Retinal and Ocular Toxicity in Ocular Application of Drugs and Chemicals – Part II: Retinal Toxicity of Current and New Drugs

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Abstract
Aims: Retinal pharmacotherapy has gained great importance for the treatment of various retinal diseases. An increasing number of drugs have been constantly released into the market, especially for wet age-related macular disease and diabetic macular edema. In this review, the issues concerning the toxicity of current and new classes of drugs are discussed. Methods: An extensive search of the literature was performed to review various aspects of drug toxicity in retinal pharmacotherapy. The different major classes of drugs, such as corticosteroids, antibiotics, antimetabolites, antineoplastic agents, monoclonal antibodies (mAbs), nonsteroidal anti-inflammatory drugs, enzymes, fibrinolytics, miscellaneous anti-inflammatory and antiangiogenic agents, as well as toxicity unrelated to the drug were identified and discussed. Results: Corticosteroids like fluocinolone, dexamethasone or triamcinolone at low dose cause little damage to the retina, but at high doses signs of toxicity have been well documented. Complications like cataract and glaucoma are quite common with corticosteroids. Aminoglycosides showed differences in the type and doses associated with toxic reactions, thereby the following order of toxicity can be described (from most toxic to least toxic): gentamicin > netilmicin = tobramycin > amikacin = kanamycin. Vancocycin at the usual dose of 1 mg is not toxic to the retina, while further studies are necessary in order to clarify the safety of new-generation quinolones. 5-Fluorouracil has been shown to be nontoxic to the retina after an injection of 2.5 mg in animals. mAbs like ranibizumab and bevacizumab were demonstrated to be safe to the retina in cell culture, animals and humans at high doses. The exact biocompatibility of nonsteroidal anti-inflammatory agents like diclofenac needs further evaluation. Preservatives like benzyl alcohol and changes in pH or osmolarity exert an influence on the toxic effects of intravitreally applied drugs. Conclusions: A great number of drugs are now used mainly intravitreally without relevant retinal toxicity.

Key Words
Avastin • Lucentis • Retina • Bevacizumab • Ranibizumab • Monoclonal antibodies • Tumor necrosis factor • Vascular endothelial growth factor • Corticosteroids • Antimetabolites • Triamcinolone • Nonsteroidal anti-inflammatory drugs • Microplasmin

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Introduction

Local ocular delivery of drugs has become a frequent and important technique for the treatment of retinal diseases [1–3]. Intravitreal injections have been applied widely for a variety of retinal diseases, including inflammatory, neovascular and edematous conditions of the macula, retina and vitreous [4–7].

The first report of intravitreal injection as a therapeutic modality was to administer air for the repair of rhegmatogenous retinal detachment [8]. However, later in the 1970s, intravitreal injections arose again as a common approach in the therapy of sight-threatening endophthalmitis, with aminoglycosides and cephalosporins. In 1987, the first case report described the benefit of delivery of ganciclovir sodium to a patient with cytomegalovirus (CMV) retinitis secondary to acquired immunodeficiency syndrome (AIDS) [9]. Later, in 1998 fomivirsen sodium (Vitravene™) was approved by the US Food and Drug Administration (FDA) for intravitreal injection in the treatment of CMV retinitis. Further studies were performed in the 1980s and 1990s of intravitreal administration of 5-fluorouracil (5-FU) for patients with proliferative vitreoretinopathy (PVR), dexamethasone for diabetic retinopathy after vitrectomy, and tissue plasminogen activator for the management of submacular hemorrhage [10, 11]. Over the past two decades, numerous drugs have been developed for the treatment of different retinal and choroidal diseases. Triamcinolone acetonide (TA) has been widely used for the treatment of different edematous retinal diseases, such as diabetic macular edema and choroidal neovascular membranes. Recently, monoclonal antibodies (mAbs) emerged as the first therapeutic option with gain lines in visual acuity for choroidal neovascularization associated with age-related macular disease, with off-label bevacizumab and FDA-approved ranibizumab.

In this review, the safety issue of major classes of drugs applied or with potential use in retinal disease will be discussed. Additionally, biocompatibility issues regarding several drugs that may be potentially exploited to treat diseases of the back of the eye are presented. The issues surrounding the toxicity after systemic delivery to the posterior segment will not be discussed in this work.

Major Classes of Drugs and Their Safety Profile after Local Ocular Application for Retinal Therapy

Corticosteroids

Corticosteroids are powerful drugs widely used in ophthalmology, including the therapy of retinal diseases. In the past two decades, much research has been conducted to evaluate the indications of various types of steroids for vitreoretinal diseases. However, beyond the efficacy, one of the most important issues for the application of corticosteroids is to define the limitation of their side effects.

Fluocinolone acetonide is a synthetic corticosteroid with low solubility in aqueous solution. Experiments in rabbits provided the preclinical toxicity profile of fluocinolone with regard to the retina. Clinical examination, electroretinography or histological examination determined the safety of both 2 and 15 mg synthetic fluocinolone acetonide in a preclinical study by Jaffe et al. [12]. Neither electroretinographic (ERG) alterations nor histology abnormalities were observed in their in vivo investigation. However, further clinical experience with implants containing 2 or 6 mg of fluocinolone acetonide for therapy of uveitis revealed that a considerable number of patients experienced complications, which included glaucoma, cataract and retinal vein occlusion [13].

Dexamethasone belongs to the synthetic glucocorticoid class of steroid hormones with potent anti-inflammatory and immunosuppressant activities. Kwak and D’Amico [14] evaluated the retinal toxicity of doses ranging from 440 to 4,000 μg dexamethasone in rabbit eyes and observed that with the 440-μg dose, only a transient increase in staining of the Müller cells was observed, which normalized after 2 days. Recent research in cell culture accorded with the previous works, as a higher dose of 800 μg dexamethasone was found to cause a reduction in the number of retinal cells. These experimental studies provided the basis for the safety profile of dexamethasone in humans, but future studies need to determine which dose, 400 or 800 μg, is safer in the treatment of edematous, proliferative and neovascular retinal diseases.

The first commercially available TA was Kenalog 40 (Bristol-Myers Squibb, Princeton, N.J., USA), an injectable suspension of TA for intramuscular or intra-articular use only, not recommended for intravenous or intraocular use. However, a series of contradicting studies have recently been published regarding retinal toxicity after intravitreal TA injection. On the one hand, various experiments with intravitreal injection of 4, 16, 20 or...
30 mg TA resulted in normal histological and ERG retinal findings after 7 months. On the other hand, in one investigation the authors injected escalating doses from 0.5 to 20 mg of suspended preservative-free TA into rabbits and found, at doses of 4 mg or higher, prominent retinal damage manifested by damage to the photoreceptor outer segments and retinal pigment epithelium (RPE) [15]. Only few studies have addressed thus far the subretinal toxicity of TA, which could occur accidently after macular hole surgery. One investigation tested a subretinal injection of 3 mg/ml TA in primate eyes and described neither ultrastructural nor cellular retinal damage [16]. However, Maia et al. [17] disclosed in a morphological study disturbance to photoreceptor segments after subretinal injection of preservative-free TA (Ophthamos, São Paulo, Brazil), although no clinical abnormality on funduscopy or angiography was observed. These contrasting findings suggest that other factors may contribute to intravitreal retinal toxicity of TA, such as preservatives that will be discussed in a later section in this paper (fig. 1).

