Current Management of Patients with Severe von Willebrand Disease Type 3: A 2012 Update

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Introduction

Von Willebrand disease (VWD) is a bleeding disorder resulting from inherited defects in von Willebrand factor (VWF) [1, 2]. VWF is a plasma glycoprotein that mediates platelet adhesion to injured blood vessel sites and promotes platelet aggregation [3, 4]. In addition, VWF binds factor VIII (FVIII), an essential cofactor of factor IXa, and protects it from degradation by plasma proteases, thus prolonging its half-life [3, 4]. VWF is present in the form of multimers of up to 20,000 kDa composed of a different number of disulfide-linked dimers, with the largest multimers having the highest hemostatic potential [5, 6]. Type 1 VWD (VWD1) results from a partial quantitative deficiency of VWF and accounts for 60–70% of VWD cases; it is inherited mostly as an autosomal dominant trait [4]. The qualitative defects of VWF are classified according to phenotype in 4 different types, 2A, 2B, 2M and 2N [2]. Type 3 VWD (VWD3) is due to an almost complete deficiency of VWF [7]. VWD3 is inherited as an autosomal recessive trait, with heterozygous carriers being asymptomatic or exhibiting mild bleeding [1, 8, 9]. The disease is characterized by undetectable levels of VWF antigen (VWF:Ag) and reduced concentrations of FVIII (<20 IU/dl) [1, 4]. The first case of VWD3 was de-
scribed by Erik von Willebrand in Finland in 1926, and, after similar cases were reported during the 1950s, Inga Nilsson et al. [10] confirmed in 1957 that the same disease was found in Sweden. VWD3 is a rare disorder, affecting 0.1–5.3 people per million [11–13]. The incidence in Europe and North America is comparable (1.51 and 1.38 per million, respectively), whereas the incidence amongst the Arab community is higher (5.3 per million).

**Clinical Markers of VWD3**

Patients with VWD3 usually meet all 3 main criteria required for the diagnosis of VWD: a history of bleeding episodes since childhood, reduced plasma VWF and autosomal recessive inheritance [14]. Clinical markers are very important and include a negative history of bleeding in parents in most families because of the autosomal recessive inheritance and moderate to severe bleeding episodes in most cases. However, some patients with VWD3 may have relatively mild symptoms, despite the VWF deficiency, probably due to residual levels of VWF and/or individual FVIII levels [15, 16]. Because VWF deficiency results in the rapid degradation and depletion of FVIII, in VWD3 both primary and secondary hemostasis is affected. The main clinical manifestations are disproportionate mucocutaneous bleeding, severe menorrhagia and hemophilia-like symptoms, such as muscle hematomas and joint bleeding [1, 14, 17]. The most common sites of bleeding are the nose, gingiva and gastrointestinal tract [18]. In VWD3 patients with very low FVIII levels, postoperative bleeding is often observed [19]. Patients with joint bleeding are at risk of long-term joint damage, which can limit patient mobility and independence [20].

Few reports have described VWD3 symptoms in detail after Silwer’s first report [18]. The prevalence of VWD3 symptoms has been investigated in Iranian [17] and Italian [19] populations of patients in particular. The most common symptoms in these patients were epistaxis (74–77% of cases), menorrhagia (32–69% of women) and bleeding following tooth extractions (53–70% of cases). Hematoma and joint bleeding occurred in 31–52% and in 37–42% of patients, respectively [17, 19].

A bleeding severity score can be useful to predict the risk of bleeding in VWD including VWD3. The severity of each bleeding symptom is graded from 0 (no/trivial bleeding) to 4 [blood transfusion/replacement therapy/desmopressin (DDAVP)], and an additional grade, −1, assigned to clinical situations associated with a high bleeding risk (e.g. tooth extraction, surgery or delivery) in which bleeding did not occur, despite no antihemorrhagic prophylaxis [21]. A bleeding severity score for 12 bleeding symptoms was calculated for 154 families in which VWD1 occurred (affected and unaffected members) and for 200 healthy individuals [21]. The value of this tool has also been evaluated in patients with other VWD types, such as VWD2B [22] and VWD1 with increased clearance [23]. This approach might also be useful in VWD3, and it will be introduced in the prospective study entitled ‘Type 3 von Willebrand Disease International Registries and Inhibitor Prospective Study (3WINTERS-IPS)’ organized on behalf of the European Group on VWD3 and approved by the VWF subcommittee of the International Society on Thrombosis and Haemostasis Scientific and Standardization Committee and the European Association of Hemophilia and Allied Disorders.

