Diffuse Endocrine System, Neuroendocrine Tumors and Immunity: What’s New?

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Abstract
During the last two decades, research into the modulation of immunity by the neuroendocrine system has flourished, unravelling significant effects of several neuropeptides, including somatostatin (SRIH), and especially cortistatin (CST), on immune cells. Scientists have learnt that the diffuse neuroendocrine system can regulate the immune system at all its levels: innate immunity, adaptive immunity, and maintenance of immune tolerance. Compelling studies with animal models have demonstrated that some neuropeptides may be effective in treating inflammatory disorders, such as sepsis, and T helper 1-driven autoimmune diseases, like Crohn’s disease and rheumatoid arthritis. Here, the latest findings concerning the neuroendocrine control of the immune system are discussed, with emphasis on SRIH and CST. The second part of the review deals with the immune response to neuroendocrine tumors (NETs). The anti-NET immune response has been described in the last years and it is still being characterized, similarly to what is happening for several other types of cancer. In parallel with investigations addressing the mechanisms by which the immune system contrasts NET growth and spreading, ground-breaking clinical trials of dendritic cell vaccination as immunotherapy for metastatic NETs have shown in principle that the immune reaction to NETs can be exploited for treatment.

Introduction

The anatomical organization and the functions of the diffuse (neuro)endocrine and immune systems are exceedingly complicated, and the more we discover the more new questions arise. The diffuse endocrine system (DES) is composed of neuroendocrine cells scattered throughout the entire body, either isolated or grouped to form discrete aggregates, such as the neuroepithelial bodies in the bronchopulmonary tract or the islets of Langherans in the pancreas [1]. While at least some of the DES cells in the gut are known to regulate secretion, absorption, motility, and mucosal proliferation [2], the physiological role of the neuroendocrine cells in most other organs remains elusive.

The immune system includes two branches, i.e. innate immunity, which is evolutionarily older, and adaptive
immunity [3]. The complement, neutrophils, mast cells, eosinophils, and basophils are central to the first, lymphocytes are the cornerstone of the second, and phagocytic cells (monocyte/macrophages and dendritic cells, DCs) are at the interface between the two. Innate immunity provides generic, non-pathogen-specific, and – most notably – short-term protection. In contrast, the adaptive immunity’s defense is specific and lasts much longer (see also the relevant paragraph).

Historically, studies on the interactions between the DES and the immune system have been addressed to establish whether and how the first influences the latter. This research direction was taken as early as the DES began to be characterized, when neuroendocrine cells were found in lymphoid organs and neuroendocrine mediators (biogenic amines, neuropeptides) and their receptors in immune cells, suggesting the existence of a neuroendocrine modulation of immune functions, based on a biochemical language of autocrine, paracrine, and endocrine signals [4]. As exemplified by somatostatin receptors (SSTR) 1–5 (fig. 1) [5–12], the expression profile of many neuropeptide receptors varies across different types of immune cells, and changes upon their differentiation or activation status, further pointing to a fine regulation of immunity by neuropeptides [13]. For instance, human monocytes do not transcribe any SSTR gene in resting conditions, but express SSTR2A when activated with lipopolysaccharide, and both SSTR1 and SSTR2A when induced to differentiate into macrophages or DCs [7–9]. In vivo, SSTR2A is present on the membrane of macrophages and DCs [5, 11, 12], making tissues physiologically or pathologically rich of SSTR2A detectable.
by SSTR scintigraphy with radiolabelled somatostatin (SRIH) analogs having high affinity for SSTR2A, such as the routinely used $^{111}$In-pentetreotide (Octreoscan®) [12, 13].

The analysis of the expression profile of neuropeptides and their cognate receptors in immune cells has been paralleled by the study of animal models and, to a far lesser extent, human diseases. As a result, we have been able to partially understand the meaning of the biochemical language connecting the endocrine and immune systems.

