Plasma Substance P Levels in Patients with Persistent Cough

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Key Words

Persistent cough · Asthma · Substance P · Capsaicin cough sensitivity · Airway responsiveness · Plasma

Abstract

Background: Substance P (SP) is involved in the pathogenesis of cough in animal models. However, few studies in humans have been reported and the roles of SP in clinical cough remain obscure. **Objectives:** To clarify the relevance of plasma levels of SP in patients with persistent cough. **Methods:** We studied 82 patients with cough persisting for at least 3 weeks and 15 healthy controls. Patients were classified as having asthmatic cough (cough-variant asthma and cough-predominant asthma; n = 61) or nonasthmatic cough (n = 21; postinfectious cough, n = 6; gastroesophageal reflux disease, n = 5; idiopathic cough, n = 5, and others, n = 5). Correlations were evaluated between plasma SP levels as measured with ELISA and methacholine airway hyperresponsiveness (airway sensitivity and airway reactivity), capsaicin cough sensitivity, sputum eosinophil and neutrophil counts, and pulmonary function. **Results:** Plasma SP levels were sig-

nificantly elevated in patients with both asthmatic and nonasthmatic cough compared with controls [31.1 pg/ml (range 18.0–52.2) and 30.0 pg/ml (range 15.1–50.3) vs. 15.4 pg/ml (range 11.3–23.7); p = 0.003 and p = 0.038, respectively] but did not differ between the two patient groups (p = 0.90). Plasma SP levels correlated with airway sensitivity (threshold dose of methacholine) in the patients with asthmatic cough (r = –0.37, p = 0.005) but not with airway reactivity, cough sensitivity, FEV₁ values, or sputum eosinophil and neutrophil counts in either group. **Conclusions:** Increased levels of SP in plasma are associated with persistent cough in humans and might be related to airway sensitivity in asthmatic cough.

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Introduction

Substance P (SP) is one of several neuropeptides that are widely distributed in sensory peripheral nerves [1] and in the central nervous system [2]. Ample evidence supports a role for SP in the mechanism of cough in ani-

mal models [3–5]. Although its activity in cough induction is controversial [3, 6], SP elicits a sensitizing effect on the cough reflex [4, 5], while conflicting results also exist [7].

A few studies have examined the relationship between SP and cough in humans. Patients with attenuated cough sensitivity associated with advanced Parkinson's disease and aspiration pneumonia have reduced SP levels in sputum [8, 9]. SP-immunoreactive nerve densities of the bronchial epithelium are increased in patients with cough-variant asthma (CVA) compared with those of patients with classic asthma and healthy controls [10]. Levels of SP are also increased in nasal lavage fluid [11] and sputum [12] from patients with nonasthmatic cough. Furthermore, elevated numbers of calcitonin gene-related peptide (CGRP)-immunoreactive nerves in the bronchial epithelium of patients with idiopathic persistent cough correlate with cough sensitivity to inhaled capsaicin, whereas levels of SP are not elevated or correlated with cough sensitivity [13]. Yoshihara et al. [14] have shown that plasma SP levels are elevated in patients with paroxysmal cough due to pertussis. However, further information about plasma SP levels in persistent cough of other etiologies has not been reported.

Cough is attributed to various causes [15–18]. Persistent cough due to asthmatic and nonasthmatic causes might involve different mechanisms since asthmatic cough is elicited by bronchoconstriction whereas nonasthmatic cough might be primarily ascribed to increased cough sensitivity. Although cough and bronchoconstriction often occur simultaneously, they are regarded as separate reflexes [19]. Bronchoconstrictors including methacholine provoke cough without altering the cough reflex [20].

Other than cough, evidence shows that SP functions in bronchoconstriction and airway hyperresponsiveness (AHR) [21, 22]. Therefore, SP might be differently involved in the mechanisms of both asthmatic and nonasthmatic cough. Indeed, SP contents in the nasal lavage fluid of patients with nonasthmatic cough are associated with increased cough sensitivity [11], whereas in asthma sputum levels of SP correlate with airflow obstruction [23].

Here, we compared plasma SP levels in patients with combined subacute cough (duration of 3–8 weeks) and chronic cough (>8 weeks) as defined by a guideline [15] with those of healthy controls. We also examined the relationship between plasma SP levels and various clinical and functional indices to determine the roles of SP in patients with asthmatic and nonasthmatic cough.