**Laboratory Markers of VWD3**

VWD3 can be more easily diagnosed than other VWD types due to the severity of the VWF and FVIII deficiency. Whereas in VWD1, VWF is commonly at the lower end of the normal range (with approximately 95% of plasma VWF levels between 50 and 200 IU/dl [8, 24]), in VWD3, VWF is virtually absent. Furthermore, the FVIII level is usually 2–20 IU/dl in patients with VWD3 (FVIII normal range: 50–150 IU/dl) [1]. In addition, abnormal hemostasis test results can suggest the presence of VWD3. The hallmark of VWD3 is a markedly prolonged bleeding time, sometimes >35 min, which is due to the extremely low levels of VWF in both the plasma and platelets. Partial thromboplastin time is also prolonged due to the low concentrations of FVIII, although platelet count is normal [25]. In patients with very low VWF and FVIII levels who have a prolonged bleeding time and partial thromboplastin time, plasma levels of VWF:Ag should be always measured to confirm the diagnosis of VWD3 and to distinguish this VWD type from patients affected by severe forms of VWD1; concentrations below 1 IU/dl are usually considered ‘undetectable’ even if they are measurable by currently-used high-sensitivity methods (e.g. ELISA) [19]. The presence of discrete amounts of VWF:Ag circulating in VWD3 patients is still under investigation; the role of platelet VWF in VWD3 should be analyzed further. In addition, in the plasma of some patients with VWD3, VWF might be found predominantly in the form of monomers, and multimers cannot be detected by agarose gel electrophoresis [19].
Due to a lack of detectable VWF:Ag, traditional VWF activity measurements for ristocetin cofactor (VWF:RCo) and collagen-binding assay (VWF:CB) are also undetectable. However, in most laboratories, these assays are less sensitive than the VWF ELISA, and so results may be reported as <5 or <10 IU/dl.

**Molecular Markers of VWD**

Molecular biology techniques have identified several VWF gene defects that can cause VWD3, including large gene deletions, nonsense mutations, small deletions/insertions or frameshift mutations, splice-site mutations and missense mutations.

Complete VWF gene deletions are considered rare. A complete homozygous VWF gene deletion was identified in 2 patients with VWD3 [26] and in 4 family members with severe VWD [27]. The latter study showed that seven asymptomatic individuals from 2 families were heterozygous for the complete deletion [27]. A complete deletion (Delta253k) was identified in homozygous, compound heterozygous and heterozygous members of 2 nonconsanguineous families from Germany and Italy [28], confirming previous reports of complete deletion in families in these countries [16, 29]. Seven partial homozygous VWF gene deletions including exons 1–3, 6–16, 42, 33–38, 22–43, 23–52 and 17–18 have been described [30–35]. Of these, a deletion of exons 1–3 resulting from an Alu-mediated recombination event was found to be the most common defect in Hungarian patients with VWD3 [32].

Most VWF defects are caused by single nucleotide substitutions or small deletions/insertions. More than 30 different nonsense mutations have been identified by DNA sequencing and various screening techniques, the majority of which cause a frame shift resulting in a premature stop codon [36].

Over 15 splice-site mutations have been reported, half of which affect the guanine at position +1 or –1 [37]. However, only 2 of these mutations have been confirmed at the mRNA level [16, 38].

Missense mutations have also been reported in patients with VWD3, yet it is not clear how these mutations could cause the low VWF level in plasma. Some of the approximately 30 missense mutations found so far may have a common mechanism of impairing VWF function, such as preventing VWF dimerization [37].

**Treatment and Prevention of Bleeding in VWD3**

The aim of treatment for VWD is to correct the defect in platelet adhesion that is due to low/defective VWF and the abnormal coagulation that is due to low levels of FVIII. There are 2 main treatments available for VWD: stimulation of the release of endogenous VWF from endothelial cells with desmopressin and supplementation with plasma-derived VWF/FVIII-containing concentrates.

By definition, patients with VWD3 are considered unresponsive to desmopressin as successful treatment depends upon endogenous synthesis of functional VWF, which is impaired in these patients [39–41]. Therefore this therapy is not recommended, although a small group of patients (reported to have VWD3) responded to desmopressin treatment with an increase in VWF and FVIII [15]. It would be interesting if these patients could be enrolled into the current prospective study on VWD3 (3WINTERS-IPS) [42] in order to be able to evaluate their clinical, laboratory and molecular markers to confirm VWD3 diagnosis.

