Stem Cell Treatment for Chronic Lung Diseases

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Asthma · Chronic lung diseases · Chronic obstructive pulmonary disease · Cystic fibrosis · Idiopathic pulmonary fibrosis · Mesenchymal stem cells

Abstract
Chronic lung diseases such as idiopathic pulmonary fibrosis and cystic fibrosis or chronic obstructive pulmonary disease and asthma are leading causes of morbidity and mortality worldwide with a considerable human, societal and financial burden. In view of the current disappointing status of available pharmaceutical agents, there is an urgent need for alternative more effective therapeutic approaches that will not only help to relieve patient symptoms but will also affect the natural course of the respective disease. Regenerative medicine represents a promising option with several fruitful therapeutic applications in patients suffering from chronic lung diseases. Nevertheless, despite relative enthusiasm arising from experimental data, application of stem cell therapy in the clinical setting has been severely hampered by several safety concerns arising from the major lack of knowledge on the fate of exogenously administered stem cells within chronically injured lung as well as the mechanisms regulating the activation of resident progenitor cells. On the other hand, salient data arising from few ‘brave’ pilot investigations of the safety of stem cell treatment in chronic lung diseases seem promising. The main scope of this review article is to summarize the current state of knowledge regarding the application status of stem cell treatment in chronic lung diseases, address important safety and efficacy issues and present future challenges and perspectives. In this review, we argue in favor of large multicenter clinical trials setting realistic goals to assess treatment efficacy. We propose the use of biomarkers that reflect clinically inconspicuous alterations of the disease molecular phenotype before rigid conclusions can be safely drawn.

Introduction

Stem cells are considered to be cells that are capable of unlimited self-renewal and differentiation into several cellular subtypes depending on their origin and the resident microenvironment. In humans, stem cells can be

subdivided into two main categories: embryonic stem cells (ESCs) and adult stem cells [1]. The latter are found in a number of tissues, including the bone marrow (BM), blood, adipose tissue, liver, kidney, heart and the lungs; they contribute to tissue repair and provide a continual source of cells throughout an individual’s life span [1–3]. In the lung, current evidence suggests that these cells may participate in tissue homeostasis and regeneration after injury [2–7] and may originate from within the lung itself, nesting in protected niches in the distal airways [8], called resident progenitor cells (alveolar, endothelial and interstitial), or from distant sites such as the blood, BM and adipose tissue, namely endothelial progenitor cells (EPCs) [5] and mesenchymal stem cells (MSCs) [3, 9]. While it is possible that such cells will ultimately be useful as alternatives to ESCs to prevent or treat lung disease, the therapeutic potential of adult stem cells in patients with lung diseases has to be better defined.

Chronic lung diseases such as idiopathic pulmonary fibrosis (IPF) and cystic fibrosis (CF) or chronic obstructive pulmonary disease (COPD) and asthma are leading causes of morbidity and mortality worldwide with a considerable human, societal and financial burden; chronic lung diseases rank second after cardiovascular diseases and amount to a gross estimate of more than 11 million deaths in the United States in 2020 [6, 10].

During the past 5 years, since the first clinical trial of the feasibility, safety and ability of EPCs to treat idiopathic pulmonary hypertension [11, 12] was conducted, studies of stem cells and cell therapies in lung biology and diseases have continued to expand rapidly [13]. The latter scientific explosion reflects the amenable need of chest physicians to respond to the will of patients suffering from end-stage chronic lung diseases that will culminate into a fatal outcome irrespective of currently available treatments. Starting from seminal observations demonstrating the potency of stem cells, notably adult mesenchymal stromal cells, to differentiate into alveolar epithelial cell (AEC) and endothelial cell lineages and rapidly extending to experimental and human lung explant models providing evidence of their efficacy, we witnessed significant advances regarding the mechanisms of action, biological properties and regenerative capacity of adult stem cells [13].

