Mean Platelet Volume Is Increased in Infective Endocarditis and Decreases after Treatment

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\textbf{Key Words}
Mean platelet volume · Infective endocarditis · Thrombus · Vegetation

\textbf{Abstract}
\textbf{Objectives:} The aim of this study was to assess the mean platelet volume (MPV), an indicator of platelet activation in patients with infective endocarditis. \textbf{Subjects and Methods:} Twenty-nine patients with infective endocarditis and 29 healthy subjects were studied. Plasma MPV values in patients and control subjects were measured on admission and after 2 weeks of specific treatment of infective endocarditis. \textbf{Results:} The MPV was significantly higher among patients with infective endocarditis when compared with the control group (9.86 ± 1.1 vs. 8.0 ± 1.0 fl, respectively; \(p < 0.01\)). The MPV values of patients with infective endocarditis decreased significantly after treatment from 9.86 ± 1.1 to 7.86 ± 1.0 fl (\(p < 0.01\)). Total platelet counts increased significantly after treatment from 193.4 ± 96.5 \times 10^9 to 243.7 ± 92.4 \times 10^9 (\(p = 0.04\)). \textbf{Conclusion:} MPV values were higher in patients with infective endocarditis and decreased significantly after treatment. Elevated MPV values indicate that patients with infective endocarditis have increased platelet activation and infective endocarditis treatment decreases this platelet activation by decreasing MPV.

\textbf{Introduction}
Infective endocarditis remains a diagnostic and therapeutic challenge, as evidenced by the stability of its incidence over time and its morbidity and mortality rates [1, 2]. Endocardial vegetations can cause direct valvular damage. Embolic events are frequent and represent one of the life-threatening complications of infective endocarditis [3]. The endocardial thrombotic vegetation represents a specific model of pathogen/host tissue interaction, involving the formation of a septic thrombus leading to injury of both underlying valvar and cardiac tissue, and to possible peripheral septic dissemination [4]. Pathogen-platelet molecular interactions are probably one of the main determinants of vegetation formation [5] and growth that are linked to septic thrombus formation, including platelet activation and aggregation [6] and fibrin-fibronectin deposition [7, 8].

Previous studies have shown that platelet activation occurs in patients with infective endocarditis [9–11]. The mean platelet volume (MPV) is an indicator of platelet activation and is important in the pathophysiology of cardiovascular disease [12, 13]. MPV is an important and easily obtainable biological variable and larger platelets have higher thrombotic potential [14]. In comparison to smaller ones, larger platelets have more granules, aggre-
gate more rapidly with collagen, have higher thrombox-
ane A2 levels and express more glycoprotein Ib and IIb/ 
IIIa receptors [15–17]. Because of these important factors, 
we investigated MPV, an indicator of platelet activation 
in patients with infective endocarditis, and change in 
MPV values after treatment.

**Subjects and Methods**

Twenty-nine consecutive patients (19 M/10 F; mean age 56.2 ± 19.3 years, range 22–80) were studied between December 2005 and January 2010, with 29 control subjects (20 M/9 F; mean age 49.9 ± 12.1 years, range 32–67). All patients were hospitalized for infective endocarditis. Nine patients had prosthetic heart valves and 4 of them had pacemakers previously. All cases of left-sided infective endocarditis defined according to Duke criteria [18] with later modifications [19] for definite and possible infective endocarditis were included in the study. All patients initially received empiric antibiotic therapy containing ampicillin sul-bactame and gentamicin. The choice of antimicrobial agents was made on the basis of culture results or, in the case of culture-negative endocarditis, on an empirical basis. Treatment was given during a 2-week hospital stay and if clinical status was stable, patients were discharged from hospital and oral ampicillin sul-bactame were continued for at least 4 weeks according to ESC-AHA guidelines. Second blood samples for MPV levels were collected after 2 weeks of specific treatment of infective endocarditis. Exclusion criteria were left ventricular dysfunction, acute coro-
nary syndromes, lung disease and chronic renal or hepatic dis-
ees. No patients were using lipid lowering drugs. The protocol 
conformed to the ethical guidelines of our institution and in-
firmed consent was obtained from each participant.

**Blood Sampling**

Blood samples were drawn for blood culture from the antecu-
bital vein by puncture in a 21G sterile syringe without stasis. Pa-
tients’ MPV, white blood cell, hemoglobin, glucose, creatinine lev-
els and lipid profiles were also evaluated for analysis. The venous 
peripheral blood samples for MPV measurements were drawn on 
admission and 2 weeks after treatment. MPV was measured in a 
control blood sample collected in dipotassium EDTA tubes within 30 min 
after blood collection to prevent EDTA-induced swelling. An au-
tomatic blood counter (Beckman Coulter, Fullerton, Calif., USA) 
was used for whole blood counts. Glucose, creatinine and lipid 
profiles were also evaluated for analysis. The venous 
peripheral blood samples for MPV measurements were drawn on 
admission and 2 weeks after treatment. MPV was measured in a 
control blood sample collected in dipotassium EDTA tubes within 30 min 
after blood collection to prevent EDTA-induced swelling. An au-
tomatic blood counter (Beckman Coulter, Fullerton, Calif., USA) 
was used for whole blood counts. Glucose, creatinine and lipid profiles were measured with an auto analyzer (Olympus AU 640).