The clinical experience in recent years unraveled the risks to patients injected with intravitreal or periocular TA. The two most frequent complications are cataract and glaucoma, which occur in approximately 40 and 30% of cases, respectively [17]. Most patients with cataract may be managed with surgery; however, glaucoma can be treated with eyedrops, and surgical procedures may be required in only around 1% of eyes [17]. Less common complications are endophthalmitis and pseudophthalmitis, encountered in around 0.5% of patients [17]. Recently, 2 new TA injectable suspensions, specifically produced and FDA approved for intraocular use, were launched on the market: Triesence (Alcon Inc., Fort Worth, Tex., USA) and Trivaris (Allergan Inc., Irvine, Calif., USA). Triesence is a 40-48/ml preservative-free TA solution, and Trivaris is an 80-mg/ml drug solution, available in a dose of 4 mg in a volume of 50 μl. Triesence was evaluated clinically for visualization during pars plana vitrectomy. This phase III, observer-masked study was conducted in 6 centers by 10 surgeons, involving 60 enrolled patients undergoing pars plana vitrectomy, where up to 4 mg was administered to enhance visualization of vitreous and membranes. Triesence was well tolerated and effectively enhanced visualization of posterior segment structures during pars plana vitrectomy without any safety issues [18]. Moshfeghi et al. [19] conducted an interesting study comparing the crystal size of 3 different TA preparations: Kenalog 40 (Bristol-Myers Squibb), compounded preservative-free TA (PFTA, New England Compounding Center, Framingham, Mass., USA) and Triesence (Alcon Inc.). In vitro, PFTA had more aggregates of smaller size than either TA or Triesence. In contrast, Triesence had a much larger aggregate size than both PFTA and TA, and this increased over time. These findings correlate with the clinical observations that PFTA and TA tend to disperse throughout the vitreous, whereas Triesence tends to conglomerate and gravitate toward the lowest portion of the eye in a globular form [19].

Antibiotics

The current application of antibiotics in retinal therapy refers to infectious endophthalmitis and uveitis. Postoperative endophthalmitis following cataract surgery is the main indication of intravitreal injection of antibiotics, even though in many cases there is no known microorganism responsible for the infection. Another important indication is posterior segment viral uveitis which is usually treated with systemic or intravitreal injections of acyclovir, valacyclovir and ganciclovir. Every intravitreally injected antibiotic drug could potentially lead to retinal toxicity, which will be addressed in further detail with consideration of the clinically safe dose (table 1).

Aminoglycosides

Amikacin used to be one of the most commonly used agents in the treatment of endophthalmitis in the past. Nowadays, it is well known that the aminoglycosides may pose a high risk of toxic effects to the retina. In vitro studies with isolated retinas showed that the b-wave in electroretinography is reduced in amplitude in the presence of low-dose (1 mg/ml) and completely eliminated by high-dose gentamicin (10 mg/ml) [20]. In in vivo ERG studies in animals, intraocular administration of gentamicin eliminated the b-wave and reduced the c-wave amplitude [20]. Histopathological evaluation demonstrated diffuse disruption of the nerve fiber layer and the inner plexiform layers in eyes exposed to this aminoglycoside antibiotic [20].

Regarding amikacin, an animal study disclosed retinal toxicity manifested by macrophages in the subretinal space, disorganization of the outer segments and RPE, and discontinuities in Bruch’s membrane after repeated intravitreal injection of 400 μg of amikacin [21]. D’Amico et al. [22] performed a comparative toxicity study of the aminoglycoside antibiotics (tobramycin, amikacin, netilmicin and kanamycin) after intravitreal injection into rabbits with doses ranging from 100 to 3,000 μg. The earliest manifestations of toxicity were confined to...
Fig. 1. After subretinal injection of preservative-free TA, the preservative, and control, different types of retinal cells manifest various signs of damage. The damage induced by the preservative (I–M) is more severe to neuroretinal posterior cells than that caused by the drug itself (E–H) and balanced salt solution (A–D). (Reprinted from Maia et al., 2008.)
the outer retina with each drug, with lamellar lysosomal inclusions in the retinal pigment epithelium as the earliest finding. However, the aminoglycosides displayed marked differences in the threshold dose required to produce toxic reactions, permitting the following order of toxicity (from most toxic to least toxic): gentamicin = netilmicin = tobramycin = amikacin = kanamycin [22].

Further experience demonstrated that repetitive injections in nonvitrectomized eyes may result in increasing retinal toxicity. Later on, many cases of macular infarction after intravitreal injection of aminoglycosides such as amikacin have been reported in humans [23]. An accidental intravitreal injection of gentamicin could also promote macular infarction (fig. 2). This has led ophthalmologists to search for other Gram-negative targeting antibiotics for routine intravitreal injections.

**Cephalosporins**

Ceftazidime, a third-generation cephalosporin emerged as a good alternative to aminoglycosides for the treatment of endophthalmitis. An in vivo animal study with monkeys showed no toxicity at the commonly used concentration of 2.25 mg ceftazidime, but others showed that ceftazidime may not be toxic in vitrectomized rabbit eyes [24]. However, it appears that cephalosporin may cause some degree of toxicity at otherwise nontoxic concentrations in a silicone-filled eye. Based on these preliminary data, surgeons have applied ceftazidime intravitreally in doses up to 2.25 mg for the treatment of endophthalmitis.

**Vancomycin**

Vancomycin remains the antibiotic of choice, targeting highly pathogenic Gram-positive microorganisms, usually applied to patients intravitreally at a dose of 1 mg in 0.1 ml. In vivo animal studies have shown no toxic effect of this antibiotic when used in an infusion solution that was given intracoarcularly after or during vitrectomy in rabbits [25]. However, in a silicone-filled eye, nontoxic concentrations of vancomycin may cause toxicity; there-

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**Table 1. Safe doses for various ophthalmic uses of antibiotics**

<table>
<thead>
<tr>
<th>Drug</th>
<th>Topical</th>
<th>Subconjunctival</th>
<th>Intravitreal</th>
</tr>
</thead>
<tbody>
<tr>
<td>Amikacin</td>
<td>10 mg/ml</td>
<td>25 mg</td>
<td>400 µg</td>
</tr>
<tr>
<td>Cefazolin</td>
<td>50 mg/ml</td>
<td>100 mg</td>
<td>2,250 µg</td>
</tr>
<tr>
<td>Ceftriaxone</td>
<td>50 mg/ml</td>
<td>–</td>
<td>2,000 µg</td>
</tr>
<tr>
<td>Clindamycin</td>
<td>50 mg/ml</td>
<td>15–50 mg</td>
<td>1,000 µg</td>
</tr>
<tr>
<td>Gentamicin</td>
<td>8–15 mg/ml</td>
<td>10–20 mg</td>
<td>100–200 µg</td>
</tr>
<tr>
<td>Imipenem</td>
<td>5 mg/ml</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Penicillin G</td>
<td>100,000 units/ml</td>
<td>0.5–1 × 10⁶ units</td>
<td>300 units</td>
</tr>
<tr>
<td>Piperacillin</td>
<td>12.5 ng/ml</td>
<td>100 mg</td>
<td>–</td>
</tr>
<tr>
<td>Tobramycin</td>
<td>8–15 mg/ml</td>
<td>10–20 mg</td>
<td>–</td>
</tr>
<tr>
<td>Vancomycin</td>
<td>20–25 mg/ml</td>
<td>25 mg</td>
<td>1,000 µg</td>
</tr>
</tbody>
</table>

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**Fig. 2.** A Color fundus picture showing extensive damage of neurosensory retina and its vasculature as well as whitening and hemorrhages of the posterior pole after intravitreous injection of gentamicin. B Fluorescein angiography demonstrating signs of vasculities, vascular occlusion, and capillary non-perfusion after intravitreous injection of gentamicin (courtesy of Dr. Harry Flynn).
fore, vitreous status should be evaluated when vancomycin is the antibiotic of choice as well [26, 27]. In clinical practice, vancomycin has been associated with postoperative cystoid macular edema when infused as intracameral injection for prophylaxis during cataract surgery.

**Quinolones**

The ocular toxicity of another class of antibiotics, the quinolones, has been investigated in the recent past specifically for antibiotic prophylaxis in ocular surgery. Regarding intravitreal use, ciprofloxacin has not been associated with toxicity at the therapeutic levels of 100–500 μg in rabbits, and significant retinal damage has been observed only at 2 mg [28].