Nevertheless, despite relative enthusiasm arising from promising therapeutic applications of stem cells in experimental models of chronic lung diseases, there has been a paucity of preclinical and clinical studies regarding frequent chronic destructive lung diseases, like IPF, CF and COPD. It is generally recognized that this significant lack could be attributed to unresolved ethical and safety concerns mainly arising from the yet unknown disease immunopathogenesis and the need for a better understanding of the mechanisms underlying pleiotropic properties of adult stem cells. With a gradually increasing worldwide incidence and no proven therapies other than lung transplantsations, the role of stem cells in the treatment of this group of diseases is of significant interest.

The main scope of this review article is to summarize the current state of knowledge, based on both human and experimental studies, regarding the application status of stem cell treatment in chronic lung diseases, including IPF, COPD, CF and asthma, to address some important safety and efficacy issues that pose significant limitations to their implementation in current clinical practice and to present future challenges and perspectives. More importantly, there is an urgent need for large multicenter clinical trials with a painstaking study design by setting realistic goals to assess efficacy using biomarkers that reflect clinically inconspicuous alterations of the disease molecular phenotype before rigid conclusions can be safely drawn.

**Stem Cells**

**Mesenchymal Stem Cells**

Among the stem cell population, MSCs are the most extensively studied and probably have the best results in medical research. Since their seminal identification, almost 40 years ago, as a nonhematopoietic stem cell of mesodermal origin with a fibroblast-like morphology and potency to differentiate into both mesenchymal and nonmesenchymal cell lineages, the body of evidence relating to their differentiation, immunophenotype, preclinical use as well as their mechanisms of action has increased dramatically [3, 7, 9, 13, 14]. MSCs, besides BM, can also be readily harvested from other tissues, including adipose tissue, skeletal muscle, dental pulp and cord blood [15–18]. Of special interest is adipose tissue since it represents an abundant and easily accessible source of MSCs denominated adipose-derived stem cells (ADSCs), also including stromal vascular fraction cells. MSCs possess outstanding pleiotropic properties, including differentiation and regenerative and migratory capacity. The latter characteristic allows them to home selectively to sites of tissue injury and exert their immunosuppressive activity with the secretion of angiogenic, anti-apoptotic and anti-inflammatory factors [15, 18–21], such as stromal-derived factor-1, monocyte chemoattractant protein-3, vascular endothelial growth factor (VEGF), hepatocyte growth
factor, stimulating angiogenesis, fostering a protected environment for host cell recovery and preserving or even rescuing injured tissue from damage [19–22].

The above impressive characteristics render MSCs major candidates for therapeutic applications in patients with chronic lung diseases and provide a strong rationale to explore their potentially beneficial use.

Endothelial Progenitor Cells

EPCs represent microvascular endothelial cells originating from circulating BM-derived vascular progenitor cells and expressing specific surface antigens. They are divided into two cell subsets, namely early and late outgrowth EPCs, respectively, based on their morphology, timing of appearance in colony assays and immunophenotypic and functional profile. In particular, early outgrowth EPCs express the leukocyte markers CD45, CD11 and CD14, the endothelial markers CD31 and VEGF-A and the hematopoietic marker CD133. They are now known to be derivatives of the hematopoietic lineage differentiating among the myeloid lineage in response to variable soluble mediators. The other type, called late outgrowth, is characterized by CD31, CD144, CD146, CD105 and VEGF-R2 expression and possesses the unique ability to participate in angiogenesis, i.e. the formation of new blood vessels [23–25]. Therefore, they have been directly implicated in the restoration of endothelial function and vascular structure. Nevertheless, studies have also revealed a dual role for EPCs because they may also be involved in vascular remodeling by differentiating into smooth muscle cells leading to intimal hyperplasia and vascular muscularization and vasoconstriction with increased vascular resistance [23–29].

The exact mechanisms orchestrating the differentiation of circulating EPCs towards either mature endothelial cells or smooth muscle cells are still a matter of debate, and several studies propose that the source of EPCs (BM, peripheral blood or umbilical cord) and the microenvironment of engraftment fostering neighboring cells, growth factors and cytokines are presumably the key regulators of the final decision on the fate of EPCs [23–29].