**Regulation of Immunity by the DES**

**Innate Immunity**

Inflammation is a stereotyped, but essential, innate immunity response. Completing a previous, detailed research work on neuropeptide-induced inflammation [14], a SRIH-mediated anti-inflammatory loop has been fully characterized in rodents in recent times (fig. 2).

Trigeminal and dorsal root ganglia contain the cell bodies of capsaicin-sensitive sensory neurons, whose unmyelinated or thinly myelinated fibers transmit electrical impulses coding for pain to the central nervous system (CNS) [14]. The nociceptive nerve terminals express the transient receptor potential vanilloid 1 (TRPV1) protein, a cation channel which binds several ligands, including capsaicin. TRPV1-dependent and -independent stimulation of capsaicin-sensitive nerve endings triggers the release of substance P (SP) and calcitonin gene-related peptide (CGRP), that initiate inflammation: in this way, nociception is immediately coupled to a neurogenic inflammatory response against noxious, potentially tissue-damaging stimuli [14, 15]. In extreme cases, irritation of a deep organ, such as the colon, may lead to 'cross-sensitization' and neurogenic inflammation of another otherwise healthy organ, e.g. the bladder, via reflexes through axon collaterals in the spinal cord and/or in the brain [16]. Besides SP and CGRP, other neuropeptides can mediate neurogenic inflammation, such as neurokinin A [17]. Noteworthy, the same neuropeptides that drive neurogenic inflammation may be protective of tissue homeostasis in other contexts. For instance, in type I diabetes mellitus-prone NOD mice, a hypofunctional mutant TRPV1 receptor leads to depressed neurogenic inflammation, but also to $\beta$-cell stress and autoimmune insulitis, secondary to deficient release of SP into the pancreatic islets of Langerhans by TRPV1-expressing sensory neurons [18].

Capsaicin-sensitive, TRPV1-positive fibers contain also SRIH and release it upon TRPV1 receptor activation [19]. Neuronally-derived SRIH acts in a paracrine/autoimmune fashion, but also enters the circulation. In the same body area where it has been released, SRIH binds to presynaptic SSTRs and stops its own, SP, and CGRP exocytosis with negative feedback [14, 20]. Moreover, SRIH from peptidergic nerve endings can inhibit the pro-inflammatory responses of neutrophils, macrophages, lymphocytes [21], as well as of non-immune cells like vascu-
lar endothelial cells, smooth muscle cells [21], and intestinal epithelial cells [22]. This inhibitory activity of SRIH on the effectors of inflammation is both local and systemic, via the bloodstream; in the latter case, SRIH can operate as an anti-inflammatory hormone (fig. 2). SRIH might also contribute to the anti-inflammatory action of glucocorticoids, which have been reported to stimulate SRIH expression at the inflammatory site in rats [23]. Analgesia is another effect of SRIH, obtained by inhibiting peripheral nerve firing and acting within the CNS [14]. Clinically, SRIH analog-induced analgesia is effective in controlling headache in patients with acromegaly, and partly also cancer pain in advanced neuroendocrine tumors (NETs).

A significant anti-inflammatory effect of SRIH has been demonstrated in different animal models in which the experimental induction of inflammation is associated with capsaicin-sensitive nerve terminal stimulation, such as arthritis, subacute pneumonia, and contact dermatitis [19, 24]. Recently, elegant studies with SSTR4–/– animals have revealed that this SSTR subtype is crucial for SRIH anti-inflammatory effects in mice [24, 25].