**VWF/FVIII Concentrates**

The first-line treatment for VWD3 is replacement therapy with plasma-derived VWF/FVIII concentrates. These can be given to stop bleeding episodes when they occur (treatment on demand), to prevent bleeding during surgery (prophylaxis for surgery) or to prevent recurrent bleeding at specific sites (secondary long-term prophylaxis). The minimum requirements for plasma-derived VWF/FVIII concentrates for the management of VWD are: (1) they must contain enough biologically active VWF to correct the primary hemostasis defect and to stabilize the endogenous FVIII molecule (the latter objective can be achieved independently of the content of exogenous FVIII), (2) they should be treated by virucidal methods, (3) before clinical use, their pharmacokinetics and efficacy should be tested in retrospective or prospective clinical trials in relatively large numbers of VWD patients. Only a few of the many VWF/FVIII concentrates available on the market meet these requirements (table 1).

Alphanate® (Grifols Biologicals Inc., Los Angeles, Calif., USA) effectively stopped active bleeding and provided adequate hemostasis for surgical procedures in a multicenter, prospective study in 81 patients with VWD, 32 of whom had VWD3 [43]. The plasma concentrations of VWF:RCo and FVIII increased to normal levels 15 min...
Table 1. Characteristics of available VWF/FVIII concentrates

A Concentrates with published VWF activity assessed in a relatively large number of VWD subjects

<table>
<thead>
<tr>
<th>Concentrate</th>
<th>Published studies demonstrating activity in VWD patients</th>
<th>Purification procedures</th>
<th>Viral inactivation methods</th>
<th>VWF: RCo/Ag</th>
<th>VWF: RCo/FVIII</th>
<th>Available in</th>
<th>Manufacturer</th>
</tr>
</thead>
<tbody>
<tr>
<td>Alphanate</td>
<td>Federici et al. 2010 [46]</td>
<td>heparin ligand CT</td>
<td>SD; dry heat</td>
<td>0.6</td>
<td>1.2</td>
<td>GE, IT, UK, USA</td>
<td>Grifols (US)</td>
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<td></td>
<td>Rivard et al. 2008 [90]</td>
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<td></td>
<td>Mannucci et al. 2002 [43]</td>
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<tr>
<td>Biostate</td>
<td>Howman et al. 2011 [57]</td>
<td>precipitation, heparin ligand CT</td>
<td>SD; dry heat</td>
<td>0.8</td>
<td>2.0</td>
<td>AU, ASIA</td>
<td>CSL Behring</td>
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<td></td>
<td>Dunkley et al. 2010 [56]</td>
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<td>Fanhdi</td>
<td>Federici et al. 2010 [46]</td>
<td>precipitation, heparin ligand CT</td>
<td>SD; dry heat</td>
<td>0.6</td>
<td>1.6</td>
<td>SP, IT</td>
<td>Grifols (SP)</td>
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<td></td>
<td>Hernandez-Navarro et al. 2008 [91]</td>
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<td>Bello et al. 2007 [45]</td>
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<td></td>
<td>Federici et al. 2002 [44]</td>
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<tr>
<td>Haemate-Pb</td>
<td>Halimeh et al. 2011 [81]</td>
<td>polyelectrolyte precipitations</td>
<td>Pasteurization</td>
<td>0.8</td>
<td>2.5</td>
<td>ASIA, EUR, USA</td>
<td>CSL Behring</td>
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<td></td>
<td>Gill et al. 2011 [52]</td>
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<td></td>
<td>Federici et al. 2007 [41]</td>
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<td></td>
<td>Coppola et al. 2006 [92]</td>
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<td>Thompson et al. 2004 [51]</td>
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<td>Michiels et al. 2004 [93]</td>
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<td>Gill et al. 2003 [50]</td>
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<td>Franchini et al. 2003 [49]</td>
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<td>Lillicrap et al. 2002 [48]</td>
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<td>Lubetsky et al. 1999 [94]</td>
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<td>Wilate</td>
<td>Berntorp et al. 2009 [60]</td>
<td>affinity CT, size exclusion</td>
<td>SD; dry heat</td>
<td>0.7</td>
<td>0.8</td>
<td>GE</td>
<td>Octapharma</td>
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<td></td>
<td>Halimeh et al. 2011 [81]</td>
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<td></td>
<td>Kessler et al. 2011 [95]</td>
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<td>Windyga et al. 2011 [61]</td>
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<td>Wilfactin</td>
<td>Borel-Derlon et al. 2007 [59]</td>
<td>ion-exchange, affinity CTs</td>
<td>SD; NF; dry heat</td>
<td>0.7</td>
<td>60</td>
<td>FR</td>
<td>LFB (Lille)</td>
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<td></td>
<td>Goudemand et al. 2005 [58]</td>
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</table>

B Concentrates with limited VWF activity and/or published studies in a relatively small number of VWD subjects

