Serum Pentraxin-3 Levels Are Associated with the Severity of Metabolic Syndrome

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Objectives: The aim of the present study was to assess the association between the level of pentraxin-3 (PTX-3) and the severity of metabolic syndrome (MS).

Subjects and Method: One hundred and two patients with MS and 101 consecutive age- and sex-matched control subjects were included in the study. The MS patients were classified into three groups based on the number of MS criteria, i.e. group 1: patients with 3 MS criteria, group 2: patients with 4 MS criteria, and group 3: patients with 5 MS criteria. Serum PTX-3 and high-sensitivity C-reactive protein (hs-CRP) levels were measured.

Results: Group 1 had higher PTX-3 levels compared to the control group (0.58 ± 0.11 ng/ml vs. 0.36 ± 0.15 ng/ml, p < 0.001). PTX-3 levels were higher in group 3 than in both group 1 (0.90 ± 0.06 ng/ml vs. 0.58 ± 0.11 ng/ml, p < 0.001) and group 2 (0.90 ± 0.06 ng/ml vs. 0.63 ± 0.12 ng/ml, p < 0.001). Group 3, however, had higher hs-CRP levels than both group 1 (1.89 ± 0.45 mg/dl vs. 1.40 ± 0.44 mg/dl, p = 0.007) and group 2 (1.89 ± 0.45 mg/dl vs. 1.47 ± 0.58 mg/dl, p = 0.01). The control group had lower hs-CRP levels than group 1 (0.81 ± 0.47 mg/dl vs. 1.40 ± 0.44 mg/dl, p < 0.001) and group 2 (0.81 ± 0.47 mg/dl vs. 1.47 ± 0.58 mg/dl, p < 0.001).

Serum PTX-3 levels correlated with serum hs-CRP levels (r = 0.49, p < 0.001).

Conclusions: PTX-3, a novel inflammatory marker, was found to be associated with the severity of MS.

Key Words
Pentraxin-3 · Metabolic syndrome · High-sensitivity C-reactive protein · Inflammation

Abstract

Introduction

Metabolic syndrome (MS) is a cluster of risk factors including glucose intolerance, abnormal lipid profile, hypertension and abdominal obesity, each of which has been shown to be related to atherosclerosis and cardiovascular disease [1–5]. Most of the recent studies showed a correlation between components of MS and inflammatory mediators, namely, interleukin-6, tumor necrosis factor-α, and C-reactive protein (CRP) [6, 7]. Moreover, in patients with more risk factors comprising MS, the serum CRP levels were found to be increased and the higher serum CRP levels were associated with a higher prevalence of cardiovascular events which reflects the prognostic significance of the severity of MS [7].

Pentraxin-3 (PTX-3), a newly identified acute-phase reactant which resembles CRP both in structure and
function [8], has been found to be produced by many kinds of cells such as macrophages, dendritic cells, neutrophils, adipose cells, fibroblasts and vascular endothelial cells [9, 10]. Plasma PTX-3 levels have recently been found to be elevated in patients with vasculitis [11], acute myocardial infarction [12, 13], systemic inflammation or sepsis [14], psoriasis, unstable angina pectoris, and heart failure [15–18]. The association of MS and PTX-3 has not been extensively studied and the published data seem contradictory. Thus, the aim of the present study was to assess the association between the level of PTX-3 and the severity of MS.

**Subjects and Methods**

A total of 203 patients were included in this prospective randomized study. One hundred and two patients who were admitted to the outpatient clinic and met the MS criteria were taken as the MS group. One hundred and one age- and sex-matched subjects were the control group. Patients with secondary hypertension, renal failure, hepatic failure and/or manifest heart disease, such as cardiac failure, coronary arterial disease, arrhythmia, and cardiac valve disease, were excluded. Also, patients with infection, acute stress, chronic systemic inflammatory disease, and those receiving medications affecting the number of leukocytes were excluded as well. All participants included in the study were informed about the study, and their oral and written consent was obtained.

MS was diagnosed according to the National Cholesterol Education Program Adult Treatment Panel III criteria 3 (NCEP ATP 3) [3]. Diagnosis was established by the presence of 3 or more of the following criteria: abdominal obesity (waist circumference >102 cm in men and >88 cm in women); triglyceride level ≥150 mg/dl or ≥1.7 mmol/l; high-density lipoprotein (HDL) cholesterol level <40 mg/dl for men and <50 mg/dl for women (<1.0 mmol/l for men and <1.3 mmol/l for women); systolic blood pressure ≥130 mm Hg and/or diastolic blood pressure ≥85 mm Hg; fasting blood glucose concentration ≥110 mg/dl or ≥5.6 mmol/l. Patients were classified into three groups based on the number of MS criteria, i.e. group 1: patients with 3 MS criteria, group 2: patients with 4 MS criteria, and group 3: patients with 5 MS criteria.