Subjects and Methods

Participants

We studied 82 consecutive patients who were referred to the outpatient asthma and cough clinic of Kyoto University Hospital between October 2007 and August 2009 because of cough that had persisted for at least 3 weeks. None of the patients had abnormal chest radiographic findings or had been prescribed angiotensin-converting enzyme inhibitors, oral or inhaled corticosteroids, leukotriene receptor antagonists, or other antiallergic drugs. None had been taking drugs that may interfere with circulating SP levels, such as antihistamines, angiotensin-converting enzyme inhibitors, or centrally acting drugs such as dopamine receptor agonists. All patients were either never-smokers or former smokers who had smoked less than 10 pack-years and had quit smoking more than 1 year prior to the study.

Causes of cough were determined according to Japanese cough guidelines [16]. In brief, patients with AHR to methacholine or reversible airflow obstruction and improvement of coughing with β_2 -agonists were considered as having chronic cough due to asthma. Patients with cough as the sole or predominant symptom (CVA or cough-predominant asthma) were included and were categorized as having asthmatic cough. The others were categorized as having nonasthmatic cough caused by the following [16]: sinobronchial syndrome (chronic sinusitis complicated by neutrophilic airway inflammation of the lower airways) [17, 24] diagnosed based on positive sinus images, and symptoms related to chronic sinusitis improved with macrolides; gastroesophageal reflux disease (GERD) based on response to treatment with proton pump inhibitors; postinfectious cough based on a history of upper respiratory tract infection followed by cough that spontaneously subsided; atopic cough based on findings suggesting an atopic predisposition or induced sputum eosinophilia as well as a response to antihistamines [16, 25]; cough due to pertussis based on the typical clinical course of pertussis and a positive antipertussis toxin antibody reaction, and idiopathic cough for which extensive examinations and intensive therapeutic trials were negative or failed to reveal any conclusive findings. The numbers of patients who underwent more specialized and detailed examinations or assessment were as follows: 9 for thoracic CT, 4 for sinus CT, 3 for esophageal endoscopy, 3 for bronchoscopy, and 3 for ENT consultations. To compare the plasma SP levels among different causes of subacute and chronic cough, patients with cough due to multiple causes were not included in the cohort who had asthma and GERD ($n = 4$), postinfectious cough and GERD ($n = 2$), asthma and sinobronchial syndrome ($n = 2$), or sinobronchial syndrome and GERD ($n = 1$).

We also studied 15 healthy controls recruited from our hospital staff who had no history of respiratory disease. We excluded individuals with atopic dermatitis from this study because its presence positively affects the plasma SP levels [26]. The Ethics Committee of Kyoto University approved the research protocol (approval No. E-300) and written informed consent was obtained from all participants.

Measurement of Plasma Levels of SP

Blood was sampled at presentation in all subjects. Samples were immediately centrifuged and plasma was mixed with an equal volume of a stabilizer (Kyowa Medex Co., Ltd., Nagaizumicho, Japan) that inhibits neutral endopeptidase [27]. The samples

Table 1. Characteristics of the three subject groups

	Asthmatic cough (n = 61)	Nonasthmatic cough (n = 21)	Healthy controls (n = 15)	p values ^a
Males, n	16 (26)	5 (24)	8 (53)	0.10
Age, years	52 (33–65)	49 (34–65)	43 (38–60)	0.9989
Former smokers, n	12 (20)	3 (14)	2 (13)	0.77
FEV ₁ /FVC, %	80.3 (74.5–84.8)	80.0 (75.7–85.4)	80.6 (78.8–84.0)	0.75
FEV ₁ , %predicted	101.2 (90.4–111.4)	101.5 (89.1–106.9)	100.6 (90.6–108.6)	0.92
FEF _{25–75} , %predicted	83.7 (67.5–105.7)	93.0 (74.2–108.9)	78.3 (73.9–100.0)	0.74
Sputum eosinophil ^b , %	1.1 (0.3–4.3)	0.4 (0.2–0.9)	0.0 (0.0–0.8)	0.040 ^c
Sputum neutrophil ^b , %	59.6 (43.3–79.3)	58.8 (45.9–84.6)	60.0 (42.8–84.1)	0.94

Data are expressed as medians (25th to 75th percentiles) or numbers (%).

^a Kruskal-Wallis or χ^2 test. ^b Sputum induction was successful in 38 patients with asthmatic cough, 10 patients with nonasthmatic cough, and 9 healthy controls. ^c Asthmatic cough versus control, $p = 0.07$; asthmatic cough versus nonasthmatic cough, $p = 0.27$; nonasthmatic cough versus control, $p = 0.51$ by the Steel-Dwass test.

were frozen at -20°C . Investigators who were blinded to the clinical conditions of the patients measured plasma SP levels using a competitive ELISA method [27, 28]. The sensitivity of this assay is 4.1 pg/ml. The specificity of the assay for SP measurement is 100%, and the assay does not significantly cross-react with neurokinin A or neurokinin B.