Given their opposing pleiotropic activities, circulating levels of EPCs have been the subject of extensive investigations in both health and disease. In specific, in healthy subjects, the number of circulating EPCs decreases with age and cigarette smoke exposure while the presence of cardiovascular risk factors results in similar effects. With respect to chronic lung diseases (most notably COPD), the evidence discussed below seems rather conflicting and controversial [5, 23, 30–33].

Alveolar Epithelial Progenitor Cells

The airway epithelium is a dynamic tissue that undergoes constant and rapid renewal in order to vigorously respond to repeated exogenous and endogenous threats and reestablish an epithelial sheet with normal structure and function, mainly based on niches of resident progenitor cells. These are multipotent stem cells programmed to move down a certain pathway of differentiation and are often called progenitors to avoid implying that they are totipotent, as it happens with ESCs. Although several lines of research suggest that basal and secretory/or Clara cells are multipotent and can reconstitute a full epithelium with the contribution of the unipotent type-II AECs, stem cell niches and their microenvironments within the human lung have not been extensively characterized [4, 8].

Most recently, it has been intriguingly demonstrated that the human lung contains identifiable stem cells that give rise to completely structured respiratory units comprising bronchioles, alveoli and pulmonary vessels when injected within damaged mouse lungs. Nevertheless, it is still debatable whether these respiratory units are fully functional and participate in gas exchange. In addition, there is no definitive knowledge whether rare airway epithelial progenitor/stem cells proliferate, migrate and differentiate in a highly orchestrated procedure similar to classical models of high turnover in tissues such as the liver or skin or alternative models of simple duplication of differentiated cells (e.g. in the pancreas) are better applicable in lung tissue. Furthermore, there is significant lack of knowledge on the endogenous signals that control type-II AEC proliferation and differentiation into type-I AEC [4].

On the basis of the above predicament, a major challenge arising from developmental biology is to induce lung tissue regeneration by administering endogenous signaling molecules that are essential for lung development and maintenance [34].

ESCs and Induced Pluripotent Stem Cells

ESCs are derived from the inner cell mass of the blastocyst and are considered totipotent in their ability to regenerate all three germ layers of an organism, whereas stem cells derived from the adult are considered multi- or unipotent, and able to give rise to one or several mature cell types. Induced pluripotent stem cells (iPSCs) are mature cells derived from several tissues, including the skin, liver, kidney and lung, that have been reprogrammed back to an embryonic-like state. They are an ethical alternative to cloning or destroying in vitro fertilization embryos for pluripotent stem cells [13].
During the past 5 years, although progress using ESCs for lung regeneration or repair has been accomplished, e.g. by the generation of cells with phenotypic characteristics of type-II AECs, derivation of fully functional airway epithelium from ESCs has proven even more elusive and controversial. To our disappointment, several ethical concerns have hampered the efforts of investigators, and thus the number of available studies on the effects of ESC administration to the lung in vivo is scarce, e.g. regarding survival of mouse ESC-derived type-II AEC and maintenance of pro-surfactant protein C expression for 1 day [35].

Regarding iPSCs, rapid advances in isolation and generation technologies have raised hope that these cells could serve as reliable alternatives for tissue renewal and restoration [36–38]. As described below, these advances generated disease-specific human iPSCs from patients with both genetic and acquired chronic lung diseases, including CF, α1-antitrypsin deficiency and scleroderma [36–38].