The fourth-generation quinolones, which include moxifloxacin and gatifloxacin, have recently been shown to be useful for clinical use. An in vitro study showed that at concentrations higher than 160 μg/ml, moxifloxacin induced adverse effects on primary RPE and neuronal retinal cells with regard to cell proliferation and cell viability [29]. Further studies in vivo showed that intravitreal injection of moxifloxacin did not cause retinal toxicity up to 100 μg/ml in mice or 150 μg in rabbits [30]. In vivo, intravitreal injection of the other quinolone, gatifloxacin, at doses varying from 50 to 400 μg, caused no retinal toxicity, assessed clinically and microscopically in rabbits [30].

Clinical experience in retinal toxicity studies revealed that the current recommended dose for intracameral injection of ciprofloxacin is less than 25 μg. In humans, intravitreal injections of ciprofloxacin 100 μg, ofloxacin 50 μg/ml, trovafloxacin 25 μg or less, moxifloxacin 160 μg/0.1 ml or less and pefloxacin 200 μg/0.1 ml are considered nontoxic to the retina and intraocular structures [31].

**Antifungal Agents**

Fungal infections may cause severe and poor-prognosis endophthalmitis. Amphotericin B has been traditionally used for the treatment, either systemically or intravitreally usually injected at doses varying from 1 to 50 μg. In one study with the application of escalating doses from 10, 20, 30 to 50 μg, the 3 higher doses of amphotericin B appeared to be associated with stronger degrees of retinal toxicity [32]. Based on animal experiments, intravitreal amphotericin B in doses of 5 or 10 μg remains an appropriate therapeutic option for patients with severe fungal endophthalmitis, for instance secondary to *Aspergillus* [32].

An alternative to amphotericin is the use of intravitreal fluconazole. An in vitro study showed no toxicity with 20 μg/ml exposure to fluconazole [33]. Further animal data revealed no retinal toxicity resulting from vitrectomy with a 2 mg/ml fluconazole infusion in an experimental model of candidal endophthalmitis. A single intravitreal injection of fluconazole at a concentration of 100 μg/ml and higher caused harmful retinal changes with disorganization of the photoreceptor outer segments [34]. Clinical experience revealed that intravitreal injection of 10 μg/0.1 ml fluconazole may be the safe dosage for intraocular fungal infection [32].

New-generation triazoles, including voriconazole, posaconazole and ravuconazole, have been shown in laboratory studies and clinical experience to have very good safety profiles with few side effects [35]. Regarding toxicity, Gao et al. [36] showed that voriconazole did not cause any retinal damage by either ERG or histological studies when the intravitreal concentration was 25 μg/ml. When the voriconazole concentration increased to 50 μg/ml, focal areas of retinal necrosis were seen; however, electroretinograms remained unaffected. With these results, they postulated that intravitreal injections of up to 100 μg/ml could be safe for the human eye. Sen et al. [37] demonstrated 5 cases of culture-proven fungal endophthalmitis. In all cases, the drug was administered at 50 μg/0.1 ml, giving a final concentration of 12.50 μg/ml in the vitreous cavity [37]. No sign of retinal toxicity was observed: 60% had anatomical success and also 60% had a final visual acuity of counting fingers or better [37].

**Antiviral Agents**

Ganciclovir is commonly used in the antiviral treatment of CMV infection. Ganciclovir dosages of up to 200 μg/0.1 ml appear to be safe for serial intravitreal injections in rabbit eyes following vitrectomy and silicone oil insertion. In nonvitrectomized eyes, ganciclovir in doses >300 μg induced severe morphological retinal damage, although at a lower dose of 200 μg, ganciclovir caused only minor functional damage characterized by changes in the ERG b-wave in rabbits [26]. Similar to other types of antibiotics, ganciclovir also induces sporadic cases of macular infarction in patients. A case report of inadvertent intravitreal injection of a high dose of ganciclovir (40 mg/0.1 ml) for CMV retinitis in a patient with AIDS led to permanent retinal damage and visual loss [38]. Recently, ganciclovir was administered as consecutive intravitreal injections in doses varying from 400 μg to 5 mg for treating intraocular viral infections [39].
Foscarnet is a phosphonic acid derivative (Foscavir, Astra Zeneca, London, UK) an antiviral medication used to treat herpesviruses, including CMV and herpes simplex virus types 1 and 2. Foscarnet is used intravenously, but there are some reports involving intravitreal injection for CMV retinitis. The largest cohort is a retrospective study with 193 patients submitted to intravitreal injection of 2.4 mg/0.1 ml foscarnet. There was good clinical response regarding toxicity/complications in 301 treated eyes, where 15 had retinal detachment, 13 intraocular hemorrhage, 3 endophthalmitis and 2 cataracts. Intravitreal foscarnet appears to be an alternative treatment for patients, where intravenous anti-CMV drugs are not tolerated or affordable, but the complications of this treatment should be considered [40]. López-Cortés et al. [41] performed an experimental study in rabbits to evaluate the pharmacokinetics of intravitreal ganciclovir (196 and 800 µg) and foscarnet (960 µg). Both doses of ganciclovir yielded retinal levels above the mean inhibitory concentration against most human CMV isolates for more than 60 h, while foscarnet retinal levels were lower than the CMV mean inhibitory concentration by 36 h after drug administration. These results suggest that the intravitreal administration of ganciclovir has a better pharmacokinetic profile than does foscarnet for the treatment of CMV retinitis, requiring fewer injections, which is particularly important in viral retinitis [41].

Fomivirsen sodium (Vitravene, Isis 2922; Isis Pharmaceuticals, Carlsbad, Calif., USA) is the latest addition to the armamentarium of treatment options for local CMV retinitis. Fomivirsen is a phosphorothioate oligonucleotide, which is administered intravitreally; it inhibits CMV replication through an antisense mechanism, binding to viral messenger RNA and blocking its transcription. The FDA approved fomivirsen in August 1998, as highly active antiretroviral therapy began to dramatically decrease the incidence of CMV retinitis. Clinical trials with intravenous fomivirsen is limited, where the recommended dose is 300 µg every other week for 1 dose followed by maintenance doses every 4 weeks after induction. Reported adverse effects with the approved dosage include transient and reversible inflammation in the anterior chamber (19%), increase in intraocular pressure (IOP; 19%), vitreitis (11%) and uveitis (5%); cataract (9%) has also been reported [42]. Isolated case reports describe foveal RPE stippling with reversible bull’s eye maculopathy [43], peripheral RPE stippling associated with visual field loss [44] and marked retinal toxic side effects after 495-µg doses [45]. Bull’s eye maculopathy was diagnosed after five 330 µg doses in 1 case and after 15 injections bilaterally in another. In both cases, fomivirsen was administered every other week; RPE stippling was diagnosed after weekly injections of 165 µg for 3 weeks.

Cidofovir or hydroxyphosphonylmethoxypropyl cytosine has been shown to be a potent inhibitor of CMV proliferation in vitro. A single intravitreal injection of cidofovir was able to prevent progression of herpes simplex virus type 1 retinitis for an almost 10 times longer period of time than with a single intravitreal injection of ganciclovir in a rabbit model [46–50]. No signs of retinal toxicity were found at the doses ≤100 µg/eye injected intravitreally in rabbits and Papio cynocephalus monkeys [48]. Phase I/II clinical studies showed that a single intravitreal injection of 20 µg of cidofovir was safe and efficacious against CMV retinitis in human eyes for a median time of 55 days. At higher doses (40–100 µg/eye), however, the drug caused irreversible ocular hypotony in 33% (at 40 µg) and 100% (at 100 µg) of eyes [51, 52].