Sputum Induction and Processing

Sputum was induced and processed as described by Pin et al. [29] with slight modifications [30]. In brief, after pretreatment with salbutamol, sputum was induced by inhalation of hypertonic saline (3%) solution for 15 min from an ultrasonic nebulizer. Adequate plugs of sputum were treated with 0.1% dithiothreitol (Sputasol; Oxoid Ltd., Hampshire, UK) followed by Dulbecco's phosphate-buffered saline (PBS). Eosinophil and neutrophil percentages were determined by counting at least 400 nonsquamous cells on centrifuged preparations visualized by May-Grünwald-Giemsa staining.

Our primary purpose in evaluating sputum cells was to investigate the association of cellular inflammation of the airways with plasma levels of SP. Sputum cell differentials were also used for the diagnosis of disease, e.g. atopic cough.

Pulmonary Function Test

We measured forced vital capacity (FVC), FEV₁, and forced mid-expiratory flow (FEF_{25–75%}) using a Chestac-65V (Chest MI Corp., Tokyo, Japan) as described [31].

Methacholine Challenge Test

We determined AHR by measuring respiratory resistance (Rrs; cm H₂O/l/s) (AstographTM; Chest, Tokyo, Japan) under continuous methacholine inhalation as described [32, 33]. The index of airway sensitivity was Dmin, i.e. the cumulative dose of inhaled methacholine at the inflection point where Rrs started to continuously increase. One unit of Dmin is equivalent to a dose of 1 mg/ml of methacholine inhalation for 1 min. When the Rrs did not increase despite methacholine inhalation at the highest con-

centration, Dmin was assigned a value of 50 units, which was the total cumulative dose of methacholine. The slope of the respiratory dose-response curve was used as the measure of airway reactivity [33]. Fifty-nine patients with asthmatic cough and all 21 patients with nonasthmatic cough underwent the test.

Capsaicin Cough Sensitivity Test

Cough sensitivity was tested by continuous inhalation of capsaicin as described [34], with a slight modification of capsaicin concentrations [24]. Ten doubling concentrations of capsaicin solution (0.61–312 μM) were inhaled until ≥ 5 coughs were induced. Each concentration of capsaicin was inhaled for 15 s during tidal breathing every 60 s. The concentration of capsaicin causing ≥ 5 coughs is referred to as C5 [24, 34]. Fifty-nine patients with asthmatic cough and all 21 patients with nonasthmatic cough underwent the test.

Statistical Analysis

Data are expressed as median values (25th to 75th percentiles) except when noted otherwise and were analyzed using JMP 6.0 (SAS, Cary, N.C., USA). Comparisons of 2 and 3 groups were achieved using Mann-Whitney and Kruskal-Wallis tests, respectively, and the latter were analyzed post hoc using the Steel-Dwass test [35–37]. Categorical data were compared using the χ^2 test. Correlations between data were analyzed using Spearman's rank correlation test. $p < 0.05$ was considered statistically significant.

Results

Characteristics of the Three Groups

Table 1 shows the characteristics of the 61 patients with asthmatic cough, 21 patients with nonasthmatic cough, and 15 controls. Only the ratios of sputum eosinophils significantly differed among the three groups.

Table 2. Characteristics of the two patient groups

	Asthmatic cough (n = 61)	Nonasthmatic cough (n = 21)	p values ^a
Cough duration	12 (7–24)	8 (4–40)	0.18
Atopy ^b	38 (63)	11 (58)	0.67
Serum IgE ^c , U/ml	80.0 (28.0–190.0)	77.0 (10.0–400.0)	0.71
Dmin ^d , units	2.8 (1.4–6.6)	9.8 (2.9–29.0)	0.007
SRrs ^d			
cm H ₂ O/l/s/min	1.3 (0.8–2.4)	1.2 (0.5–2.2)	0.54
C5 ^e , μ M	9.8 (2.4–19.5)	2.4 (0.9–14.6)	0.07

Data are expressed as medians (25th to 75th percentiles) or numbers (%).

^a Mann-Whitney U test or χ^2 test.

^b Atopy was determined based on the presence of specific serum IgE antibodies to at least one common inhalant allergen, including cat dander, dog dander, weed pollens, grass pollens, molds, or house dust mites. Data are missing for 1 patient with asthmatic cough and 2 patients with nonasthmatic cough.

^c Data are missing for 2 patients each with asthmatic and nonasthmatic cough.