Cortistatin (CST) is a neuropeptide with strong homology to SRIH, which can bind to SSTRs with the same high affinity as SRIH, and with lower affinity to type 1a growth hormone secretagogue receptor (GHS-R1a) and Mas-related gene (Mrg) X2 receptor [26], the latter being interestingly highly expressed in dorsal root ganglia [27]. CST has been shown to inhibit the production of pro-inflammatory cytokines and nitric oxide by activated macrophages, and stimulate the secretion of the anti-inflammatory cytokine interleukin (IL)-10, up to affecting circulating levels of these molecules and strikingly improve survival and clinical outcomes in mouse models of sepsis, Crohn’s disease, and rheumatoid arthritis [28–30]. Therefore, two different ligands can activate SSTRs to dampen inflammation. However, CST anti-inflammatory action appears to be stronger than that of SRIH, possibly because of CST binding to GHS-R1a (and hypothetically also to MrgX2). In animals, cyclosomatostatin, a SSTR-antagonist, fully blocked the anti-inflammatory effect of SRIH, but not of CST, while a GHS-R antagonist partially antagonized CST activity [28–30]. In addition, only CST inhibited basal and IL-1β-stimulated release of prostaglandin E2 by primary rat microglia, a population of CNS-resident macrophage-like cells [31]; thus, CST action on innate immunity cells might also be broader than that of SRIH. Noteworthy, CST, but not SRIH, mRNA can be detected in human monocyte-macrophages, and is up-regulated upon activation in parallel with SSTR2 mRNA, suggesting the existence of an autocrine CST-SSTR2 loop (fig. 1) [7, 8].

In protecting mice exposed to lipopolysaccharide from endotoxemia, CST was synergic with vasoactive intestinal polypeptide (VIP) [28], which has also been shown to inhibit macrophage and microglia release of pro-inflammatory cytokines, chemokines, free radicals, nitric oxide, and prostaglandin E2, and to stimulate the secretion of IL-10 [32, 33]. Both VIP and ghrelin also blocked lipopolysaccharide-activated macrophages from secreting high mobility group box 1 (HMBG1), a DNA-binding protein considered a necessary and sufficient late mediator of multiple organ failure in advanced sepsis [34].

Sepsis is probably the best scenario to pinpoint the strength of the anti-inflammatory activity of neuroendocrine mediators, since it results merely from an excessive systemic inflammatory response to infectious agents. Those neuropeptides which also have antimicrobial activities, such as ghrelin [35], are expected to be the most protective in sepsis, because of their capability of inhibiting not only inflammation, but also the replication of the microorganisms against which inflammation is mounted. Eventually, this category of neuropeptides, or even better their analogs, might be designed for therapy [36].

Despite the abundance of experimental data, and the report of increased SRIH-like immunoreactivity in the blood of patients with sepsis or undergoing surgery [37], there is yet no direct evidence that SRIH from capsaicin-sensitive nerves or CST from other sources inhibit inflammation also in humans, and that lack of SRIH and/or CST feedback may lead to increased susceptibility to developing more severe inflammatory reactions.

More in general, it has been hypothesized that in humans inflammation activates the DES, because increased serum values of chromogranin A (CgA) are found in patients with chronic inflammatory disorders, such as inflammatory bowel diseases [38], systemic lupus erythematosus [39], and bronchitis [40]. Tumor necrosis factor-α (TNF-α) might be the main stimulus for neuroendocrine cell activation, since concentrations of CgA, TNF-α, and its soluble receptors are positively correlated [38, 41]. Interestingly, CgA and its breakdown products have been shown to protect the endothelium from TNF-α-induced inflammatory changes [42]. However, the increase in CgA in human inflammatory diseases may reflect reactive hyperplasia of the DES or hyperactivity of the adrenal medulla, rather than a finalized feedback neuroendocrine response to inflammation [38, 39]. Neither implies the raise of CgA
levels that other neuropeptides and/or amines are secreted and somehow involved in tuning the ongoing inflammation.