Recently, an experimental study investigated the highest nontoxic dose of hexadecylxyloxypropyl cyclic hydroxyphosphonylmethoxypropyl adenine (HDP-cHPMPA), a novel, potent, intravitreally injectable, slow-release crystalline drug for long-acting treatment of CMV retinitis. Three doses (55, 100 and 550 µg/eye) were tested in 9 pigmented rabbits, and a confirmation toxicity study with the dose equivalent to the highest nontoxic dose in rabbits was performed in 9 guinea pig eyes (a second species) to study the potential adverse effect on IOP. Intravitreal injection of the highest nontoxic dose of 55 µg/eye of HDP-cHPMPA in rabbit eyes yields a calculated intravitreal concentration of 65 µM, which is 3,250-fold greater than the 50% effective concentration against human CMV (0.02 µM). Regarding IOP, the drug did not cause hypotony in rabbit and guinea pig eyes. Finally, HDP-cHPMPA could be detected in the vitreous cavity over 4 months after the injection [53].

**Antimetabolites and Antineoplastic Drugs**

5-Fluorouracil

5-FU is a potent antimetabolite that inhibits the proliferation and contraction of intraocular fibroblasts and the RPE in animal models and in vitro studies. 5-FU is converted inside the cell to fluoropyrimidine nucleotides that act blocking DNA synthesis due to the inhibition of thymidylate synthetase and decrease RNA function secondary to incorporation of 5-FU metabolites into newly synthesized RNA. The antiproliferative and anticontractile effects of 5-FU are probably due to the synthesis of abnormal RNA [54, 55].
Several studies have evaluated the retinotoxicity of 5-FU in animal models [54, 56, 57]. Blumenkranz et al. [54] found that 5-FU was not toxic to rabbit retina after a single intravitreal dose of 2.5 mg in nonvitrectomized eyes or after subconjunctival injections of 10 mg daily for 7 days. Also, Barrada et al. [58] found 750 μg of intravitreal 5-FU to be nontoxic in primate eyes. However, when combined with 0.10 or 0.15 μg/ml of vincristine, toxic effects were observed. Leon et al. [59] studied the retinotoxicity of 5-FU, assessing protein synthesis in the rabbit retina, and their data showed marked inhibition after 2.5 mg of 5-FU, using 3 different methods for protein synthesis evaluation in the rabbit retina. Finally, Kivilcim et al. [60] showed that in silicone-filled eyes, the safe dose is up to 200 μg, and therefore the vitreous status is important to know before intravitreal administration of 5-FU. In another study, Cardillo et al. [61] designed a sustained-release pellet of 1.5 mg naproxen and 5-FU with continuous drug delivery. In this study, there were no drug-related toxic effects evident on histopathological or ERG examination of eyes containing the naproxen/5-FU pellet [61].

In humans, 5-FU was used to inhibit PVR after vitrectomy. In a randomized clinical trial, 5-FU 10 mg, given as a single intravitreal injection, was evaluated in 26 vitrectomized eyes [62]. It was reported that corneal edema was slightly more prevalent in the 5-FU group and that persistent retinal damage was not observed, based on visual acuity [54].

Mitomycin C

Mitomycin C (MMC) is an anticancer drug that covalently binds to DNA and causes DNA cross-linking and cell death. This drug is not currently being used intravitreally in humans, although several ophthalmic experimental studies have been carried out [56, 63].

The toxicity of 0.2, 0.3 and 0.4 mg/ml MMC to the rat retina was investigated by Kawashima et al. [64], who reported that ERG changes follow intravitreal injection. No signs of retinal toxicity were observed after 0.2 mg/ml at 7 days, mild changes with 0.3 mg/ml at 7 days and profound damage with 0.4 mg/ml in as early as 2 days. The highest dose (0.4 mg/ml) showed selective degeneration of Müller cell processes on day 2, RPE changes on day 4 and irregular arrangement of the outer nuclear layer and photoreceptors on day 7. MMC (0.4 mg/ml) administered in the anterior chamber showed no evidence of ERG and histological changes at 2 and 7 days after injection [64]. Macky et al. [63] evaluated the retinal toxicity of intravitreal injection of MMC combined with TA (0.784 μg/0.2 μl) in rats. No difference was observed between MMC-TA conjugate and a control group, based on histopathology and electroretinogram 5 or 20 days after treatment. No signs of retinal damage such as retinal necrosis, photoreceptor cell loss, cystic degeneration or inflammatory cell infiltration were observed [63].

Mitomycin was also investigated in rabbit eyes. Velez et al. [65] showed no ERG abnormalities after intravitreal injection of 2 μg/0.1 ml. On the other hand, 4 μg/0.1 ml caused a decrease of 34.5% in the b-wave, and with 8 μg/0.1 ml, the b-wave was absent. In a PVR model, Yu and Chung [66] evaluated the safety and efficacy of MMC in a PVR rabbit model. Regarding safety, a dosage of up to 4 μg/0.1 ml was not toxic in nonvitrectomized rabbits, but in gas-filled eyes, the safe dose was 2 μg/0.1 ml. In the PVR rabbit model of intravitreal injection of RPE cells, there was a remarkable reduction in advanced retinal detachment after intravitreal MMC, 9.1% with the 0.2-μg dose and no occurrence with the 1.0-μg dose, compared to 84.6% in untreated eyes [66]. There are no reports of the use of intravitreal MMC in humans; however, based on experimental studies the safe dose varies from 2 to 4 μg/0.1 ml.

Colchicine

Colchicine, a mitosis-arresting phenanthrene derivative extracted from various species of Colchicum (especially C. autumnale), is a drug commonly used in the prophylaxis and treatment of gout, by inhibiting cell proliferation and acting as an anti-inflammatory drug. This drug is known to be neurotoxic, and peripheral neuropathy associated with its use has been described clinically. This drug is not currently being used intravitreally in humans, although several experimental ophthalmic studies have been carried out [67].

The effects of 1.0–100 μg of intravitreal colchicine in monkeys were studied by ophthalmoscopy, and light and electron microscopy. Optic nerve atrophy was ophthalmoscopically evident within 4 weeks after a single dose of 10 μg or more [68]. Morphological changes included progressive swelling of retinal neurons, accumulation of a fibrillogranular material, displacement of organelles and loss of microtubules. Cell membrane rupture was observed in ganglion cells and in photoreceptors after as little as 1 μg of colchicine [68], while relative sparing of the cone cells was noted. Similar effects were observed after intravitreal injection of colchicine in the rabbit, cat and rat. However, the monkey retina appears to be more sensitive to colchicine [67].
Methotrexate

Methotrexate (MTX) is an antineoplastic agent used to treat a variety of malignant conditions. It reversibly binds to and inhibits dihydrofolate reductase, which leads to a depletion of purine and thymine nucleotides, consequently blocking DNA, RNA and protein synthesis. MTX affects cells in the S phase of the cell cycle [69], and it is most effective against rapidly proliferating cells. MTX is mostly used in combination with other antineoplastic agents in the treatment of the ocular neoplasias [65, 69, 70].

Velez et al. [65] evaluated the ocular toxicity of intravitreal combination chemotherapy, composed of 3 cycles of serial injections of MTX (400 μg on days 1, 4 and 6), a single injection of 5-FU (500 μg on day 2) and dexamethasone (500 μg on day 7). The investigation was performed in New Zealand white rabbits, and fluctuations in the a- and b-wave amplitudes in both the experimental (right) and control (left) eyes were observed. Full recovery was noted, however, with no statistically significant difference in the mean of the a- and b-wave amplitudes between the treated and control eyes after the final cycle [65]. Histopathological examination confirmed ERG findings, showing no photoreceptor or ganglion cell layer damage in the experimental eyes compared with controls, away from the site of injection. The optic nerve and medullary rays also appeared intact in both the treated and control eyes [65]. Another study determined the retinal toxicity of combined chemotherapy in rabbit eyes, which consisted of: 5-FU, 375 μg; doxorubicin, 3 μg; neomycin, 15 μg; thiotepa, 12 μg; etoposide, 150 μg, and MTX, 600 μg. This combination regimen caused a severe ERG decrease (decrease in b-wave amplitude to less than 25%) and retinal changes (generalized thinning of the retina, disorganization of retinal layers and loss of photoreceptor outer segments) [71].