Height, weight, and waist circumference were measured while fasting and standing up with standard measuring tools. The narrowest diameter between the costal arch and the anterior superior iliac spine was measured for waist circumference. Body mass index (BMI) and body area were calculated using the following formulas: BMI = weight (kg)/height (m)² and body surface area (m²) = 0.007184 × height (cm)⁰.⁷²⁵ × weight (kg)⁰.⁴²⁵. Blood pressure was measured after at least 10 min of rest in the sitting position. Blood pressure measurement was repeated two more times at 2-min intervals. The average of the 3 measurements was considered as blood pressure. Venous blood samples obtained in the morning after an 8-hour fast were used to measure serum glucose, urea, creatinine, total cholesterol, triglyceride, HDL, low-density lipoprotein, hemoglobin, high-sensitivity CRP (hs-CRP) and PTX-3 levels. Glomerular filtration rate was estimated by the Cockcroft-Gault formula. Serum PTX-3 level was measured by enzyme immunoassay using a quantitative kit (Human PTX-3/TSG-14 Immunoassay, DPTX30; R&D Systems, Inc., Minn., USA). hs-CRP was measured in serum by enzyme immunoassay (Immage hs-CRP EIA kit; Beckman Coulter Inc., USA). Complete blood count was performed in our hematology unit with the Beckman-Coulter Gen-S system device (Beckman-Coulter Inc., USA). Transthoracic echocardiography was performed and ejection fraction was measured by the biplane Simpson method.

**Statistical Analyses**

Statistical analyses were conducted with SPSS 17 (SPSS Inc., Chicago, Ill., USA) software package program. Continuous variables were expressed as means ± standard deviation, whereas categorical variables were presented as percentages. The differences between normally distributed numeric variables were evaluated using the t test or one-way ANOVA, while non-normally distributed variables were analyzed by the Mann-Whitney U test or Kruskal-Wallis variance analysis as appropriate. The χ² test was employed for the comparison of categorical variables. Univariate correlation and multivariate regression analyses were performed. A p < 0.05 was considered as statistically significant.

**Results**

Baseline clinical and laboratory characteristics of the patients according to groups are shown in table 1. Age, sex, BMI, smoking status and medications were not different amongst all groups. However, BMI and frequencies of hypertension and diabetes mellitus were higher in patients with MS.

Group 1 had higher PTX-3 levels compared to the control group (0.58 ± 0.11 ng/ml vs. 0.36 ± 0.15 ng/ml, p < 0.001). The PTX-3 levels were higher in group 3 than both group 1 (0.90 ± 0.06 ng/ml vs. 0.58 ± 0.11 ng/ml, p < 0.001) and group 2 (0.90 ± 0.06 ng/ml vs. 0.63 ± 0.12 ng/ml, p < 0.001). However, group 1 and group 2 had similar PTX-3 levels (0.58 ± 0.11 ng/ml vs. 0.63 ± 0.12 ng/ml, p = 0.44). On the other hand, there was no difference in levels of hs-CRP between group 1 and group 2 (1.40 ± 0.44 mg/dl vs. 1.40 ± 0.58 mg/dl, p = 0.97). Group 3, however, had higher hs-CRP levels than both group 1 (1.89 ± 0.45 mg/dl vs. 1.40 ± 0.44 mg/dl, p = 0.007) and group 2 (1.89 ± 0.45 mg/dl vs. 1.47 ± 0.58 mg/dl, p = 0.01). The control group had lower hs-CRP levels than group 1 (0.81 ± 0.47 mg/dl vs. 1.40 ± 0.44 mg/dl, p < 0.001) and group 2 (0.81 ± 0.47 mg/dl vs. 1.47 ± 0.58 mg/dl, p < 0.001) (fig. 1). Univariate correlation analysis revealed a positive correlation between serum PTX-3 levels and hs-CRP levels (r = 0.49, p < 0.001) (fig. 2). In the multivariate linear
Table 1. Baseline clinical and laboratory characteristics of patient groups

