A Nonsense Mutation in the *SCN9A* Gene in Congenital Insensitivity to Pain

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Key Words

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Abstract

Background: Congenital insensitivity to pain (CIP) (OMIM 243000) is a rare autosomal-recessive disorder. Clinically, CIP is characterized by insensitivity to all modalities of pain except neuropathic pain, and recurrent injuries frequently go unnoticed. CIP is caused by mutations in the *SCN9A* gene encoding for the Na1.7 channel. **Methods:** We analyzed the DNA from members of a consanguineous Pakistani family for mutations in the *SCN9A* gene through direct sequencing after performing linkage studies. **Results:** We identified a novel missense mutation designated R523X in all affected individuals. A screening assay ruled out the possibility of polymorphism. **Conclusion:** We identified a novel mutation in the Na1.7 channel leading to CIP, extending the spectrum of mutations in the Na1.7 channel, and enhancing our understanding of the physiology of pain.

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Introduction

Pain is a highly conserved sensory modality among all vertebrate species which confers protection against noxious and dangerous stimuli. Loss of pain sensation is associated with multiple injuries that compromise survival. On the other hand, hypersensitivity to pain lies on the other extreme of the spectrum and would also adversely affect individuals, as they would perceive several non-harmful stimuli as being noxious. Pain is categorized into several subtypes, including mechanical, visceral, inflammatory, thermal, chemical and neuropathic.

Voltage-gated sodium channels are the channels responsible for the conversion of mechanical or chemical stimuli into electrical signals within excited cells [1]. These channels are composed of functional pore-forming α subunits and auxiliary β subunits [2]. To date, nine different sodium channels encoded by nine different genes designated *SCN1*–*SCN9A* have been identified. Inactivation of *SCN1A* and *SCN2A* [3] genes have been associated with epilepsy, *SCN4A* is associated with myotonia disorders [4], *SCN5A* is associated with Brugada syndrome (OMIM 601144) [5], which is characterized by cardiac arrhythmias in young people, and *SCN8A* is associated with cerebellar atrophy and ataxia [6]. *SCN9A* is

associated with indifference to pain in cases with inactivating mutations [7], and the opposite phenotype, hyperalgesia, in cases of activating mutations [8, 9].

Nav1.7, a tetrodotoxin-sensitive sodium channel encoded by the *SCN9A* gene, and Nav1.8, a tetrodotoxin-insensitive sodium channel, are both expressed in the peripheral nervous system and mainly in the dorsal root ganglia and are closely interconnected. They are implicated in pain perception including inflammatory and mechanical pain [10], but not neuropathic pain [11]. Despite the discovery of these two channels and their modes of action, the pathophysiology of pain perception is incompletely understood. To date, mutations in Nav1.7 (OMIM 603415) have been implicated in three pain-related syndromes in humans.

Primary erythromelalgia (PE) (OMIM 133020) and paroxysmal extreme pain disorder (PEPD) (OMIM 167400) are two autosomal-dominant disorders characterized by severe pain. PE involves primarily the hands and feet and to a lesser extent the ears and nose. It usually presents in the first decade of life with severe episodic erythema, pain and flushing that respond well to cooling; symptoms worsen with age [12, 13]. PEPD is characterized by chronic episodic pain attacks starting soon after birth and involving the rectum with radiation to the lower extremities bilaterally triggered mainly by defecation, accompanied by ocular and mandibular pain. Unlike attacks of PE, these attacks are responsive to carbamazepine [9]. Symptoms usually improve with age. Pain in the two syndromes is of the inflammatory type, and is caused by activating mutations in the *SCN9A* gene [14, 15].

In contrast to PE and PEPD, congenital insensitivity to pain (CIP) (OMIM 243000) is an autosomal-recessive disease caused by inactivating mutations in the Nav1.7 channel [7]. Patients present during the first decade of life with lack of pain perception, but with preservation of all other sensory modalities including normal touch perception, proprioception, itch, warm and cold temperature discrimination and vibration. Individuals with CIP generally present with injuries involving the lips and tongue due to asymptomatic injuries during the first few years of life [7, 16]. Bone fractures are also asymptomatic and affected individuals are not aware of these fractures until a limp develops. An accompanying feature among several patients with CIP is anosmia or hyposmia [16, 17].