In humans, MTX has been used in both inflammatory and neoplastic ophthalmic disease. Intravitreal MTX has been effectively used in the treatment of intraocular lymphomas, both in primary central nervous system lymphoma and non-Hodgkin’s lymphoma [72]. It has also been used in eyes with uveitis and advanced proliferative diabetic retinopathy [70, 73]. Adverse reactions to intraocular MTX have been reported, where corneal epitheliopathy is the main complication, followed by corticosteroid-responsive sterile endophthalmitis and less common intravitreal hemorrhage. Cataract was probably related to MTX toxicity, but no special type of cataract was noted. Other conditions, including irreversible loss of visual acuity, maculopathy and progression of cataract, could not be directly attributed to the injection since these complications occurred in patients with previous severe disease [70, 73]. A single intravitreal MTX injection (200 or 400 μg), used in the treatment of proliferative diabetic retinopathy and chronic uveitis, was also studied, and the authors did not report any adverse effect [74]. The cumulative intravitreal MTX dose ranging from 200 to 1,200 μg in silicone-filled eyes was studied, and no adverse effects were observed following single or serial injections at doses up to 1,200 μg [75].

Melphalan

Melphalan, also known as L-phenylalanine mustard, phenylalanine mustard or L-sarcosylsin, is a phenylalanine derivative of nitrogen mustard. Melphalan is a bifunctional alkylating agent, which is active against selective human neoplastic diseases [76]. It is FDA approved for palliative therapy of multiple myeloma and nonsectable epithelial carcinoma of the ovary. Considering ocular tumors, melphalan has been used in cases of advanced retinoblastoma, injected intravitreously and into the ophthalmic artery [77].

Ueda et al. [76] studied the effects of an intravitreal injection of melphalan on electroretinography and on the retinal structure in rabbits to establish the nontoxic dose for its clinical use. They concluded that 5.9 μg/ml melphalan was not toxic; this concentration was higher than the 4 μg/ml necessary to completely suppress colony formation by retinoblastoma cells in vitro [76]. Another study investigated the toxic effects of intravitreal perfusion with melphalan (5, 10 and 20 μg/ml) during vitrectomy on the rabbit retina. In the 5 μg/ml perfusion group, electroretinograms and histology showed no substantial changes compared with control fellow eyes during 28 days postoperatively. However, the groups of 10 and 20 μg/ml displayed a decreased a- and b-wave amplitude and histological changes [78].

In humans, Kaneko and Suzuki [79] reported the treatment of 41 eyes with intravitreous injection of melphalan for retinoblastoma; 56.1% (23/41) of these eyes were salvaged, and 43.9% (18/41) of these eyes were enucleated in the long run. Severe side effects, such as extraocular dissemination of tumor cells related to intravitreal injection, were not experienced [79]. Vitreous injection of melphalan may be one of the promising treatment modalities for conservative therapy.

Investigators in Japan developed a technique to deliver chemotherapy to the orbit by temporarily occluding the carotid artery distal to the ophthalmic artery and injecting intra-arterial melphalan into the cervical internal ca-
rotid [77]. Recently, a phase I/II study using this technique was published [77], where neither severe systemic side effects nor toxicity to the cornea, anterior segment, pupil or motility were reported [80]. Considering the retina, retinal ischemia was observed in 1 (previously irradiated) eye, and another eye showed no toxicity after intraarterial chemotherapy but did develop a radiation-like retinopathy after brachytherapy [80].

**Monoclonal Antibodies**

Current and future indications of mAbs will be discussed, and table 2 provides information on the mechanism of action, level of evidence and FDA status.

**Table 2. Current and future mAbs in the treatment of ocular diseases**

<table>
<thead>
<tr>
<th>Drug name</th>
<th>Brand name</th>
<th>Company</th>
<th>Target</th>
<th>Antibody composition</th>
<th>Current indications</th>
<th>Future indications</th>
<th>Level of evidence in ophthalmology</th>
<th>FDA status in ophthalmology</th>
</tr>
</thead>
<tbody>
<tr>
<td>Adalimumab</td>
<td>Humira</td>
<td>Abbott</td>
<td>TNF-α</td>
<td>Fully human</td>
<td>Uveitis and inflammatory diseases</td>
<td>Wet AMD and macular edema</td>
<td>Ongoing clinical trial</td>
<td>Off label</td>
</tr>
<tr>
<td>Bevacizumab</td>
<td>Avastin</td>
<td>Genentech</td>
<td>VEGF-A</td>
<td>Humanized</td>
<td>Wet AMD, macular edema and PDR</td>
<td>Neovascular glaucoma</td>
<td>Ongoing clinical trial</td>
<td>Off label</td>
</tr>
<tr>
<td>Daclizumab</td>
<td>Zenapax</td>
<td>Roche</td>
<td>CD25</td>
<td>Chimeric</td>
<td>Uveitis</td>
<td>Dry AMD</td>
<td>Phase II</td>
<td>Off label</td>
</tr>
<tr>
<td>Etanercept</td>
<td>Enbrel</td>
<td>Amgen</td>
<td>TNF-α and β</td>
<td>Recombinant dimeric protein</td>
<td>Uveitis and inflammatory diseases</td>
<td>Wet AMD and macular edema</td>
<td>Ongoing clinical trial</td>
<td>Off label</td>
</tr>
<tr>
<td>Infliximab</td>
<td>Remicade</td>
<td>Centocor</td>
<td>TNF-α</td>
<td>Chimeric</td>
<td>Uveitis and inflammatory diseases</td>
<td>Wet AMD and macular edema</td>
<td>Ongoing clinical trial</td>
<td>Off label</td>
</tr>
<tr>
<td>Ranibizumab</td>
<td>Lucentis</td>
<td>Genentech</td>
<td>VEGF-A</td>
<td>Antibody fragment</td>
<td>Wet AMD and macular edema</td>
<td>Neovascular glaucoma and PDR</td>
<td>FDA-approved</td>
<td>Approved</td>
</tr>
<tr>
<td>Rituximab</td>
<td>Rituxican</td>
<td>Genentech</td>
<td>CD20</td>
<td>Chimeric</td>
<td>Lymphoma and inflammatory diseases</td>
<td>Dry AMD</td>
<td>Phase I</td>
<td>Off label</td>
</tr>
</tbody>
</table>

AMD = Age-related macular disease; PDR = proliferative diabetic retinopathy; TNF = tumor necrosis factor; VEGF = vascular endothelial growth factor.

In vitro cellular assays of exposure to various concentrations of bevacizumab (0.08 μg/ml to 1 mg/ml) have shown few toxic effects to neuronal cells such as the ganglion cells, neuroretinal cells and RPE cells, as well [81–83]. In rabbits, mAbs penetrated the full thickness of the retinal tissue by 24 h, and their presence gradually decreased to undetectable levels at 4 weeks [84]. Experiments in monkeys revealed some bevacizumab immunoreactivity in the inner layers of the retina and also in the choroid as early as 1 day after injection. Thereafter, the immunoreactivity was even more evident in the outer layers and the choroid within the following 7 days [85].