**Stem Cell Therapy in IPF**

IPF is an irreversible, devastating, fibroproliferative disorder of the lung that culminates into a fatal outcome irrespective of treatment [39–41]. Despite extensive research and rapid expansion of scientific knowledge, IPF pathogenesis still remains with numerous question marks. Recent data strongly suggest that the mechanisms driving IPF reflect abnormal wound healing in response to multiple sites of ongoing alveolar epithelial injury of unknown origin leading to fibroblast activation and exaggerated accumulation of extracellular matrix in the lung parenchyma [42–45]. Therefore, our present understanding of the molecular and cellular pathways has resulted in the testing of therapeutic approaches that modulate specific inflammatory and fibrotic mediators, however with minimal results [40, 46, 47]. With a gradually increasing worldwide incidence and no proven therapies other than lung transplantations, IPF treatment represents a major challenge and bottleneck for chest physicians. Therefore, the role of stem cells in the treatment of this disease is greatly warranted [7].

A continuing accumulation of data in animal models suggests that cell-based therapies may be potential therapeutic approaches for lung regeneration and normal wound healing after injury (table 1). An attempt to address this crucial issue was made by Ortiz et al. [48] who reported diminished histological and inflammatory injury in the bleomycin (BLM) model of pulmonary fibrosis following intravenous instillation of MSCs. Fueled by the same prospect, Rojas et al. [49] intravenously administered BM-derived MSCs to mice following BLM-induced lung injury and observed decreased expression of inflammatory cytokines; their findings were verified by two follow-up studies conducted by Germano et al. [50] and Zhao et al. [51].

In line with this evidence, a beneficial effect of intratracheal and systemic infusion of MSCs in the BLM model of lung injury, which was assessed by decreases in lung collagen accumulation, fibrosis score and matrix metalloproteinase levels, has also been reported by Moodley et al. [52]. Further extending the latter finding, the same group of investigators evaluated the role of human umbilical cord MSCs in the treatment of BLM-induced lung inflammation and fibrosis. MSCs were administered systemically 24 h after BLM instillation to immunodeficient mice and were notably visualized at areas of fibroblastic foci 14 days after BLM injection but they were absent thereafter (by 28 days). Although differentiation of MSCs into AECs failed in vitro, their administration resulted in a significant attenuation of inflammation and fibrosis, which was reflected by the reduction in collagen deposition as well as inflammatory and profibrotic mediators, including TNFα, TGFβ and IL-10. Furthermore, BM-derived MSCs modified to express keratinocyte growth factor via an inducible lentivirus have been found to protect from BLM-induced lung inflammation and fibrosis, which was assessed by a significant reduction in lung collagen deposition and inflammatory cytokine production [53].

Similar protective potentially paracrine effects of systemically administered BM-derived MSCs in the BLM model of lung fibrosis have also been reported by Kumamoto et al. [54] as well as Lee et al. [55], who demonstrated improvement in the lung injury score and modulation of inflammatory cytokine production, respectively. Despite these interesting and very promising data, there are significant limitations mainly arising from the lack of a specific mechanism of action through which MSCs exert their beneficial role in the experimental model of lung fibrosis.

Results were further reproduced by Cargnoni et al. [56], who reported improved body weight and decreased histological fibrosis in BLM-injured mouse lungs following administration of allogeneic MSCs via three different routes.

Finally, there are only two published reports showing beneficial effects of ESC administration in experimental
models of BLM- and silica-induced lung injury. Authors performed an impressive series of experiments and starkly demonstrated that intratracheal instillation of type-II AECs derived from human ESCs resulted in an intriguingly high percentage of prolonged engraftment in injured lungs while an attenuation of BLM- or silica-induced inflammation and fibrosis (assessed by both structural and functional measurements) was also notable [35, 57].

The above data indicate that it is conceivable to speculate that the beneficial effect of MSCs reported in the experimental model of lung fibrosis is mainly attributable to their immunomodulatory paracrine activity rather than their capacity to differentiate or alternatively their potency to generate AECs. In line with this premise, a considerable number of studies have failed to demonstrate true phenotypic differentiation of adult MSCs of different origins into AECs. There was only one study in the literature so far (Sueblivong et al. [58]) which demonstrated that a small proportion of less than 5% of umbilical-cord-blood-derived MSCs cultured in special airway growth media preferentially expressed epithelial cell markers. Nevertheless, the above observation is weakened by the finding that only a minority of these cells engrafted into airway epithelium, indicating that most of the cells only temporarily lodged in the capillary beds of the pulmonary vasculature and were then either cleared or migrated to other sites. With regard to ESCs, it re-