**Adaptive Immunity**

In superior species, the immune system is able to expand different subsets of T cells for every pathogen, which will then be attacked in a specific manner. Antigen-specific clones of T lymphocytes undergo activation and expansion when they recognize peptides derived from a pathogen, properly presented with major histocompatibility complex (also known as human leukocyte antigen, HLA) molecules by antigen-presenting cells, especially DCs. This capability of adaptation, referred to as adaptive immunity, ensures a much higher level of protection than first-line innate immunity [3]. CD4+ T helper (Th) lymphocytes orchestrate the adaptive immune response, by secreting different cytokine patterns according to the stimuli received during their activation, including those brought on by neuropeptides [43]. For instance, it has been demonstrated in mice that SRIH and VIP favor the development and maintenance of a Th2 response, typically involved in helminthic infections and allergy, over a Th1 one, which instead occurs in the presence of intracellular pathogens [32, 44]. This modulation seems to be relevant in humans too, since in more than 300 children serum levels of SRIH and VIP have been associated with circulating Th2 cytokines, expression of Th2 transcription factors in T cells, and allergic sensitization [45]. Exploiting the potential of some neuropeptides of determining a Th2 bias might be of value to treat Th1-driven autoimmune processes, such as sarcoidosis [13], multiple sclerosis [46], Crohn’s disease, and rheumatoid arthritis [32]. In animal models of the latter two, CST selectively inhibited the release of Th1 cytokines by CD4+ lymphocytes infiltrating the colon or joints, respectively, and the draining lymph nodes, without affecting the levels of the Th2 signature cytokine IL-4 [29, 30].

Adaptive immunity is achieved through the activity of T cells against foreign or abnormal (non-self) cells which are to be destroyed. When lymphocytes reactive against self-tissues, which are to be ‘tolerated’ and preserved, are generated and/or not suppressed, the system becomes potentially harmful to the host itself [32, 47]. In general, autoimmunity is underlain by ineffective control of autoreactive T cells, a function for which regulatory T cells (Treg) play a primary role. Treg are lymphocytes shown to be crucial in sustaining self-tolerance and preventing the development of autoimmune processes by inhibiting autoreactive effector cells [47]. The most extensively characterized Treg population expresses the surface marker CD25 (in addition to CD4) and the transcription factor FoxP3 [48].

Naïve T cells in the periphery can be converted into Treg upon particular conditions, such as high concentrations of transforming growth factor (TGF)-β [49]. As reviewed by Gonzalez-Rey and colleagues [36], it has been discovered recently that neuropeptides can induce Treg and restore immune tolerance in autoimmune diseases. VIP is the most interesting molecule analyzed so far, because its ability to generate Treg has been proved in primary human T cells [50], and, most notably, in patients affected by sarcoidosis [51]. Although not as thorough as for VIP, evidence has been gained that other neuropeptides, including CST, can expand/generate Treg [29, 30, 32]. Treg induction might be the conceptual basis for preventing the rejection of allogeneic transplants with neuropeptides, through the stimulation of the recipient’s immune tolerance to alloantigens (i.e. non-self-antigens of the donor tissue) [52]. In a mouse model of skin transplantation, treatment with CST, but not with the SRIH analog octreotide, almost doubled the survival time of the allograft, and increased CD4+ spleen lymphocytes with a regulatory phenotype [53].

Immune self-tolerance is firstly produced in the thymus, the primary lymphoid organ where bone marrow-derived thymocytes mature to T lymphocytes (fig. 1). In the thymus, autoreactive T lymphocytes undergo negative selection and elimination [4], and a subset of Treg is generated [47]. Several neuropeptides are present in the thymus, released by sympathetic and parasympathetic nerve fibers, thymic epithelial cells, macrophages, DCs, and lymphocytes themselves [6, 12, 54]. In different animal species and in cell culture systems, some of these neuropeptides, such as SRIH and SP, respectively, have proapoptotic or mitogenic effects on thymocytes [6, 55], which makes them potentially involved in causing the survival of self-tolerant and the death of autoreactive T cells.

**Immune Response to NETs: The Other Side of the Coin**

Growing attention is being given to the immune response to neoplasms, induced by tumor-associated antigens (TAA), in the light of both a better understanding of tumor biology and technical advances that have made cancer immunotherapy a feasible treatment perspective [56, 57].
Endocrinologists and oncologists have known that immunology may be a key to improve NET management since the introduction of interferon (IFN)-α biotherapy, which directly inhibits NET cell cycle progression and hormone synthesis, reduces neoangiogenesis, and activates immune cells [58]. Other IFN molecules, such as IFN-β, may have similar or more potent effects than IFN-α [59].