Retinal cells interact actively with cytokines and growth factors to maintain their healthy environment and to promote vision. In this context, vascular endothelial cells produce and depend on VEGF for their multiple bioactivities, thereby being susceptible to anti-VEGF drugs. To study the effects of bevacizumab on various types of retinal cells, Kaempf et al. [86] exposed cultured adult porcine neurosensory retinas joined to the RPE/choroid layer to 3 doses of bevacizumab (0.25, 0.5 and 1.25 mg/ml) for 3 days. Their results showed no toxic effects on ganglion or photoreceptor cells at any concentration of bevacizumab. However, they observed significantly enhanced smooth muscle actin expression in retinal blood vessels in the presence of bevacizumab, which may imply a loss of smooth muscle cell modulation in normal retinal vessels by VEGF [86]. In contrast to those data, Luthra et al. [83] found no toxicity to microvascular retinal cells in vitro after their exposure to 0.125, 0.25, 0.50 and 1 mg/ml bevacizumab for up to 24 h.
A large body of animal studies has been released about the biocompatibility and safety of bevacizumab for ophthalmology. Consecutive experimental investigations in rats, rabbits and primates revealed that intravitreal bevacizumab at various concentrations up to 3 mg/ml demonstrated no functional or morphological toxicity to the retina [84, 87–89]. The preclinical safety of ranibizumab has been evaluated in primate eyes, showing that 0.5 mg injection of the mAb fragment caused reduced leakage from choroidal neovascularization, while no signs of retinal toxicity have been encountered [90]. However, a few recent reports of experimental studies demonstrated some signs of retinal damage after intravitreal bevacizumab. In primates, intravitreal bevacizumab induced choriocapillaris abnormalities manifested by reduced choriocapillaris endothelial cell fenestrations by densely packed thrombocytes and leukocytes within the vascular lumen [91]. Moreover, Manzano et al. [88] reported signs of ocular inflammation after intravitreal injection of high-dose bevacizumab at 5 mg in rabbit eyes. Also in rabbits, intravitreal bevacizumab at a dose of 1.25 or 3 mg caused both mitochondrial changes in the inner segments of photoreceptors and intensive apoptotic protein expression of bax and caspase on immunohistochemistry in comparison to control [87], but no signs of toxicity were detected on light microscopy and ERG examination. The clinical importance of such findings is yet to be clarified.

Regarding human studies, to date intraocular bevacizumab and ranibizumab injections have produced few clinically important ocular side effects. In contrast to the crystalline steroid drug TA, intravitreal bevacizumab has not been shown to induce glaucoma or cataract progression. Further clinical experience with intravitreal bevacizumab revealed few sporadic cases of uveitis, vitreous hemorrhage, RPE tears or endophthalmitis [92, 93]. Clinically, Rosenfeld et al. [94] suggested that the maximum tolerated dose of ranibizumab is 0.5 mg, as higher doses over 1 mg caused clinically important intraocular inflammation. Additional clinical investigation disclosed that intravitreal ranibizumab induces few severe complications such as endophthalmitis, uveitis and vitreitis in the fellow eye, while minor ocular events reported were conjunctival bleeding, eye pain and floaters [92, 95, 96]. In 2008, a head-to-head comparison of ranibizumab versus bevacizumab to treat advanced age-related macular degeneration funded by the National Eye Institute was started, which should clarify the differences in safety issues between the two mAbs (http://www.nei.nih.gov/news/statements/amd_therapy.asp).

Anti-Tumor-Necrosis-Factor mAbs: Etanercept, Infliximab and Adalimumab

Three anti-TNF (tumor necrosis factor) agents, infliximab, etanercept and adalimumab, have been shown to
promote clinical control of uveitis and other inflammatory eye diseases [97, 98].

Although infliximab and etanercept have been mainly administered systemically, in the future, anti-TNF mAbs may be used in intraocular therapy for the management of severe intraocular inflammatory and angiogenic diseases. Intravitreal injections of infliximab at concentrations ≤1.7 mg have been shown to be safe to the retinal tissue in preliminary animal studies, although future detailed research should elucidate the intraocular safety and pharmacokinetics of infliximab and its vehicle [99]. Recently, our group investigated the intravitreal toxicity of infliximab in nonhuman primates, and it was shown to be safe up to a dose of 400 μg (fig. 3) [unpubl. data]. The pharmacokinetics and safety of intravitreal etanercept delivery at a dose of 100 μg have been investigated in rabbits in one study. Clinical examination, electoretinography and histology revealed no evidence of toxicity with retinal levels peaking at 4 weeks and still detectable after 8 weeks [100]. Another investigation on the intraocular safety of different doses of etanercept revealed that the anti-TNF mAb at doses up to 2.5 mg caused no retinal damage [101]. One study recently demonstrated the safety profile of intravitreal injections of various doses of adalimumab in rabbits. Their results showed normal retinal examination in eyes having received injections of balanced salt solution (BSS), 0.25 or 0.50 mg adalimumab; however, 1 mg adalimumab induced inflammatory changes, retinal histopathological alterations and significant ERG reduction [102]. In summary, most studies suggest a dose-dependent retinal toxicity profile for intravitreal anti-TNF mAbs, and further evaluation should determine the exact safe dosage for each drug.

Interestingly, large clinical trials have poorly addressed ocular complications after the systemic application of anti-TNF mAbs. Neuro-ophthalmic toxic effects of systemic infliximab, adalimumab and etanercept have been manifested by either anterior optic nerve neuropathy or oculomotor nerve palsy, and over 15 cases of optic neuritis have been reported [103, 104]. Ocular side effects such as cataracts, infections or increase in IOP have not been observed, although one study suggested the occurrence of mild vitreous hemorrhage in 2 patients [104, 105].

Anti-Cluster-of-Differentiation Antigen mAb: Rituximab

A few additional mAbs, including rituximab, daclizumab, efalizumab and alemtuzumab, showed positive results in animal and early clinical studies, and may represent useful adjuvant therapies for ocular lymphoma or ocular inflammation.

Rituximab (Rituxan®, Genentech, San Francisco, Calif., USA) is the chimeric mAb against cluster of differentiation 20 (CD20) antigen that was FDA approved as subcutaneous infusion for patients with recurrent low-grade B-cell lymphoma, while it may also be indicated for rheumatoid arthritis systemic lupus erythematosus, leukemia and Wegener’s granulomatosis [106]. Small clinical series revealed rituximab as an effective treatment for patients with refractory scleritis, orbital inflammation, and intraocular and extraocular lymphoma [103]. The intravitreal pharmacokinetics of rituximab has recently been investigated; high concentrations of anti-CD20 mAb were sustained, with a half-life of 4.7 days in one study, whereas in a second investigation the agent injected intravitreally at doses up to 1 mg induced neither retinal damage in rabbits nor clinical signs of toxicity in a small series of 5 patients [107, 108]. Comprehension of the intraocular safety of rituximab may enable intravitreal injection for the therapy of uveal/oculocerebral lymphoma and uveitis. Clinically, the anti-CD20 mAbs showed benefit in the therapy of intraocular lymphoma and uveitis, although much more clinical experience is warranted to clarify the indications and risks of anti-CD mAb.

Nonsteroidal Anti-Inflammatory Drugs

Nonsteroidal anti-inflammatory drugs have cystoid macular edema as their main indication for retina pharmacotherapy. They may be used either systemically or topically, and topical ketorolac, diclofenac and nepafenac are most frequently used. The use of both diclofenac and ketorolac as intravitreal injections has been assessed in rabbits. Diclofenac formulated in hyaluronan was toxic to the retina at doses ≥540 μg, as evidenced by indirect ophthalmoscopy, and light and electron microscopy. Electoretinography showed no toxic signs of intravitreal injection of 400 μg for this formulation after 25 days. Another study showed no signs of toxicity, either electoretinographically or histologically, of intravitreal injections of up to 300 μg of diclofenac and 3,000 μg of ketorolac, after 8 weeks [109]. Preservative-free ophthalmic solution of 0.25 or 0.5% (500 μg) ketorolac tromethamine in 0.1 ml was nontoxic to the retina in animals when assessed up to 4 weeks after injection, as shown by microscopic and ERG evaluation [109]. However, commercially available ketorolac (3 mg/0.1 ml) was toxic following multiple intravitreal injection in albino rabbit eyes [110].