### Table 1. Experimental data of stem cell therapy in animal models of pulmonary fibrosis

<table>
<thead>
<tr>
<th>Study year</th>
<th>Type of stem cells</th>
<th>Animal model</th>
<th>Route of administration</th>
<th>Outcome</th>
<th>Potential mechanisms of action</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ortiz et al. [48] 2003</td>
<td>Mouse BM-MSCs Plastic adherent</td>
<td>BLM</td>
<td>Intravenous</td>
<td>Decreased histologic and inflammatory injury, hydroxyproline, matrix metalloproteinase-2 and -9</td>
<td>None specific</td>
</tr>
<tr>
<td>Rojas et al. [49] 2005</td>
<td>Mouse BM-MSCs</td>
<td>BLM</td>
<td>Intravenous</td>
<td>Decreased expression of inflammatory cytokines</td>
<td>None specific</td>
</tr>
<tr>
<td>Germano et al. [50] 2009</td>
<td>Mouse BM-MSCs Plastic adherent</td>
<td>BLM</td>
<td>Intravenous</td>
<td>Decreased systemic inflammatory cytokines (IL-1β, IFN-γ, IL-6, IL-8, MIP-1α)</td>
<td>None specific</td>
</tr>
<tr>
<td>Zhao et al. [51] 2008</td>
<td>Rat BM-MSCs Plastic adherent</td>
<td>BLM</td>
<td>Intravenous</td>
<td>Decrease in histologic injury, hydroxyproline, laminin, hyaluronan, TGFβ, PDGF, IGF</td>
<td>None specific</td>
</tr>
<tr>
<td>Moodley et al. [52] 2009</td>
<td>Human umbilical cord MSCs</td>
<td>BLM</td>
<td>Intravenous</td>
<td>Reduction in histologic injury, collagen deposition, hydroxyproline, TIMP-2</td>
<td>None specific</td>
</tr>
<tr>
<td>Aguilar et al. [53] 2009</td>
<td>Mouse BM-MSCs</td>
<td>BLM</td>
<td>Intravenous</td>
<td>Decrease in collagen deposition, αSMA, TNFα, CCL-2, CCL-9</td>
<td>Keratinocyte growth factor secretion</td>
</tr>
<tr>
<td>Kumamoto et al. [54] 2009</td>
<td>Mouse BM-MSCs Plastic adherent</td>
<td>BLM</td>
<td>Intravenous</td>
<td>Decrease in histologic injury, hydroxyproline, inflammatory cells</td>
<td>None specific</td>
</tr>
<tr>
<td>Lee et al. [55] 2010</td>
<td>Rat BM-MSCs Plastic adherent</td>
<td>BLM</td>
<td>Intravenous</td>
<td>Decreased inflammation (neutrophils, BALF inflammatory cytokines), collagen deposition</td>
<td>None specific</td>
</tr>
<tr>
<td>Cargnoni et al. [56] 2009</td>
<td>Placenta MSCs</td>
<td>BLM</td>
<td>Intravenous</td>
<td>Improved body weight</td>
<td>None specific</td>
</tr>
<tr>
<td>Wang et al. [35] 2010</td>
<td>ESCs</td>
<td>BLM</td>
<td>Intratracheal</td>
<td>Abrogation of lung injury/fibrosis, improvement in tidal volume, gas exchange</td>
<td>Differentiation of human ESCs to type-II AECs</td>
</tr>
<tr>
<td>Spitalieri et al. [57] 2012</td>
<td>Human ESCs</td>
<td>Silica</td>
<td>Intratracheal</td>
<td>Diminished lung injury, decreased levels of TGFβ, improved gas exchange</td>
<td>Differentiation of human ESCs to type-II AECs</td>
</tr>
</tbody>
</table>

αSMA = α-Smooth muscle actin; BALF = bronchoalveolar lavage fluid; IFN = interferon; MIP = macrophage inflammatory protein; PDGF = platelet-derived growth factor; TGF = transforming growth factor; TNF = tumor necrosis factor; TIMP = tissue inhibitor of metalloproteinases.
mains to be elucidated whether their beneficial effect reflected structural engraftment and increased capacity to differentiate or was simply attributable to paracrine properties.