<table>
<thead>
<tr>
<th>Concentrate</th>
<th>Purification procedures</th>
<th>Viral inactivation methods</th>
<th>VWF: RCo/Ag</th>
<th>VWF: RCo/FVIII</th>
<th>Available in</th>
<th>Manufacturer</th>
</tr>
</thead>
<tbody>
<tr>
<td>Emoclot</td>
<td>ion-exchange CT</td>
<td>SD; dry heat</td>
<td>0.5</td>
<td>1.2</td>
<td>BR, IT</td>
<td>Kedrion</td>
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<tr>
<td>Immune</td>
<td>ion-exchange CT</td>
<td>SD; vapor heat</td>
<td>0.2</td>
<td>0.2</td>
<td>EUR</td>
<td>Baxter</td>
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<tr>
<td>Innobrand</td>
<td>ion-exchange CT</td>
<td>SD</td>
<td>0.7</td>
<td>2.5</td>
<td>FR</td>
<td>LFB (Lille)</td>
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<tr>
<td>Koate DVI</td>
<td>precipitations, size exclusion</td>
<td>SD; dry heat</td>
<td>0.5</td>
<td>1.2</td>
<td>USA</td>
<td>Talecris</td>
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<td>8Y</td>
<td>heparin/glycine precipitations</td>
<td>dry heat</td>
<td>0.3</td>
<td>0.8</td>
<td>UK</td>
<td>BioProducts</td>
</tr>
</tbody>
</table>

Adapted from [37] and reproduced with permission from John Wiley, Copyright 2011. AU = Australia; BR = Brazil; CT = chromatography; EUR = Europe; FR = France; GE = Germany; IT = Italy; NF = nanofiltration; SD = solvent-detergent (t-N-butyl-PO₄ with polysorbate, Tween, or otoxynol, Triton); SP = Spain.

* Ratio of ristocetin cofactor activity (VWF:RCo) to VWF:Ag or FVIII activity expressed as IU/dl (or % of a normal pool).

* Humate-P in USA.
after infusion, with a corresponding decrease in bleeding
time from >30 to 10.5 min. Bleeding episodes in VWD3
patients were controlled with an average of 3 infusions
administered at a median interval of 16 h. Bleeding due
to surgical procedures was effectively controlled with a
median of 3 infusions at a median interval of 24 h. Al-
phanate tolerability was good, with 14% of patients expe-
riencing nonserious adverse events (AEs) possibly related
to study medication.

Results of a retrospective study in 22 patients with
VWD, 6 of whom had VWD3, showed that Fanhdi® (Gri-
fols, Barcelona, Spain) was effective in controlling bleed-
ing episodes and preventing excessive bleeding in surgi-
cal procedures [44]. No AEs were reported. In 2 open,
noncontrolled, multicenter studies in VWD patients un-
responsive to DDAVP, Fanhdi showed excellent or good
efficacy in controlling both spontaneous and surgery-re-
lated bleeding episodes [45]. Two nonserious AEs were
reported (phlebitis, considered possibly related and se-
roma, considered doubtfully related to study medica-
tion).

In one of the largest studies available on Fanhdi and
Alphanate, (Fanhdi and Alphanate Italian Retrospective
Study on Willebrand disease, FAIRest.Will), 120 patients
of all VWD types (10 VWD3, 19 VWD2A, 25 VWD2B,
10 VWD2M and 56 desmopressin-unresponsive VWD1)
were analyzed retrospectively. The patient population
had a median bleeding severity score of 8 (>5 indicates
moderate and severe cases). Clinical responses to the
products were good-to-excellent in 97% of bleeding epi-
sodes and in 99% of surgical procedures. A partial re-
response was observed in gastrointestinal bleeding, one
of the most complicated situations in VWD, possibly be-
cause other causes of bleeding were involved [46].