<table>
<thead>
<tr>
<th>Variables</th>
<th>Control group (n = 101)</th>
<th>Group 1 (n = 29)</th>
<th>Group 2 (n = 48)</th>
<th>Group 3 (n = 25)</th>
<th>p value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age, years</td>
<td>46.9 ± 13.8</td>
<td>49.8 ± 11.3</td>
<td>47.9 ± 9.1</td>
<td>47.6 ± 7.5</td>
<td>0.30</td>
</tr>
<tr>
<td>Male</td>
<td>53 (52.5)</td>
<td>16 (55.2)</td>
<td>24 (50)</td>
<td>13 (52)</td>
<td>0.85</td>
</tr>
<tr>
<td>BMI</td>
<td>24.7 ± 4.3</td>
<td>28.1 ± 6.1</td>
<td>29.2 ± 4.6</td>
<td>28.3 ± 2.2</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Waist circumference, cm</td>
<td>85 ± 11.5</td>
<td>96.7 ± 12.6</td>
<td>97.9 ± 10.9</td>
<td>105.6 ± 4.7</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>SBP, mmHg</td>
<td>130 ± 19.3</td>
<td>133.6 ± 22.2</td>
<td>132.5 ± 12.0</td>
<td>143.4 ± 18.8</td>
<td>0.005</td>
</tr>
<tr>
<td>DBP, mmHg</td>
<td>82.2 ± 12.7</td>
<td>82.8 ± 11.6</td>
<td>86.9 ± 6.1</td>
<td>89.8 ± 7.9</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Diabetes</td>
<td>8 (7.9)</td>
<td>10 (34.5)</td>
<td>17 (35.4)</td>
<td>8 (32)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Hypertension</td>
<td>39 (38.6)</td>
<td>16 (55.2)</td>
<td>27 (56.3)</td>
<td>16 (64)</td>
<td>0.045</td>
</tr>
<tr>
<td>Smoking</td>
<td>40 (39.6)</td>
<td>14 (48.3)</td>
<td>27 (56.3)</td>
<td>8 (32)</td>
<td>0.14</td>
</tr>
<tr>
<td>Ejection fraction</td>
<td>64 ± 4.3</td>
<td>60.3 ± 2.8</td>
<td>61.6 ± 2.1</td>
<td>62.3 ± 2.2</td>
<td>0.44</td>
</tr>
<tr>
<td>Aspirin</td>
<td>47 (46.5)</td>
<td>15 (51.7)</td>
<td>24 (50)</td>
<td>13 (52)</td>
<td>0.32</td>
</tr>
<tr>
<td>ACE-I or ARB</td>
<td>30 (29.7)</td>
<td>10 (29.4)</td>
<td>15 (31.2)</td>
<td>8 (32)</td>
<td>0.42</td>
</tr>
<tr>
<td>β-Blocker</td>
<td>10 (9.9)</td>
<td>3 (10.3)</td>
<td>5 (10.4)</td>
<td>3 (12.1)</td>
<td>0.24</td>
</tr>
<tr>
<td>CCB</td>
<td>3 (2.9)</td>
<td>1 (3.4)</td>
<td>2 (4.1)</td>
<td>1 (4)</td>
<td>0.40</td>
</tr>
<tr>
<td>Statin</td>
<td>10 (9.9)</td>
<td>3 (10.3)</td>
<td>5 (10.4)</td>
<td>2 (8)</td>
<td>0.35</td>
</tr>
<tr>
<td>Glucose, mg/dl</td>
<td>90.8 ± 13</td>
<td>101 ± 2.6</td>
<td>113.7 ± 23.4</td>
<td>121.6 ± 30.2</td>
<td>0.001</td>
</tr>
<tr>
<td>HDL-C, mg/dl</td>
<td>38.9 ± 7.6</td>
<td>37.6 ± 8.5</td>
<td>40.3 ± 11.5</td>
<td>37.3 ± 6.6</td>
<td>0.11</td>
</tr>
<tr>
<td>LDL-C, mg/dl</td>
<td>118.3 ± 31.2</td>
<td>113.9 ± 23.1</td>
<td>115.8 ± 18.2</td>
<td>121.7 ± 38.5</td>
<td>0.33</td>
</tr>
<tr>
<td>Triglyceride, mg/dl</td>
<td>130.0 ± 48.8</td>
<td>179.9 ± 74.2</td>
<td>169.3 ± 71.8</td>
<td>189.3 ± 79.9</td>
<td>0.04</td>
</tr>
<tr>
<td>BUN, mg/dl</td>
<td>12.3 ± 5.7</td>
<td>16.4 ± 9.2</td>
<td>15.9 ± 7.3</td>
<td>14.6 ± 7.3</td>
<td>0.12</td>
</tr>
<tr>
<td>Creatinine, mg/dl</td>
<td>0.78 ± 0.42</td>
<td>0.81 ± 0.30</td>
<td>0.82 ± 0.33</td>
<td>0.97 ± 0.24</td>
<td>0.21</td>
</tr>
<tr>
<td>eGFR, ml/min</td>
<td>104.3 ± 12.9</td>
<td>105.0 ± 14.1</td>
<td>100.6 ± 13.5</td>
<td>102.5 ± 14.8</td>
<td>0.15</td>
</tr>
<tr>
<td>hs-CRP, mg/dl</td>
<td>0.81 ± 0.46</td>
<td>1.41 ± 0.44</td>
<td>1.47 ± 0.58</td>
<td>1.88 ± 0.45</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Hb, g/dl</td>
<td>12.4 ± 1.1</td>
<td>13.2 ± 1.7</td>
<td>12.9 ± 2.0</td>
<td>13.2 ± 1.3</td>
<td>0.77</td>
</tr>
<tr>
<td>PTX-3, ng/ml</td>
<td>0.36 ± 0.15</td>
<td>0.58 ± 0.11</td>
<td>0.63 ± 0.12</td>
<td>0.90 ± 0.05</td>
<td>&lt;0.001</td>
</tr>
</tbody>
</table>