Flurbiprofen is a member of the phenylalkanoic acid derivative family of nonsteroidal anti-inflammatory
drugs used to treat the inflammation and pain of arthritis. Morales et al. [111] investigated the retinal toxicity of intravitreal ketorolac Tris salt and flurbiprofen. Clinical examination, electroretinography results and histological examination demonstrated no signs of retinal toxicity for either drug at any dose (125, 250 or 500 μg or 1 mg).

Finally, intravitreal use of nonsteroidal anti-inflammatory drugs has not been assessed in depth in humans to date. Unlike corticosteroids, the nonsteroidal anti-inflammatory drugs may not be associated with a rise in IOP.

**Enzymes and Fibrinolytics**

Pharmacological vitreolysis refers to the capacity of altering the molecular organization of the vitreous to achieve posterior vitreous detachment (PVD) induction and liquefaction. Thus, many pharmacological agents have been studied with the aim of inducing PVD in order to facilitate the surgical procedure and reduce complications of vitrectomy. Tissue plasminogen activator, plasmin, microplasmin and hyaluronidase have shown an ability to induce PVD, when given as a single intravitreal injection in experimental and human studies [112].

Tissue plasminogen activator safety was assessed experimentally in both rabbits and cats. It was injected intravitreally (doses from 50 to 100 μg), and mild to severe vitreous inflammation with or without vitreous strands was observed. Reduced scotopic a-waves and b-waves on electroretinography as well as diffuse pigment alterations, severe photoreceptor losses coupled with RPE necrosis and proliferation with pigment clumping have been documented [113]. The clinical experience available points out that doses of 25–50 μg of intravitreal tissue plasminogen activator injection may be safe to the retina.

Microplasmin is a recombinant protein limited to the enzymatic moiety of plasmin without any of its cringle domains. Its enzymatic activity is similar to that of the enzyme plasmin. De Smet et al. [114] evaluated the vitreolytic ability in a porcine model with escalating doses of microplasmin (62.5, 125, 250, 400 μg) for 1 h or with 125 μg microplasmin with increasing time of exposure (15, 30, 60, 120 min). PVD was assessed by scanning electron microscopy, and microplasmin caused vitreolysis and posterior vitreous separation in an apparent dose- and time-dependent fashion. In this model system, the minimal effective dose appeared to be 125 μg and no signs of retinal toxicity were observed [114].

Intravitreal injection of recombinant microplasmin at doses from 12.5 to 250 μg in the rabbit induces no ERG or retinal ultrastructural abnormalities. Thrombogenics NV (Euronext Brussels, Leuven, Belgium), a biotechnology company focused on vascular disease, announced the results of the MIVI-I phase IIa trial (Microplasmin in Vitrectomy), which included 60 patients. Preliminary results available for all patients through 28 days after vitrectomy revealed that microplasmin was generally well tolerated even up to the highest dose tested (125 μg) [112].

Hyaluronidase cleaves the glycosidic bonds of hyaluronan as well as other mucopolysaccharides resulting in dissolution of hyaluronan and collagen complex and subsequent vitreous liquefaction. Intravitreal injection of hyaluronidase in doses of 20 IU or less does not appear to adversely affect the biomicroscopic morphology or function of ocular structures in a preclinical investigation in rabbits. A preservative-free, highly purified ovine hyaluronidase formulation (Vitrase, ISTA Pharmaceuticals, Irvine, Calif., USA) has been evaluated in phase III clinical trials. Intravitreal application of hyaluronidase was well tolerated, where dose-dependent iris was the most common adverse event, occurring in 62% of eyes treated with 55 IU and in 59% of eyes treated with 75 IU. Iritis was often self-limited and did not result in a serious adverse event in any hyaluronidase-treated eye [112]. Wang et al. [115] conducted an experimental study in diabetic and healthy rats, evaluating PVD induction after intravitreal injection with hyaluronidase, plasmin, hyaluronidase plus plasmin and BSS used as control. Scanning electron microscopy showed that complete PVD was found after hyaluronidase and plasmin in healthy rats. However, in diabetic rats, hyaluronidase alone was ineffective, plasmin alone induced partial PVD, and the combination of hyaluronidase and plasmin induced complete PVD. No obvious toxic reaction was observed in any group [115].

**Miscellaneous Anti-Inflammatory and Antiangiogenic Agents**

Some forms of drugs are characterized by a wide variety of actions due to their complex chemical structure and receptor affinity. Two examples of such drugs are thalidomide and MTX. Thalidomide is a glutamic acid derivative with proved antiangiogenic and anti-inflammatory efficacy in animal models of retinal ischemia, retinal neovascularization, uveitis and diabetic retinopathy. On the other hand, MTX is an antimetabolite and antifolate drug used in the treatment of cancer and autoimmune diseases. Due to their large variety of effects, the agents could be applied in different ophthalmic diseases; however, MTX may be indicated mainly in the therapy of primary intraocular lymphoma.
Few consecutive investigations have examined the safety of MTX in animals. As early as 1985, it has been shown that 600 µg MTX combined with other chemotherapeutic agents such as doxorubicin, 5-FU and bleomycin enhanced the risk of retinal toxicity [71]. However, in one recent study, the ocular pharmacokinetics and retinal toxicity of intravitreal MTX sodium at 400 µg were studied in New Zealand white rabbits. This research group reported that intravitreal MTX, combined with FU and dexamethasone, showed no evidence of retinal toxicity determined by electroretinography and histological examination [65]. Frenkel et al. [116] reported their 10-year experience with frequent intravitreal injection of MTX at 400 µg/0.1 ml. The effective agent rarely induced adverse reactions, the most common being self-limited corneal epitheliopathy, while the vitreoretinal involvement of lymphoma could be controlled effectively [116]. In another retrospective case series, complications that occurred during the period of treatment with intravitreal MTX included cataract (73%), corneal epitheliopathy (58%), maculopathy (42%), vitreous hemorrhage (8%) and optic atrophy (4%). However, no patient had irreversible loss of vision that could be definitely attributed to the intravitreal injections of MTX. Overall, the intravitreal use of MTX in doses up to 400 µg seems to be a safe approach in clinical practice. Although no preclinical investigation has been performed to investigate the retinal biocompatibility of thalidomide for local ocular application, one case report demonstrated corneal endothelium toxicity after oral use of the powerful agent [117].

Toxicity Unrelated to the Drug

Toxicity due to Preservatives

One of the main potential sources of bias in the assessment of retinal toxicity of certain pharmaceutical preparations is undoubtedly the adverse effect of the vehicle and preservatives.

Recently, many authors reported favorable therapeutic results after intravitreal injection of TA for the treatment of exudative age-related macular degeneration [118–122], diabetic macular edema [123, 124], proliferative diabetic retinopathy [125], macular edema due to central [126] and branch [127] retinal vein occlusion, and many other conditions. After the popularization of the intravenous administration of TA, the presence of benzyl alcohol (BA) in the Kenalog (Kenalog 40, Bristol-Myers Squibb) formulation and its deleterious effects in the intraocular environment became notorious [128–132].

Macky et al. [131] injected 0.1 ml of pure BA in the vitreous cavity of rabbits to assess its sole toxic properties. Evaluation included IOP measurements, slitlamp examination, indirect ophthalmoscopy, electroretinogram and electron microscopy for quantitative morphometric measurements. The mean amplitudes of the a- and b-waves of the BA-injected eyes were significantly reduced compared with the BSS-injected eyes and the noninjected contralateral eyes. These ERG responses continued to be significantly reduced in the BA-injected eyes (p < 0.01, t test) throughout the study period. The mean ganglion cell count was significantly reduced in the BA-injected eyes compared with the BSS-injected and noninjected eyes. Electron microscopy showed moderate to severe intracellular changes in the ganglion cell layer, inner nuclear cell layer, outer nuclear cell layer and photoreceptor layer at 6 weeks in BA-injected eyes, with no significant changes in the BSS-injected eye. There was no significant rise in the IOP or clinical evidence of increased lens density during the study period in any of the eyes.