Indirect evidence that a functional immune system reacts to growing NETs, and in principle slows down or halts their progression, comes also from the observation that patients affected by X-linked hyperimmunoglobulin M syndrome (XHIGM), a primary immunodeficiency disorder with defective B- and T-cell functioning, can develop neuroendocrine carcinoma (NEC) [60]. Erdos and colleagues [60] recently reported a 7-year-old child who died from a poorly-differentiated NEC, retrieved six further published cases of NECs in XHIGM, and found a total of 4 poorly-differentiated NECs in XHIGM out of 6,965 patients registered in the European Society for Immunodeficiencies online database. Although an estimate cannot be equated with reality, an incidence of NECs in XHIGM of 57.4/100,000 can be extrapolated from the ratio 4/6,965, and it might be even higher, since the European database includes other primary immunodeficiencies besides XHIGM.

That an immune response opposes to NETs is also substantiated by the frequent finding of lymphocyte infiltration of NETs, as assessed by immunohistochemistry for CD3 (a general T lymphocyte marker), CD4, and CD8 [61–63]. In the biggest case series analyzed so far, CD3+ T cells were found to infiltrate 68% of 87 well-differentiated pancreatic NETs. In patients with intermediate-grade tumors who were followed up for more than 5 years, CD3+ T cell infiltration was a significant univariate predictor of improved recurrence-free survival following surgical resection [63]. CD3+ infiltration was also found in 97% of 39 resected NET liver metastases, the pancreas being the most common verified primary site [63].

Carcinoid TAA-specific CD8+ T cells have been described in patients with midgut carcinoid tumors [64, 65]. The highest CD8+ T cell recognition was for the TAA CgA, which makes it an attractive target for immunotherapy. Interestingly, IFN-γ secretion by CD8+ lymphocytes in response to TAA was significantly higher in patients with a low tumor burden, while those who did not show recognition of any CgA-derived epitope had either very high or low plasma CgA levels. Based on this finding, the authors of the study argue that a minimal tumor bulk is necessary to activate T cells, and there is a threshold of NET growth beyond which T cells are anergized or deleted [65].

A subpopulation of DCs with natural killer properties (CD56 positivity, direct cell lysis via tumor necrosis factor-related apoptosis-inducing ligand, activation of T cells) can be generated in vitro by incubating monocytes with IFN-α [66]. In humans, precursors of these DC subtype might be CD14+/CD56+ monocytes, which are characterized by upregulation of costimulatory molecules chiefly involved in T cell activation, and cytolytic activity toward tumor cells upon stimulation with IFN-α [67]. Significantly more CD14+/CD56+ monocytes have been reported in 4 NEC patients than in healthy control subjects, being >5% of all monocytes in 3 cases [67].

As other cancer types, NETs often elude immunosurveillance regardless of the functionality of the immune system: clinicians’ experience is that NETs will eventually progress and/or relapse in most immunocompetent patients, if no therapy is given. As illustrated in figure 3, NET cells may escape the immune response in many ways [56].

An altered expression of HLA class I molecules has been demonstrated by immunohistochemistry in 10 out of 11 pancreatic NETs, loss of β2-microglobulin being the most frequent alteration [61]. In the absence of HLA class I antigens, TAA-derived peptides cannot be presented to T cells and the development of a tumor-specific immune response is hindered.

Immune cells can be inhibited by soluble factors secreted by NETs. Generation of DCs from umbilical cord blood precursors was completely abrogated if the latter were incubated with cell-free supernatant from bronchial carcinoid tumor cells, which are therefore likely to produce molecules blocking DC differentiation. Consistently, no CD1a and CD83-positive DCs were found in pulmonary carcinoid tumor samples [68]. Also non-tumor stromal cells may release immune inhibitory factors.