To date, only a limited number of inactivating mutations in the *SCN9A* gene have been found, among which the majority were nonsense mutations resulting in CIP. Here we present a family from Pakistan with CIP and a novel mutation in the *SCN9A* gene, further extending

the spectrum of mutations in this gene and providing a framework for the better understanding of pain physiology.

Materials and Methods

DNA Extraction

After obtaining informed consent, we collected peripheral blood samples from family members and 100 population-matched unrelated healthy control individuals in EDTA-containing tubes under institutional approval and in adherence to the Declaration of Helsinki Principles. Genomic DNA was isolated from these samples according to standard techniques.

Haplotype Analysis

In order to determine if the phenotype of patients linked to the *SCN9A* gene, we performed haplotype analysis using two microsatellite markers, D2S1379 and D2S2330, closely linked to the gene. The amplification conditions for each PCR were 94°C for 2 min, followed by 35 cycles of 94°C for 30 s, 55°C for 30 s, and 72°C for 30 s, with a final extension at 72°C for 7 min. The amplified PCR products were run on an 8% polyacrylamide gel and genotyping was done by visual inspection.

Mutation Analysis

All exons of the *SCN9A* gene with adjacent sequences of exon-intron borders were amplified by PCR with primers and conditions described previously [7]. The amplified PCR products were directly sequenced in an ABI Prism 310 Automated Sequencer, using the ABI Prism Big Dye Terminator Cycle Sequencing Ready Reaction Kit (PE Applied Biosystems).

Screening Assays

We performed a PCR and restriction endonuclease digestion assay for the novel mutation. We amplified exon 10 of the *SCN9A* gene using the same primers for mutation analysis. This was followed by *Cac8I* digestion at 37°C for 2 h. The product fragments were separated on 2% agarose gel.

Results

Patients

The pattern of inheritance of CIP in this family was autosomal-recessive (fig. 1a). By history, patients reported having experienced no pain from trauma including fractures, burns, and accidental tongue or lip biting, with some individuals losing the distal parts of the tongue (fig. 2a). In addition, none reported having abdominal or chest pain events. All patients reported the ability to discriminate among different odors. On physical examination, all patients had injuries over the lips and tongues. Neurological examination was consistent with defective perception of pain stimuli, while the other sensory mo-

Fig. 1. a Pakistani family showing an autosomal-recessive pattern of inheritance for CIP. Circles indicate females, squares indicate males, filled squares and circles indicate affected individuals and double lines indicate consanguinity. **b** Affected individual with part of her tongue missing.

Fig. 2. a Affected individuals (II-5, III-1, III-2, III-3, III-4, and III-5) show a homozygous mutation at position 1567C>T leading to substitution of arginine by a stop codon. Carrier individuals from the same family (II-4, II-6, II-7, and III-7) are heterozygous for the mutation. Control individuals are homozygous for the wild-type allele. **b** The restriction enzyme *Cac8I* was used for the screening assay for the mutation R523X. The enzyme is expected to digest the wild-type allele into two fragments, while the mutant allele is resistant to the digestion. The smaller fragment is not seen.

dalities including touch, tickle and pressure, warm and cold temperatures, and proprioception were within normal limits. Motor examination and cranial nerve examination including the olfactory nerve were normal.

Genetic Analysis

Haplotype analysis revealed phenotype linkage to the region on chromosome 2 spanning the *SCN9A* gene

(fig. 1). A homozygous nonsense mutation in exon 10 of the *SCN9A* gene was identified at position 1567 designated R523X and was found in the homozygous state among all affected individuals (fig. 2a). Unaffected individuals were either heterozygous for the mutation or carried the wild-type allele. Screening assays performed in 100 control individuals (200 chromosomes) from the same population ruled out the possibility of polymorphism (fig. 2b).