^d Data are missing for 2 patients with asthmatic cough. Two patients among those with asthmatic cough were not eligible for analysis of AHR.

^e Data are missing for 2 patients with asthmatic cough.

Cough in the nonasthmatic cough group was due to postinfection (n = 6), GERD (5), atopic cough (2), pertussis (2), and sinobronchial syndrome (1) but was idiopathic in 5 patients.

Outcomes in Patients with Asthmatic and Nonasthmatic Cough

Table 2 shows the outcomes of the two patient groups. Two of the patients with asthmatic cough were ineligible for AHR analysis since the inflection point where Rrs increased could not be determined because severe coughing was elicited. Patients with asthmatic cough were significantly more sensitive to methacholine as determined by Dmin than those with nonasthmatic cough. C5 was marginally lower in patients with nonasthmatic cough than in those with asthmatic cough.

Comparison of Plasma SP Levels among the Three Groups

Plasma SP levels were significantly higher in patients with asthmatic and nonasthmatic cough compared to healthy controls [31.1 pg/ml (range 18.0–52.2) and 30.0 pg/ml (range 15.1–50.3) vs. 15.4 pg/ml (range 11.3–23.7)]

Fig. 1. Comparison of plasma SP levels among groups. Plasma SP levels are elevated in patients with asthmatic and nonasthmatic cough compared with controls, while the two patient groups do not significantly differ.

but did not significantly differ between the two patient groups (fig. 1).

Relationships between Plasma SP Levels and Clinical Indices

Plasma SP levels significantly correlated with airway sensitivity determined by Dmin only in patients with asthmatic cough (table 3; fig. 2). Plasma SP levels did not correlate with cough duration, airway reactivity, capsaicin cough sensitivity, spirometric indices, or sputum neutrophil and eosinophil counts in either patient group (table 3).

Discussion

We measured plasma levels of SP in patients with subacute and chronic cough of various origins and healthy subjects. We discovered that plasma SP levels are elevated in patients with cough of asthmatic and nonasthmatic origins. We also found that plasma SP levels correlate with airway sensitivity in patients with asthmatic cough. These results indicate that SP is involved in both asthmatic and nonasthmatic cough, and its role in the mechanisms of these types of cough might differ.

SP synthesized in the cell body of C-fibers is transported along axons towards the peripheral and central terminals, where it is stored in large granular vesicles. C-

Fig. 2. Correlation between plasma SP levels and airway sensitivity. Plasma SP levels negatively correlate with Dmin in asthmatic cough but not in nonasthmatic cough. Logarithmic data are presented for Dmin.

fiber activation evokes SP release into the airway dependent on the axon reflex in guinea pigs. Airway SP causes bronchospasm, vasodilation, edema, and mucus secretion, which secondarily evokes the activation of rapidly adapting receptors (RARs) in the airway, resulting in an enhanced cough reflex [5, 6]. The sensitizing effect of SP on RARs has also been demonstrated in the central nervous system, especially in the nucleus tractus solitarius, which also results in an enhanced cough reflex [38, 39]. While airway SP levels or expression in patients with persistent cough are conflicting [10–13, 40, 41], we found elevated plasma SP levels in patients with persistent asthmatic and nonasthmatic cough.

We found that SP may be associated with airway sensitivity in asthmatic cough. Airway sensitivity and airway reactivity are two major components of AHR and might have different underlying mechanisms [33, 42]. Both airway sensitivity and reactivity are associated with SP. Umeno et al. [22] reported that intravenous SP increases airway sensitivity to histamine in guinea pigs and Cheung et al. [21] showed that inhaled SP enhances maximal airway narrowing to methacholine in patients with asthma. Airway sensitivity may be determined by the strength of the stimulus that triggers the airways to constrict, e.g. epithelial damage, neural control, and inflammatory cell numbers, while airway reactivity may be caused by the responsiveness of airways to applied stimuli such as smooth muscle contractility, viscous and elastic loads,

Table 3. Correlations between plasma SP levels and clinical indices in the two patient groups

	Asthmatic cough		Nonasthmatic cough	
	r	p	r	p
Cough duration	0.10	0.42	–0.24	0.29
Serum IgE (U/ml)	0.18	0.19	0.35	0.14
FEV ₁ /FVC (%)	–0.18	0.17	0.04	0.88
FEV ₁ (%predicted)	–0.02	0.88	0.12	0.62
FEF _{25–75} (%predicted)	–0.05	0.73	0.05	0.84
Dmin (units)	–0.37	0.005	0.08	0.73
SRrs (cm H ₂ O/l/s/min)	0.01	0.93	0.05	0.85
C5 (μM)	0.16	0.23	0.01	0.95
Sputum eosinophil (%)	0.15	0.36	0.09	0.80
Sputum neutrophil (%)	0.18	0.28	–0.39	0.26

and airway swelling [42]. Our results suggest that SP plays a role in the pathophysiology of asthmatic cough by affecting airway sensitivity.