Haemate P®/Humate-P® (CSL Behring, Marburg,
Germany) is a preparation with a very high content of
high molecular weight VWF multimers (HMWM) and a
high ratio of VWF to FVIII content (VWF:FVIII; table 1).
The efficacy and tolerability of Haemate P/Humate-P
have been well documented in 3 retrospective and 3 pro-
spective studies. In a retrospective evaluation in 97 Ca-
nadian patients, 28 of whom had VWD3, the efficacy of
Haemate P/Humate-P was rated as excellent in 99% of
surgeries, in 97% of bleeding episodes and in all prophyl-
lactic treatments [47, 48]. Tolerability was good, with re-
lated AEs being nonserious and of mild or moderate inten-
sity (chills, phlebitis, vasodilatation, paresthesia, pru-
ritus, rash and urticaria). Another retrospective study
confirmed the efficacy and good tolerability of Haemate
P/Humate-P in 26 patients with VWD1 or VWD2A un-
dergoing surgical interventions and showed that the
mean daily doses and duration of treatment were higher
for major than for minor surgery (mean 39.3 VWF:RCo
IU/kg, mean 10.3 infusions vs. 28.7 VWF:RCo IU/kg,
mean 2.9 infusions, respectively) [49]. No related AEs or
thrombotic episodes were reported. The largest retro-
spective study enrolled 100 Italian patients, 37 of whom
had VWD3 [41]. In 95% of the patients treated for bleed-
ing events (n = 59) and in 97% of those treated for surgical
procedures (n = 56), responses were rated as excellent or
good. Tolerability was good, with no serious AEs or
thrombotic episodes. Two prospective, open-label, non-
randomized studies conducted in the USA assessed the
clinical efficacy and safety of this preparation in acute
bleeding events [50] and in urgent surgical situations [51].
The first study reported excellent or good efficacy in 98%
of the 53 bleeding events, 27 of which were in patients
with VWD3 (median loading dose 67.0 VWF:RCo IU/kg
and median daily maintenance dose 74.0 VWF:RCo IU/
kg) [50]. One related AE was reported (mild allergic reac-
tion) and 2 treatment episodes were associated with seri-
ous AEs unrelated to the study treatment. The other
study evaluated 39 patients with VWD, 8 of whom had
VWD3, and reported excellent or good efficacy in all
evaluated treatment events (median loading dose 82.3
VWF:RCo IU/kg and median daily maintenance dose
52.8 VWF:RCo IU/kg) [51]. Eight related AEs (2 events of
paresthesia, allergic reaction, vasodilatation, peripheral
edema, extremity pain, pseudo-thrombocytopenia and
pruritus) and 4 unrelated serious AEs were reported. An-
other prospective, open-label, multinational study in 35
patients undergoing surgery, 13 of whom had VWD3,
showed effective hemostasis in 91% of patients immedi-
ately after surgery [52]. Treatment-related AEs (headache,
itching, nausea and dizziness) were generally mild and of
short duration. Dosing of Haemate P/Humate-P for
bleeding prophylaxis in surgery was determined by mea-
suring the individual preoperative in vivo recovery of
VWF in a prospective open-label multicenter study [53].
Of the 29 patients in this study, 9 were diagnosed with
VWD3. The median recovery of VWF:RCo was 1.9 IU/dl
per IU/kg infused and no significant differences related
to VWD type were observed [53]. Based on these results,
a median dose of 62.4 VWF:RCo IU/kg was used for pro-
phylaxis in surgery, with efficacy rated as excellent or
good in 96.3% of patients on the day of surgery and in all
patients on the next day.

Biostate® (CSL Ltd, Melbourne, Australia) is a prepa-
ration with a high VWF:FVIII ratio (table 1). With re-
spect to FVIII:C and VWF markers, Biostate showed
pharmacokinetic behavior similar to that of AHF (high-purity, CSL), a VWF/FVIII concentrate previously licensed in Australia for the treatment of hemophilia A [54]. In a retrospective study of 43 VWD patients undergoing surgical procedures, 5 of whom had VWD3, Biostate showed an excellent (78%) or good (22%) hemostatic activity [55] in all of the 58 evaluated surgical procedures. No AEs related to Biostate administration were observed. In a prospective trial of 23 patients with VWD, 7 with VWD3, the hemostatic efficacy of Biostate was rated as excellent or good in all patients for all major and minor surgery events, for long-term prophylaxis, and for 4 of the 6 assessable nonsurgical bleeding events [56].

More recently, the results of a retrospective trial assessing the efficacy and safety of Biostate were reported for 43 children with VWD. In the VWD3 subgroup (n = 7), the hemostatic efficacy of Biostate was rated as excellent or good for 94% of all surgical and nonsurgical events. Biostate was well tolerated, with 1 case of nausea reported, possibly related to the study drug [57]. The current global clinical development program, SWIFT (Studies with von Willebrand factor/Factor VIII), is evaluating the pharmacokinetics, efficacy and safety of Biostate. It includes centers in Europe, Latin America and South America and consists of 6 clinical trials, 2 of which – SWIFT-VWD in adults (NCT00941616) and SWIFTLY-VWD in children (NCT01213446) – involve patients with VWD who may continue in the extension study – SWIFT-VWD-xt (NCT01224808) – to investigate the long-term safety and efficacy of Biostate.