**Statistical Analysis**

Data were analyzed with the SPSS software version 15.0 for 
Windows (SPSS Inc., Chicago, Ill., USA). Data are expressed as 
mean ± standard deviation. The Kolmogorov-Smirnov test was 
used to test the distribution of numeric variables. To compare 
continuous variables, the Student t test or Mann-Whitney U test 
was used as appropriate. Because glucose levels can influence 
MPV values, glucose was taken as a covariate and covariate 
analysis (ANCOVA) was performed for MPV comparison. Categori-
cal variables were compared with the χ² test. To compare con-
tinuous variables before and after treatment, a paired sample t test 
was used. Statistical significance was defined as p < 0.05.

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Table 1. Comparison of the clinical and laboratory characteristics of the study and control groups

<table>
<thead>
<tr>
<th></th>
<th>Patients (n = 29)</th>
<th>Control (n = 29)</th>
<th>p value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age, years</td>
<td>56.2 ± 19.3</td>
<td>49.9 ± 12.1</td>
<td>0.11</td>
</tr>
<tr>
<td>Male</td>
<td>56.0 ± 20.0</td>
<td>48.9 ± 11.8</td>
<td>0.16</td>
</tr>
<tr>
<td>Female</td>
<td>56.7 ± 19.1</td>
<td>52.2 ± 13.7</td>
<td>0.53</td>
</tr>
<tr>
<td>Male/female</td>
<td>19/10</td>
<td>20/9</td>
<td>0.41</td>
</tr>
<tr>
<td>Hypertension</td>
<td>6 (20.7)</td>
<td>7 (24.1)</td>
<td>0.12</td>
</tr>
<tr>
<td>Medications</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>ACEi/ARB</td>
<td>5 (17.2)</td>
<td>4 (13.7)</td>
<td>0.71</td>
</tr>
<tr>
<td>CCB</td>
<td>1 (3.4)</td>
<td>3 (10.3)</td>
<td>0.30</td>
</tr>
<tr>
<td>Smoking</td>
<td>3 (10.3)</td>
<td>4 (13.8)</td>
<td>0.46</td>
</tr>
<tr>
<td>Obesity</td>
<td>4 (13.8)</td>
<td>5 (17.2)</td>
<td>0.06</td>
</tr>
<tr>
<td>Glucose, mg/dl</td>
<td>114.5 ± 31.8</td>
<td>88.4 ± 8.99</td>
<td>&lt;0.01</td>
</tr>
<tr>
<td>Creatinine, mg/dl</td>
<td>1.02 ± 0.21</td>
<td>0.89 ± 0.25</td>
<td>0.11</td>
</tr>
<tr>
<td>Total cholesterol, mg/dl</td>
<td>124.0 ± 25.0</td>
<td>182.8 ± 36.7</td>
<td>&lt;0.01</td>
</tr>
<tr>
<td>Triglycerides, mg/dl</td>
<td>135.5 ± 72.5</td>
<td>114.2 ± 63.1</td>
<td>0.32</td>
</tr>
<tr>
<td>LDL cholesterol, mg/dl</td>
<td>70.1 ± 24.1</td>
<td>109.8 ± 34.9</td>
<td>&lt;0.01</td>
</tr>
<tr>
<td>HDL cholesterol, mg/dl</td>
<td>28.3 ± 9.4</td>
<td>48.1 ± 12.0</td>
<td>&lt;0.01</td>
</tr>
<tr>
<td>Hemoglobin, g/dl</td>
<td>12.4 ± 1.3</td>
<td>13.9 ± 1.7</td>
<td>0.01</td>
</tr>
<tr>
<td>Platelet count, × 10⁹</td>
<td>193.4 ± 96.5</td>
<td>213.1 ± 39.5</td>
<td>0.32</td>
</tr>
<tr>
<td>MPV, fl</td>
<td>9.86 ± 1.1</td>
<td>8.0 ± 1.0</td>
<td>&lt;0.01</td>
</tr>
</tbody>
</table>

Figures in parentheses are percentages. p value is for comparison between control and study population. LDL = Low-density lipoprotein; HDL = high-density lipoprotein; ACEi = angioten-
sin-converting enzyme inhibitor; ARB = angiotensin receptor blocker; CCB = calcium channel blocker.