Although levels of plasma SP were higher in patients with nonasthmatic cough than in healthy controls, we found no correlation between plasma SP levels and various indices including capsaicin cough sensitivity. This is in conflict with the findings of Cho et al. [11], who showed that SP levels in nasal lavage fluid correlate with capsaicin

cough sensitivity in patients with nonasthmatic cough. This discrepancy might be attributed to the different sample sources or the smaller sample size in our study. Indeed, although Cho et al. [11] found a correlation between SP in nasal lavage fluid and cough sensitivity in all 38 of their patients with nonasthmatic cough, no correlation was evident when the patients were separated into 2 equal, separately analyzed groups ($n = 19$ for each) with increased and normal cough sensitivity.

The two phenotypes of asthma might involve different pathophysiological mechanisms: CVA or cough-predominant asthma and classic asthma that predominantly presents with wheezing. De Diego et al. [43] reported that although classic asthma and CVA have similar profiles of airway inflammatory markers their relationships with AHR and cough sensitivity differ. Lee et al. [10] found increased SP-immunoreactive nerve densities in patients with CVA but not in those with classic asthma. SP is associated with neurogenic inflammation and subsequent airflow obstruction in asthma [23, 44]. We found no correlation between plasma SP levels and spirometric indices, which contradicts the findings of Tomaki et al. [23], who found a negative correlation between sputum SP levels and FEV₁/FVC in patients with classic asthma. This discrepancy might be attributed to the different sample sources and less prominent airflow obstruction in our patients (FEV₁/FVC of 80.3% by average) compared with those of Tomaki et al. [23] (71.3%).

Airway inflammation stimulates receptors of unmyelinated C-fibers of the vagus nerve, thus causing the release of tachykinins such as SP from the C-fibers [11, 45, 46]. A correlation between sputum SP levels and sputum eosinophil count has been reported in asthma [23]. Since patients with CVA show evidence of airway inflammation with increased eosinophils [47] and since nonasthmatic cough might be associated with increased neutrophils [48], we evaluated the correlation between plasma SP levels and sputum counts of eosinophils and neutrophils in each patient group. However, we found no correlation between plasma SP levels and these sputum cells in either patient group. Circulating levels of SP might not be related to cellular inflammation of the airways in patients with persistent cough.

Our study has some limitations. Firstly, we could not determine whether SP levels in plasma reflect those in the airways. Moreover, although SP is widely distributed in the central and peripheral nervous system, there is increasing evidence that it may be synthesized and released from inflammatory cells such as eosinophils, monocytes and macrophages, lymphocytes, and dendritic cells [49–

52]. Therefore, elevated levels of SP in plasma may reflect overexpression of SP in inflammatory cells as well as those in sensory nerves. Secondly, atopic dermatitis might have influenced the levels of SP in plasma [53]. Atopic dermatitis was present in 8 patients with asthmatic cough but in none of the patients with nonasthmatic cough. However, we found no difference in plasma SP levels between patients with and without atopic dermatitis ($p = 0.51$). Thirdly, the sample size of patients with nonasthmatic cough was small, and the etiology of their cough was diverse. We found no difference in plasma SP levels among the 3 most common diagnostic subgroups of nonasthmatic cough [postinfectious cough ($n = 6$), GERD ($n = 5$), and idiopathic cough ($n = 5$); $p = 0.196$ by Kruskal-Wallis test]. Future larger studies might clarify the roles of SP, especially in nonasthmatic cough. Fourthly, while the success rate of sputum induction in our previous series of 407 patients with classic asthma was 73.0% [54], the success rate in the present study was lower (57/97, 59%). This may be because cough due to CVA or nonasthmatic origins such as GERD is more dry or nonproductive in nature compared with the cough of classic asthma [55], and 'healthy' controls do not present with sputum by definition. Fifthly, patients diagnosed as not having GERD might actually have had GERD, because the diagnosis of GERD solely depended on the response to specific therapy not involving the gold standard for the diagnosis. The same is true for more uncommon causes of cough such as obstructive sleep apnea and tracheal collapse.

Despite these limitations, the elevated plasma SP levels in patients with asthmatic and nonasthmatic cough are notable. SP in plasma is associated with persistent cough and it might be related to airway sensitivity in asthmatic cough.

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