubertal age, the increase in testosterone may partially down-regulate AMH: serum AMH declines but remains higher than expected for patient’s testosterone level [26, 27].

Primary Hypogonadism with Leydig Cell-Specific Dysfunction

Inactivating mutations of the LH/hCG receptor or of the steroidogenic enzymes in 46,XY individuals result in DSD due to isolated hypoandrogenism leading to the development of female or ambiguous external genitalia. At variance with what is observed in whole gonadal dysfunction (i.e., testicular dysgenesis), serum AMH is normal/high, mainly during the first year of life and at pubertal age, owing to the lack of androgen inhibitory effect (fig. 4; table 3) [27].

Primary Hypogonadism with Germ Cell-Specific Dysfunction

In XX males, AMH and testosterone are normal, indicating that the somatic cells of the gonads are not primarily affected [27]. However, germ cells fail to progress through gametogenesis and enter apoptosis at puberty. AMH levels have not been evaluated in other disorders.
where germ cells are the primarily affected population, like DAZ (deleted in azoospermia) microdeletions of the Y chromosome.

Combined Hypogonadism (Central and Primary) with ‘Whole Testicular Dysfunction’

DAX1 and SF1 are genes expressed at several levels of the hypothalamic-pituitary-gonadal and adrenal axes [54]. Mutations in the DAX1 gene cause an X-linked disorder presenting with adrenal hypoplasia congenita and combined hypothalamic-pituitary-testicular defects. The latter is characterised by a primary testicular defect, as observed in DAX1 knockout mice [55], and lack of elevation of gonadotrophins, suggestive of an impaired functional capacity of the pituitary. Serum AMH is low, like inhibin B, from early pubertal stages onwards, indicating the existence of a primary failure of testicular function [56].

The phenotypic spectrum associated with SF1 mutations includes different degrees of insufficient virilisation with or without müllerian duct remnant and normal or impaired adrenal function. Endocrine investigations are consistent with gonadal dysgenesis associated with a partial form of hypogonadotrophic hypogonadism [57]. AMH and testosterone are very low, probably due to a multifactorial pathophysiology including gonadal dysgenesis, lack of SFI as a transcription factor for AMH gene expression and insufficient gonadotrophin secretion.

In prepubertal male patients with Prader-Willi syndrome, a specific form of combined central and peripheral hypogonadism, AMH is normal/low, inhibin B is low, FSH is normal/height, and LH and testosterone are normal/low [58–60]. Sertoli cell function has not been assessed either in children exposed to total body irradiation during cancer treatment.

Concluding Remarks

Hypogonadism can be overlooked in infancy and childhood if attention is driven only to gonadotrophins and testosterone. Sertoli cells are the main active population between birth and pubertal onset. Small testes in this period reflect Sertoli cell defects, either primary or secondary to FSH deficiency. Sertoli cell markers, like serum AMH and inhibin B, are most helpful in the evaluation of gonadal function, without any need for stimulation tests, and orientate the aetiological diagnosis of male hypogonadism in paediatric patients. Serum AMH is an excellent marker of FSH and androgen action in the testis.

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References


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