**Results**

The clinical and laboratory characteristics of the study 
population and the control subjects are presented in table 
1. There were no statistically significant differences 
between the two groups with respect to age, gender, history 
of hypertension, smoking status, obesity and levels of creatinine, triglyceride and platelet count. MPV was 
found to be normally distributed (p = 0.38). Fasting glu-
cose levels of 114.5 ± 31.8 versus 88.4 ± 8.99 mg/dl (p < 
0.01) and MPV values of 9.86 ± 1.1 versus 8.0 ± 1.0 fl 
(p < 0.01) were significantly higher in patients with infec-
tive endocarditis than in controls. The covariate analysis 
(ANCOVA) on effect of glucose levels on MPV did not 
show any effect on MPV (p = 0.21). Total cholesterol 124.0 
± 25.0 versus 182.8 ± 36.7 mg/dl (p < 0.01), low-density 
lipoprotein cholesterol 70.1 ± 24.1 versus 109.8 ± 34.9 
mg/dl (p < 0.01), high-density lipoprotein cholesterol 
28.3 ± 9.4 versus 48.1 ± 12.0 mg/dl (p < 0.01) and 
hemoglobin levels 12.4 ± 1.3 versus 13.9 ± 1.7 g/dl (p =

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MPV in Infective Endocarditis
Table 2. Comparison of the hematological parameters of the study population before and after treatment

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Before treatment</th>
<th>After treatment</th>
<th>p value</th>
</tr>
</thead>
<tbody>
<tr>
<td>WBC, $\times 10^3$</td>
<td>11.8 $\pm$ 5.8</td>
<td>10.1 $\pm$ 4.6</td>
<td>0.07</td>
</tr>
<tr>
<td>Hemoglobin, g/dl</td>
<td>10.6 $\pm$ 1.8</td>
<td>10.4 $\pm$ 1.6</td>
<td>0.42</td>
</tr>
<tr>
<td>Platelet count, $\times 10^9$</td>
<td>193.4 $\pm$ 96.5</td>
<td>243.7 $\pm$ 92.4</td>
<td>0.04</td>
</tr>
<tr>
<td>MPV, fl</td>
<td>9.86 $\pm$ 1.1</td>
<td>7.86 $\pm$ 1.0</td>
<td>&lt;0.01</td>
</tr>
</tbody>
</table>

*p value is for comparison between parameters before and after treatment. WBC = White blood cells.*

Discussion

The MPV was significantly higher in patients with infective endocarditis when compared with control subjects thereby implicating platelet activation in patients with infective endocarditis as previously reported [10]. Our finding that MPV values decreased significantly after treatment also confirmed the previous finding of Gunebakmaz et al. [20].

MPV has recently become an active area in cardiovascular research. It is a simple and easy laboratory measurement of assessing platelet function [12, 21] because it can be measured in almost all laboratories. Platelets are heterogeneous in size, density and reactivity. In comparison to smaller ones, larger platelets have more granules, aggregate more rapidly with collagen, have higher thromboxane A2 level and express more glycoprotein Ib and IIb/IIIa receptors [14, 16, 17].

The reason for increased MPV in infective endocarditis is not exactly known. There are a number of probable causes. Firstly bacteria-platelet interactions might play a critical role in platelet activation and thrombus formation in infective endocarditis [4–6]. Direct or indirect binding of bacteria to platelets can provoke their activation and aggregation, resulting in fibrin formation [5, 22]. Secondly, it has been shown that serum levels of IL6, IL-1β and C-reactive protein were significantly elevated in patients with infective endocarditis as compared to controls [23]. Elevated inflammatory cytokines such as IL-3 and IL-6 can lead to the production of more reactive and larger platelets as reported previously [24]. Hence, elevated levels of IL-6 in infective endocarditis might be a cause of increased MPV. Thirdly, endothelial injury due to bacterial toxins, hemodynamic forces and autoimmune reactions might lead to platelet activation and consequently increased MPV in infective endocarditis. It has been reported that once endothelial injury occurs, it leads to the denudation of the vessel wall, hence the local antithrombotic function of the endothelium cannot be maintained and local thrombosis with the formation of vegetations occur immediately [25].

Thrombocytopenia in critically ill patients on admission has been recognized as a poor prognostic sign [26, 27] and is associated with a higher relative risk of mortality in septic patients [26]. We found lower platelet count in patients with infective endocarditis than the control group and platelet count increased significantly after treatment.

The limitations of this study include the small number of patients and that the analysis was based on a simple baseline determination that may not reflect the patient status over long periods. Also, even though age difference was not statistically different, the range of age was narrower in the control subjects and the control group was not also followed up and compared.

Conclusion

In this study, MPV values were significantly higher among patients with infective endocarditis when compared with the control group and the values decreased significantly after treatment. Elevated MPV indicated that patients with infective endocarditis have an increased platelet activation and infective endocarditis treatment decreased platelet activation. We recommend further prospective studies in order to establish the pathophysiological and clinical significance of increased MPV in patients with infective endocarditis.
References