**Stem Cell Therapy in COPD**

COPD represents a disease paradigm where gas exchange abnormalities resulting in respiratory failure reflect pathogenetic dramatic changes in lung architecture, including loss of lung elasticity, vascular remodeling and luminal obstruction with inflammatory mucoid secretions, underlying the physiologic hallmarks of the disease [25, 59, 60]. Evidence accumulates that these key pathologic alterations are associated with an exaggerated inflammatory process in response to repeated inciting stimuli, including cigarette smoke exposure, and viral or microbial infections coupled with genetically predisposed accelerated mesenchymal cell senescence and potentially impaired mobilization of resident progenitor cells. This lethal combination of genetic susceptibility and epigenetic stimuli ultimately leads to endothelial dysfunction, destruction or stiffening of pulmonary capillaries with progressive accumulation of connective tissue and AEC apoptosis resulting in pulmonary hypertension and emphysema. On the basis of the above-mentioned different aspects of disease pathogenesis and pleiotropic properties of stem cells, including immunomodulatory, anti-inflammatory and anti-apoptotic factors, a steadily increasing number of studies has evaluated the efficacy of systemic or intratracheal stem cells, notably MSC administration in a variable spectrum of lung injury models in mice [25, 34, 59–62] (table 2).

**MSCs and Immunomodulation**

The first attempt to address this crucial issue was made by Shigemura et al. [63, 64] who demonstrated that ADSCs ameliorated pulmonary emphysema in an experimental model by secreting large amounts of hepatocyte growth factor. Similar protective paracrine effects of BM-derived MSCs have also been demonstrated by several investigators in experimental models of pulmonary emphysema induced by papain [65, 66] or intratracheally administered elastase [67]. Based on the above observations, Schweitzer et al. [68] suggested that the protective effect of intravenously given human or mouse adult ADSCs in a model of inflammation and injury induced by cigarette smoke exposure was mediated via a paracrine pathway involving growth factors and angiogenic modulators. In addition, beneficial effects of ADSCs were also observed on a systemic basis, i.e. by the prevention of weight loss and restoration of BM dysfunction.

**Table 2. Experimental data of stem cell therapy in animal models of COPD**

<table>
<thead>
<tr>
<th>Study year</th>
<th>Type of stem cells</th>
<th>Animal model</th>
<th>Route of administration</th>
<th>Outcome</th>
<th>Potential mechanisms of action</th>
</tr>
</thead>
<tbody>
<tr>
<td>Shigemura et al. [63] 2006</td>
<td>Rat ADSCs Plastic adherent</td>
<td>Rat elastase</td>
<td>Direct topical application</td>
<td>Improved histologic repair</td>
<td>Hepatocyte growth factor secretion</td>
</tr>
<tr>
<td>Shigemura et al. [64] 2006</td>
<td>Rat ADSCs Plastic adherent</td>
<td>Rat elastase</td>
<td>Intravenous</td>
<td>Decreased apoptosis, improved histologic repair, improved gas exchange and exercise tolerance</td>
<td>Hepatocyte growth factor secretion</td>
</tr>
<tr>
<td>Zhen et al. [65] 2008</td>
<td>Rat BM-MSCs</td>
<td>Rat papain</td>
<td>Intravenous</td>
<td>Decreased histologic injury and AEC apoptosis</td>
<td>None specific</td>
</tr>
<tr>
<td>Zhen et al. [66] 2010</td>
<td>Rat BM-MSCs</td>
<td>Rat papain</td>
<td>Intratracheal</td>
<td>Decreased histologic injury</td>
<td>None specific</td>
</tr>
<tr>
<td>Katsha al. [67] 2011</td>
<td>Mouse BM-MSCs Plastic adherent</td>
<td>Mouse elastase</td>
<td>Intratracheal</td>
<td>Reduction in collagen deposition and inflammatory and profibrotic cytokines (TNFa, TGFβ, IL-10)</td>
<td>None specific</td>
</tr>
<tr>
<td>Schweitzer et al. [68] 2011</td>
<td>ADSCs (mouse and human)</td>
<td>Cigarette smoke</td>
<td>Intravenous</td>
<td>Decrease in collagen accumulation, fibrosis score, metalloproteinase levels, weight loss, BM suppression</td>
<td>None specific</td>
</tr>
</tbody>
</table>
EPCs and Endothelium Repair