Wilfactin® (LFB, Les Ulis, France) is a VWF concentrate depleted of FVIII. Two crossover, randomized, pharmacokinetic studies in 25 patients with VWD, 14 of whom had VWD3, showed that bleeding time was reduced in 71–86% of patients [58]. The only difference between Wilfactin and the VWF/FVIII concentrates was the 6-hour delay in FVIII increase with Wilfactin. Thus, for the treatment of acute bleeding, particularly in patients with VWD3 who had very low FVIII levels, supplementation with exogenous FVIII was recommended [58]. Wilfactin has been used successfully for many years in the treatment of VWD patients in hemophilia centers in France. Its efficacy and safety were investigated by pooling the results of 2 comparable prospective studies conducted in 5 European and 12 French centers. Of a total of 139 spontaneous bleeding episodes treated, 53 (38%) required a concomitant FVIII dose. Outcome was excellent or good in 89% of the episodes. Forty-four patients underwent 108 surgical or invasive procedures. Outcome was excellent or good in 95 of the scheduled procedures (only VWF was used) and 13 emergency procedures (a priming FVIII dose was coadministered with the first VWF infusion). There were no thrombotic complications and none of the VWD3 patients developed anti-VWF or anti-FVIII inhibitors [59].

Data on Wilate® (Octapharma, Lachen, Switzerland), a preparation with a low VWF:FVIII ratio (table 1), are available from a recent clinical review that included 4 European prospective, open-label, noncontrolled, nonrandomized, multicenter phase II or phase III clinical trials. Its efficacy and safety in spontaneous or posttraumatic bleeding was demonstrated in 44 patients with VWD (1,095 bleeding episodes), 24 of whom had VWD3 [60]. Tolerability was rated as very good or good in 99% of the 5,310 infusions, and no thrombotic events were reported; 13 related AEs in 5 patients were documented. The efficacy in the perioperative management of bleeding (57 surgical procedures in 32 patients, 19 of whom had VWD3) was rated as excellent or good in 96% of the interventions, with tolerability rated as very good or good in all interventions: according to the data of this study there was no accumulation of FVIII after repeated injections [61].

The recommended doses of VWF/FVIII concentrates depend on the product used and the clinical situation of the patient (table 2). The VWF:FVIII ratio is important to ensure that VWF is raised to an appropriate level, whilst maintaining a concentration of FVIII that will not induce thrombosis [62, 63]. Data comparing VWF/FVIII factor concentrates with a high (Haemate P) or low (Wilate) VWF:FVIII ratio in a rabbit model showed that the cumulative exposure to FVIII was 84% higher in animals treated with Wilate compared to those treated with Haemate P [64]. These results suggested that elevated FVIII levels could be avoided through the use of VWF/FVIII concentrates with higher VWF:FVIII ratios [64]. Factor concentrates vary also in their HMWM content (table 1). Because large VWF multimers are essential for platelet adhesion to sites of vascular injury and only the largest multimers are hemostatically active [4], a higher HMWM content is expected to be favorable for clinical efficacy.

Despite the differences in purity, VWF/FVIII content and virucidal techniques used, there is no evidence from retrospective or prospective clinical studies that the 6 VWF-containing concentrates (Alphanate, Biostate, Fanhdi, Haemate P, Wilate and Wilfactin), discussed above and shown in table 1, differ with regard to efficacy and safety, because, to date, no head-to-head clinical study has been carried out. Therefore, all of these VWF...
concentrates can be considered to be effective and well tolerated in the treatment or prevention of bleeding in VWD patients when the appropriate dosage (20–60 U/kg, see Table 2) of VWF:RCo is administered [37].

**Adjuvant Treatments**

Antifibrinolytic agents, such as tranexamic acid and e-aminocaproic acid, may be used in combination with plasma-derived VWF/FVIII concentrates to promote hemostasis and wound healing in dental extractions, tonsillectomy, menorrhagia and mucosal hemorrhage, but not in muscle hematomas or joint bleeding [1]. Platelet transfusions may be required in patients experiencing excessive and recurrent bleeding (e.g. gastrointestinal) despite treatment with VWF/FVIII concentrates and have been shown to be effective in some, if not all, patients [65–67]. Their presence not only renders VWF/FVIII replacement therapy ineffective, due to their interaction with the VWF domains critical for the therapy [69–73], but also introduces the risk of life-threatening anaphylactic reactions developing [70]. In fact, an increased level of circulating immune complexes is often observed after infusion of VWF/FVIII concentrates in patients with anti-VWF alloantibodies, which, in turn, may lead to classic complement pathway activation [70, 71, 73]. Patients who do not respond to VWF/FVIII replacement therapy with an increase in the plasma levels of both factors should be tested for the presence of anti-VWF alloantibodies. In such cases, recombinant factor VIIa (rFVIIa) and recombinant FVIII (rFVIII) products, which are devoid of VWF, may be used [70, 74–76]. As with VWF/FVIII concentrates, care should be taken to maintain the FVIII level below 150–200 IU/dl to reduce the risk of thrombosis [62].