Lang et al. [133] compared intravitreal injection of commercial Kenalog versus its vehicle alone, pure triamcinolone or BSS. Kenalog and its vehicle alone caused approximately 50% reduction in the ERG b-wave amplitude at the end of follow-up. Pure TA caused only mild (up to 14%) reduction of the ERG b-wave amplitude. Histological examination of the retina exposed to Kenalog or its vehicle showed severe damage to all retinal layers in areas close to the site of Kenalog injection. The authors of this study emphasized that a simple separation of the vehicle from the TA is not absolutely safe, and even a small amount of the vehicle may still be toxic. It has also been shown that following filtered or nonfiltered separation of triamcinolone from the vehicle, the concentration of BA increased to 3.6–4.3% (compared to 0.99% in commercial Kenalog suspension distributed in the USA) due to the high affinity of BA for a lipophilic environment such as TA crystals [132].

Li et al. [134] evaluated the toxic effects of two TA vehicles separated by centrifugation from commercially available preparations – Transton (vehicle A; Kunming Jida, China) and Kanacort-A (vehicle B; Bristol-Myers Squibb, Pakistan) – on rabbit retina at two different volumes (0.1 and 0.2 ml). Both vehicles A and B consisted of 9.9 mg/ml BA, 7.5 mg/ml sodium carboxymethylcellulose and 0.4 mg/ml polysorbate 80. Eyes with vehicle A appeared normal under the ophthalmoscope but showed disorganization in the retinal inner nuclear layer and photoreceptor layer in pathological analyses. Eyes with vehicle B disclosed more significant retinal changes in-

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including retinal hemorrhage, vascular narrowing, myelin-ated fiber edema, retinal necrosis and atrophy, and photoreceptor apoptosis. There was an increase in the degree of the above damages as the volume of either vehicle was increased.

Kai et al. [128] compared intravenous injections in rabbits of 4 or 25 mg of TA, with vehicle either reduced or not. Elevation of IOP was noted in all experimental groups after intravitreal TA. A significant uniformly distributed increase in lens density measured by a Pentacam system was noticed 1 week after intravitreal TA in the group administered 25 mg + vehicle. A significant loss of glutathione peroxidase activity was noticed at the end of the study, while superoxide dismutase activity increased. Since the function of glutathione peroxidase, which catalyzes the reduction of hydrogen peroxide to water, is to protect the lens from oxidative damage, the decrease in glutathione peroxidase activity may suggest damage of the antioxidative system of the lens. Amplitudes of ERG waves declined significantly in vehicle-treated groups at the end of the study. Pathological examination showed obvious retinal toxicity in vehicle-treated groups [128].

Formerly, Loewenstein et al. [135] had already reported the deleterious effects of preservatives in another frequently used commercial corticosteroid preparation, methylprednisolone acetate (Depo-Medrol®; Upjohn Company, Kalamazoo, Mich., USA). At that time, myristyl-γ-picolinium chloride was found to be the major toxic component of the formulation. When myristyl-γ-picolinium chloride solutions were injected at concentrations at least twice as high as that in Depo-Medrol, significant reductions in the light- and dark-adapted ERG responses were seen. The effect of the drug on the ERG responses was seen as early as 3 days after injection and developed to its maximal level within 1–2 weeks. No ERG recovery was seen over a period of more than 2 months. The visual evoked potential, elicited by applying light stimuli to the experimental eye, was characterized by low amplitude and delayed implicit time compared with the response obtained from the control eye [135].

Recently, Lüke et al. [136] studied retinal tolerance to bevacizumab in co-application with a recombinant tissue plasminogen activator (rtPA). During application of bevacizumab (0.25 mg/ml) in co-application with 20 mg/ml rtPA, the ERG amplitudes remained stable. The concentrations of rtPA alone (20 and 60 mg/ml) did not induce significant reduction of the b-wave amplitude. In addition, 20 mg/ml rtPA did not alter the a-wave amplitude. However, 60 mg/ml rtPA caused a slight but significant reduction in the a-wave amplitude. A full recovery was detected for both concentrations during the washout. At the highest tested concentration of 200 mg/ml rtPA, a significant reduction in the a- and b-wave amplitudes occurred during the exposure. The reduction of ERG amplitudes remained irreversible during the washout. The retinal toxicity of rtPA was thought to be due to the L-arginine component of the vehicle [136]. Further evidence for this hypothesis comes from a study of intravitreal injections of aztreonam that also contain L-arginine in the vehicle, which showed almost equivalent retinal damage when vehicle equivalent to the aztreonam dose was injected [137, 138].

Many other commercially available systemic medications could probably be useful for intraocular use, but every component of the vehicle or preservative can carry itself significant toxic potential, and much more research is still needed in this area.

Toxicity due to Osmolarity and pH Changes in the Retina

The group around Marmor [139–143] first reported retinal toxicity secondary to changes in osmolarity at the vitreomacular interface. They described cellular damage at the vitreoretinal interface, including nonspecific shrinkage and disruption of the cellular architecture caused by induction of osmotic levels higher than 500 mOsm. Despite the fact that differences in osmolarities between the subretinal space and choroid are corrected rapidly by the adjacent tissues, substances with nonphysiological osmolarity can produce retinal damage when in contact with the subretinal space, as in macular hole surgery [139–143]. Intravitreal dye injections may rapidly change the osmolarity in the vitreous cavity. Several in vivo and in vitro studies proposed that hypo-osmotic indocyanine green solutions could harm the RPE, and this effect could be augmented by additional intraoperative light exposure [144–146].

It is clear that homeostatic conditions are essential for the delicate intraocular environment, and many of the recently intraocularly administered off-label drugs lack an adequate adjustment to deal with intraocular pH and osmolarity requirements. Moreover, the lack of definite safe ranges for these parameters in the literature makes it even more difficult to predict whether a particular pharmacological preparation is toxic or not.

Anyway, it is obvious that extreme pH and osmolarity values for intraocular solutions, even when only a short exposure time is intended, should be definitely avoided.
**Final Conclusions**

Intravitreal pharmacotherapies have been used with increasing frequency in the treatment of retinal disease. Indications for their use include choroidal neovascular membranes, diabetic macular edema, ischemic neovascularization, inflammatory and infectious processes, and neoplasia. Complications of intravitreal therapies include cataract formation, glaucoma and endophthalmitis. Recent developments of pharmacological agents administered intravitreally and the new applications of systemic medications in retinal disease present the practitioner with expanded treatment options. Current and emerging data will help guide therapy in order to maximize the benefits and limit the systemic and ocular complications of these new treatment options.

Ocular toxicity caused by the exposure of the retinal tissue to a high concentration of drug for certain periods of time remains an important topic in the approach to intravitreal injection in the therapy of retinal diseases. In the evaluation of drug safety for retinal pharmacotherapy, both exposure concentration and exposure time can be important, but the toxic effect may also be directly related to the transient drug distribution inside the vitreous body. The retina is subject to the risk of drug-induced toxicities owing to its rich blood supply, complex neuroretinal organization and lifelong exposure to focused light rays. To deal with the risk of injury, the retina is protected by specific defense mechanisms including the endogenous cytochrome P450 detoxification system and the blood-retina barrier. An understanding of these cellular and molecular principles is a key aspect in elucidating the pathological pathways leading to retinotoxicity.

Currently, there are several approved drugs for intravitreal use, such as ranibizumab (Lucentis; Genentech) and pegaptanib (Macugen; Pfizer, New York, N.Y., USA). However, there are numerous off-label uses of drugs and substances injected into the eye, and the scientific literature remains confusing with regard to the intraocular properties and toxicology of off-label drugs such as commercial triamcinolone, antibiotics, antivirals and tissue plasminogen activator.

In the near future, several drugs that have already undergone preclinical evaluation and small case series in patients, and that are now tested in randomized clinical trials, will be available soon for use in clinical practice.

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**References**


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