Anti-tumor immune responses can be limited by Treg like any immune response. A heavier tumor infiltration by Treg is associated with weaker anti-tumor immunity and thus worse prognosis [69]. Treg are induced or recruited into the tumor by cytokines and chemokines secreted by cancer cells, tumor-associated macrophages, and DCs, such as TGF-β and CCL22, respectively (fig. 3); TAA then drive their expansion [69].

The number of circulating Treg was significantly higher in 68 patients with midgut carcinoid tumors than in 53 healthy blood donors [62]. In a subgroup of 25 patients compared with 18 controls, serum levels of the cytotoxic T lymphocyte-promoting cytokines IL-12p70 and...
IL-1β were significantly lower, and concentrations of the Treg cytokine TGF-β were increased, even though not significantly. After polyclonal activation, patients’ T cells proliferated less than those from controls, possibly because of an anergized state caused by carcinoid-associated Treg [62]. NET-infiltrating Treg have been detected by immunohistochemistry for single Treg markers [62, 63], even though such an approach is not optimal to identify Treg and no Treg within NETs have been reported too [61]. Treg infiltration tended to be heavier in metastases than in primary carcinoid tumors [62]. In NET liver metastases (most from pancreatic NETs) it was significantly denser in intermediate/high-grade than low-grade lesions and correlated with a shorter overall survival in univariate analysis [63].

In Merkel cell carcinoma, a higher number of intratumoral mast cells was a predictor of shorter survival. A possible explanation is that mast cells may also stimulate immunosuppression, along with angiogenesis, tumor proliferation, and extracellular matrix degradation [70].

Even before most of the aforementioned data were available, pioneering reports about DC vaccination NETs were published [71, 72]. Briefly, for DC vaccination DCs are generated and/or purified ex vivo, loaded with TAA, and administered back to a patient. DCs pulsed with TAA should cause the activation of tumor-specific T cells, and eventually induce an anti-tumor response reducing or arresting tumor outgrowth; this really happens, but to a variable extent, largely depending on technical issues as the choice of the TAA, the way they are delivered into DCs, the route and schedule of TAA-loaded DC administration (for a review, see [44]). Even though few patients, especially affected by medullary thyroid carcinoma [72, 73], have received DC immunotherapy for NETs so far, the field is rapidly expanding and great hopes are pinned on it [74, 75].

Conclusions

Although a clear, comprehensive picture of how the DES and the immune system influence each other has not yet been established, our knowledge has remarkably grown. Research on animal models has shed light on the physiological, significant modulation of immune functions by neuropeptides, and has opened exciting therapy perspectives. These mainly concern neuropeptides such as VIP or CST, which can inhibit exuberant and thus det-
rimental responses of both innate and adaptive immu-
nity. Nevertheless, the puzzle is far from being solved. For
instance, VIP-deficient mice have recently been found to
be resistant to experimental autoimmune encephalitis, a
model of multiple sclerosis, because VIP exerts an unan-
ticipated, permissive role in the CD4+ T cell infiltration
of the CNS [76].

In contrast to the wealth of information from basic and
translational studies, there is very little evidence that the
preclinical findings about DES-immune interactions ap-
ply to humans. Neither can data from animals be trans-
lated into the human situation without verification, be-
cause of differences between species in the expression pat-
tern of neuropeptides and their receptors. As for SSTRs
and SRIH/CST, SSTR4 is abundantly expressed in rodent
immune cells [21], its expression in human immune cells,
if any, has however been reported to be negligible [10, 11],
but for one immunohistochemistry study [21].

The awareness that the immune system reacts to the
development of any tumor has prompted the study of the
immune response to NETs. Although promising, the pre-
liminary, small-sized clinical trials of DC vaccination
have left open many issues about the optimal immuniza-
tion strategy, which should be defined before proceeding
to a direct comparison with the current best treatment
options for NETs. Further research on the immune reac-
tion to NETs may also lead to the identification of new
prognostic markers (e.g. the degree of lymphocyte infil-
tration) and the establishment of other types of immuno-
therapy, such as adoptive tumor-specific T cell transfer
or targeting of tumor-associated Treg.

Disclosure Statement

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