**Complications/Difficulties Associated with Treatment of VWD3**

In rare cases (approximately 7.5–9.5% [68]), patients with VWD3 may develop anti-VWF alloantibodies. Most patients developing anti-VWF alloantibodies have a complete VWF gene deletion and thus a complete VWF deficiency [26, 27, 69]; however, not all of those with a complete deletion develop the antibodies [29]. All reported anti-VWF alloantibodies belong to the IgG class, yet their titer and specificity differs between patients [69, 70]. Their presence not only renders VWF/FVIII replacement therapy ineffective, due to their interaction with the VWF domains critical for the therapy [69–73], but also introduces the risk of life-threatening anaphylactic reactions developing [70]. In fact, an increased level of circulating immune complexes is often observed after infusion of VWF/FVIII concentrates in patients with anti-VWF alloantibodies, which, in turn, may lead to classic complement pathway activation [70, 71, 73]. Patients who do not respond to VWF/FVIII replacement therapy with an increase in the plasma levels of both factors should be tested for the presence of anti-VWF alloantibodies. In such cases, recombinant factor VIIa (rFVIIa) and recombinant FVIII (rFVIII) products, which are devoid of VWF, may be used [70, 74–76]. As with VWF/FVIII concentrates, care should be taken to maintain the FVIII level below 150–200 IU/dl to reduce the risk of thrombosis [62].

**Secondary Long-Term Prophylaxis**

Patients who experience frequent and severe bleeding episodes should be considered for regular prophylaxis with VWF/FVIII factor concentrates. For example, in patients with frequent bleeding in joints, long-term prophylaxis should be commenced before the development of joint disease, as is recommended for most bleeding disorders [77, 78]. The most common indications for long-term prophylaxis include joint bleeding, nose/mouth bleeding, menorrhagia and gastrointestinal bleeding [79, 80].

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**Table 2. Commonly recommended doses of VWF/FVIII concentrates in the treatment of VWD3**

<table>
<thead>
<tr>
<th>Indication</th>
<th>Dosea (VWF:RCo units)</th>
<th>Frequency of infusions (treatment duration)</th>
<th>Target VWF:RCo and FVIII levels</th>
</tr>
</thead>
<tbody>
<tr>
<td>Spontaneous or traumatic bleeding</td>
<td>25 IU/kg</td>
<td>daily infusions (2–4 days)</td>
<td>maintain trough VWF:RCo and FVIII level &gt;50% of normal level</td>
</tr>
<tr>
<td>Tooth extraction</td>
<td>20–40 IU/kg</td>
<td>single infusion</td>
<td>achieve trough VWF:RCo and FVIII level &gt;50% of normal level</td>
</tr>
<tr>
<td>Minor surgery</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Loading</td>
<td>30–60 IU/kg</td>
<td>every 12–48 h (1–5 days)</td>
<td>maintain trough VWF:RCo and FVIII level &gt;50% of normal level</td>
</tr>
<tr>
<td>Maintenance</td>
<td>20–40 IU/kg</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Major surgery</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Loading</td>
<td>40–60 IU/kg</td>
<td>every 8–24 h (7–14 days)</td>
<td>maintain trough VWF:RCo and FVIII level &gt;50% of normal level</td>
</tr>
<tr>
<td>Maintenance</td>
<td>20–40 IU/kg</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Data are from [63, 80].

*a Doses in pediatric patients are usually 20% higher than those in adults to correct for the greater plasma volume.*
The overall percentage of VWD patients treated with prophylaxis is low (reported as 1.6% in a study of 6,208 patients [77]), with the use of prophylaxis being more common in VWD3 than VWD1 or VWD2 (23.9% versus 0.2 or 1.8%, respectively, in a study of 5,343 patients [78]). The largest study of secondary long-term prophylaxis in VWD was conducted in 35 patients with severe VWD (28 of whom had VWD3) in Sweden [79]. The results showed that 63% of adult patients had been treated for more than 10 years with a resulting marked decrease in the number of joint bleeds. A recent study in 32 patients with VWD (13 of whom had VWD3) receiving prophylaxis for a median duration of 3 years reported a significant reduction in monthly bleeding frequency (from 3 to 0.07, \(p < 0.001\)) and the bleeding score (from 3 to 0, \(p < 0.001\)) over the 12-month observation period [79]. Preliminary results of an ongoing study in Germany in 24 patients with VWD3 showed reduction in bleeding events during periods of prophylaxis [81].

So far, the experience available does not allow the estimation and comparison of the costs for prophylaxis with the costs for treatment on demand [78]. The reported patient benefits of long-term prophylaxis include a decreased number of bleeding episodes, decreased severity of bleeds, prevention of arthropathy, an improved quality of life and allowing patients to lead a more normal life [82, 83]. In a survey in 34 federally funded Hemophilia Treatment Centers in the USA, 73% of respondents considered prophylaxis very/extremely helpful [20].