Discussion

The molecular pathways controlling pain perception are highly complex. Given this complex neural circuitry, it is surprising that an alteration in a single sodium channel (Na1.7) can have such profound effects leading to loss of all pain sensation (with the exception of neuropathic pain) when deactivated. This has great implications, mainly at the pharmacological level, as finding antagonists for Na1.7 would have great impact in the field of analgesia.

Na1.7 channels function mainly as gatekeeper channels [18], such that they amplify small signals and bring up neurons to certain voltages, which in turn activate the Na1.8 channels that have a much higher threshold for activation. Na1.8 channels then produce most of the current necessary for the generation of action potentials and therefore transduction of pain [19]. The pathway of pain mediated through both channels is most likely to be acting at the level of peripheral nociceptive transmission since these channels are mainly expressed in the peripheral nervous system, although a central nervous system contribution cannot be entirely ruled out, as there is some expression of Na1.7 and Na1.8 channels at these sites [20].

In cases of PE and PEPD, the mutations detected in the Na1.7 are missense mutations and result in lowering of the threshold of the channels to be activated by small depolarization events, thus enhancing the activity of the channels [21]. Moreover, these mutations slow the deactivation of the channels, keeping them open for an increased duration and in addition there are exaggerated sodium currents in the mutated channels [21, 22]. All these factors contribute to the pathogenesis of PE and PEPD, although it is not yet clear why certain regions in the body are more predisposed to pain development and why pain is paroxysmal [23]. It is also unclear why patients with PE respond well to cooling, which renders the threshold for pain comparable to people with wild-type Na1.7 channels.

CIP is a rare autosomal-recessive condition first described in patients from Pakistan presenting with insensitivity to all modalities of pain including visceral pain. CIP presents immediately after birth and is characterized mainly by insensitivity to pain which is the common feature among all affected individuals, although clinical heterogeneity exists, as some patients develop anosmia while others have intact smell sensation. Although the Na1.7 channels are expressed in the sympathetic nervous system, there is no autonomic nervous system instability, suggesting that Na1.7 channels have secondary roles at these sites or are compensated for by other channels.

The main differential diagnosis that should be considered is hereditary sensory neuropathy type 2 (HSAN2) (OMIM 201300), hereditary sensory neuropathy type 4 (HSAN4) (OMIM 256800) and hereditary sensory neuropathy type 5 (HSAN5) (OMIM 608654). Clinical features in these syndromes in addition to the loss of pain sensation include thermal and tactile abnormalities in addition to loss of neuropathic pain and autonomic nervous system instability [24]. Biopsies of the sural nerve in patients with these syndromes reveal loss of large and small myelinated fibers with some loss in unmyelinated fibers [25], in contrast to patients with CIP in whom the nerves are intact.

The SCN9A gene which encodes for Na1.7 channels is linked to chromosome 2q24. The protein is composed of 1,977 amino acids which form four similar domains. Each domain is composed of six transmembrane regions, with the channel pore being formed from the fifth and sixth transmembrane regions and the voltage sensor located in the fourth transmembrane segment of each domain. The domains are connected to each other by linkers that attach the sixth transmembrane segment of one domain to the first transmembrane segment of the next domain [7, 25].

All mutations in the cases of CIP are homozygous nonsense [7, 17] mutations, except for one compound heterozygous mutation [16]. These mutations are expected to result in either nonsense-mediated mRNA decay with complete absence of channel formation, or to a truncated protein which lacks the normal protein function [7, 17].

Here, we identified a novel nonsense mutation in a Pakistani family, designated R523X, which is located in exon 10 and is part of the first linker which connects domain 1 to domain 2. This site appears to be critical for the normal formation and function of the channel, since a previous mutation in the same linker designated S459X was associated with loss of function [7].

This mutation expands the body of evidence supporting the critical role of the Na1.7 channel in the perception of pain, and suggests that the absence of this channel cannot be compensated for by other channels.

In conclusion, we have extended the spectrum of mutations in Na1.7 channels in CIP, providing a framework for unraveling the complex mechanisms of pain perception.

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