Systolic blood pressure; DBP = diastolic blood pressure; ACE-I = angiotensin-converting enzyme inhibitor; ARB = angiotensin receptor blocker; CCB = calcium channel blocker; eGFR = estimated glomerular filtration rate; HDL-C = high-density lipoprotein cholesterol; LDL-C = low-density lipoprotein cholesterol. Categorical variables were expressed as numbers with percentages in parentheses whereas numerical variables were expressed as means ± standard deviation. The comparisons between groups were done by the χ² or Kruskal Wallis test where appropriate. p values are overall trend values between groups and p < 0.05 was considered as significant. The comparison between the control group and group 1 was done by the Mann-Whitney U test and χ² test. The significant p values for these two groups were as follows: BMI, p = 0.005; waist circumference, p < 0.001; diabetes mellitus, p = 0.001; glucose, p < 0.001; triglyceride, p < 0.001; BUN, p = 0.045; hs-CRP, p = 0.001; PTX-3, p < 0.001.

Discussion

PTX-3, a novel inflammatory marker, was significantly higher in MS groups than the control group, and serum PTX-3 levels increased as the severity of MS increased. Many recent studies have shown that there is a correlation between components of MS and inflammatory mediators, namely, interleukin-6, tumor necrosis factor-α, and CRP [6]. Among these inflammatory biomarkers, the best characterized and standardized is the hs-CRP [19]. hs-CRP has been shown to be associated with insulin resistance [20], endothelial dysfunction [21] and adverse cardiovascular events [7, 22].

PTX-3 is produced by different kinds of cells such as adipocytes, macrophages, dendritic cells, neutrophils, fibroblasts, vascular endothelial cells [9, 10] and released by inflammatory stimuli and known cardiovascular risk factors, including oxidized low-density lipoprotein [23, 24], and is systematically correlated with age, BMI, waist circumference, glucose level, and HDL cholesterol.
24]; therefore, it may reflect the local inflammatory status in tissues [16]. Zanetti et al. [20] reported that the hs-CRP and the PTX-3 levels were higher in MS patients and they postulated that this elevation was related to inflammation and subclinical atherosclerosis. Another study reported that plasma PTX-3 levels were lower in individuals who had more than one MS component compared to healthy individuals. In the same study, however, hs-CRP levels were higher in individuals who had more than one MS component. They postulated that PTX-3 and CRP antagonistically participate in the development of MS. In the present study, we found that PTX-3 levels were higher in MS patients and as the number of MS components increased, the PTX-3 and the hs-CRP levels also increased.

**Fig. 1.** The mean PTX-3 and hs-CRP levels according to groups.

**Fig. 2.** Correlation plots for PTX-3 and hs-CRP.
This result seems to conflict with the aforementioned study. In their paper, they postulated that lower PTX-3 levels might be associated with the HDL cholesterol-induced PTX-3 inhibition. In fact, in our study, the included patients were different from those of the aforementioned study, which included only healthy males without any disease or medication. Firstly, most of our patients had risk factors such as hypertension, diabetes mellitus and therefore had an inflammatory background. In addition, the patients in our study had relatively lower HDL cholesterol levels. This may be the reason why PTX-3 levels were higher as the number of MS components increased. PTX-3 was well correlated with hs-CRP levels, which supports the baseline inflammatory background.

The reason why the patients with 5 MS components had higher PTX-3 and hs-CRP levels compared to patients with 3 or 4 components may be explained by the synergistic interactions of the 5 components resulting in an increased inflammatory status. Although elevated PTX-3 levels were found to be associated with adverse cardiovascular events in stable coronary artery disease [25] and acute coronary syndromes [13, 26], there is a lack of data about the prognostic significance of elevated PTX-3 in MS. Seemingly, for a more thorough understanding of the role of PTX-3 in MS, further studies are needed.

The major limitation of this study was the number (n = 203) of patients included. Another limitation was the lack of long-term follow-up of the patients, which could have provided prognostic information about the levels of PTX-3. Inflammatory markers other than hs-CRP such as interleukin-6 and tumor necrosis factor-α would support collecting more data about the inflammatory status.

Conclusion

PTX-3, a novel inflammatory marker, was associated with the severity of MS, which may reflect a higher inflammatory status associated with the increased number of components comprising MS.

References


Pentraxin-3 and Severity of Metabolic Syndrome

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