Not all patients may require long-term prophylaxis. The VWD Prophylaxis Network (VWD PN; http://www.vwd-prophy.com) has initiated a study to evaluate the benefits of long-term treatment. The aim is to determine the responsiveness to prophylaxis in patients with clinical manifestations of joint, gastrointestinal and/or nasal/oral bleeding, or menorrhagia, and the dose and intervals of treatment required to control bleeding episodes. In the prospective phase of the study, patients are treated with increasing doses of VWF/FVIII concentrate depending on the clinical events; patients will be observed for a follow-up period of 1 year. The VWD PN International Prophylaxis (VIP) nonrandomized dose-escalation study aims to identify people with VWD who may benefit from prophylaxis, to evaluate the effects of prophylaxis on bleeding frequency and to establish the optimal treatment regimens for joint bleeding, gastrointestinal bleeding, epistaxis and menorrhagia. Patients will receive prophylaxis with VWF/FVIII concentrate at doses increasing in 3 steps until bleeding is controlled, with initial doses being individually determined; patients will be followed up to 1 year. The VIP study also includes 2 retrospective studies to evaluate the efficacy of prophylaxis on the frequency of bleeding episodes in patients who are already receiving prophylaxis at the study centers, and to capture the natural history of gastrointestinal bleeding in patients with VWD.

The cost-effectiveness of prophylaxis regimens versus on-demand therapy is currently under investigation in another prospective study (PRO.WILL) organized in Italy, Spain and the UK [84].

Future Treatments

The currently available treatments with plasma-derived VWF/FVIII concentrates are suitable for all patients with VWD and mandatory for those with VWD3. Recombinant products are being developed and tested in clinical trials. Recombinant VWF has been characterized in vitro and in animal models, with promising results, and is currently undergoing clinical trials concerning treatment for VWD [85]. The interim results of a phase I pharmacokinetic study conducted in a cohort of 31 patients with VWD3 or VWD1 have recently been reported [86]. Recombinant interleukin 11 (IL-11) induces a gradual and sustained increase in VWF and FVIII levels by a mechanism different from that of desmopressin, as shown in a phase II, prospective, open-label, escalating-dose trial [87]. Therefore, IL-11 may be used in combination with desmopressin to overcome tachyphylaxis, or for the treatment of patients who are refractory to desmopressin. A recent phase II study in 7 women with mild VWD and refractory menorrhagia showed that recombinant IL-11 reduced the severity and duration of bleeding in 71 and 85% of patients, respectively [88]. Potential long-term or lifelong correction of VWF deficiency may be possible by delivery of the VWF protein by gene therapy [89]. As VWD results from a defect in a single gene, and because VWF is secreted in the circulation, which obviates the need for specific targeting, VWD is a good candidate for gene therapy [89]. However, with large proteins such as VWF, the technical difficulties are not negligible.

Conclusion

The treatment of VWD3 aims at correcting platelet adhesion and at restoring normal coagulation. The recommended first-line treatment is replacement therapy with plasma-derived VWF/FVIII concentrates, as pa-
patients with VWD3 do not respond to desmopressin. An important issue in VWD3 therapy is maintaining adequate levels of FVIII by using repeated infusions of exogenous VWF contained in VWF/FVIII concentrates. However, the circulating levels of FVIII should always be monitored after frequent infusions in order to avoid thrombotic events. Antifibrinolytic agents and platelet transfusions may be used concomitantly with VWF/FVIII concentrates as adjuvant therapy. Prophylaxis should be considered at least in some VWD3 patients, particularly those who experience frequent bleeding in joints or recurrent gastrointestinal bleeding. The patient benefits and cost-effectiveness of long-term prophylaxis need to be evaluated in prospective studies. Individual tailoring of the therapy to a patient’s clinical situation and response to treatment is currently the best choice.

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References


36 ISTH-SSC VWF Online Database: http://vwfgroup.ssh.edu/uk/ (accessed December 2011).


40 Cattaneo M, Simoni L, Gringeri A, Mannucci PM: Patients with severe von Willebrand disease are insensitive to the releasing effect of DDAVP: evidence that the DDAVP-induced increase in plasma factor VIII is not secondary to the increase in plasma von Willebrand factor. Br J Haematol 1994;86:333–337.


42 Federici AB, Mannucci PM, Steering Committee of the Type 3 von Willebrand International Registries Inhibitor Prospective Study (3WINTERS-IPS): http://www.vwd-3inters-ips.com (accessed December 2011).