As mentioned above, the exact pathogenetic mechanisms underlying restoration of endothelial dysfunction and pulmonary vascular remodeling are currently under investigation with studies supporting opposite roles for EPCs (both beneficial and detrimental) depending on the origin of these cells as well as their microenvironment comprising neighboring cells and soluble cytokines and growth factors. The latter seem to be responsible for the differentiation route of EPCs either towards mature endothelial cells reconstituting damaged endothelium or smooth muscle cells favoring vascular muscularization and deleterious remodeling.

In this context, a steadily increasing number of studies revealed distinct profiles regarding EPC numbers and function between COPD patients with stable disease and acute exacerbations. In specific, several lines of extensive investigation proposed that stable COPD is associated with a significant reduction in the number of circulating EPCs while a strong correlation with functional parameters of disease severity was also noted [5, 31, 33]. Moreover, a positive correlation between reduced EPC levels and a low body mass index, a marker of systemic impairment, has also been reported in COPD patients [32]. Nonetheless, circulating EPCs have been assessed using different methodology techniques among different studies with conflicting results [30]. On the other hand, it seems rationale to suggest that acute disease exacerbations are associated with a significant increase in CD34+ cells followed by upregulation of serum VEGF concentrations, reflecting either a compensatory response or simply a bystander effect [69].

The mechanisms orchestrating the reduction in EPCs in stable COPD patients have not been delineated yet. However, various premises have been suggested, e.g. impaired mobilization and the proliferative capacity of BM-derived EPCs driven by hypoxemia [31, 70] and/or increased apoptosis of EPCs due mainly to oxidative stress and inflammatory cytokines that directly affect BM production of progenitor cells [31, 71]. An alternative notion supports that diminished circulating levels of EPCs may simply reflect enhanced recruitment in injured pulmonary vessels [27, 71].

Alveolar Epithelial Progenitor Cells and Epithelium Restoration

An alternative approach to the complications arising from exogenously administered stem cells, including low engraftment levels and rejection, is to exploit the stemness of resident lung stem cells. It is becoming increas-ingly apparent that a considerable number of stem cell populations with broad regenerative and differentiative capacity, such as basal, secretory or Clara cells and type-II AECs, reside in small protected niches across the tracheobronchial tree. While in otherwise healthy individuals the above cells are recruited to sites of injury to accelerate tissue repair and restoration, it is believed that in genetically predisposed individuals, e.g. patients with chronic lung disease experiencing repetitive injurious stimuli, impaired mobilization coupled with diminished regenerative capacity and/or limited reservoir of resident stem cells is responsible for abnormal wound healing.

It seems rationale that the re-awakening of developmental pathways being in a dormant state in adult or injured lung may provide an effective alternative to alveologenesis [34]. One such cascade is the retinoid pathway with the major candidate all-trans retinoic acid (atRA) derived from vitamin A (retinol), atRA functions via nuclear retinoic acid receptors and modulates the synthesis of elastin, an essential structural component of the lung matrix in neonatal fibroblasts [34]. Exogenous instillation of atRA in rats induced alveolization [72] whereas mice mutant for retinoic acid receptor genes exhibited disrupted alveolar formation [73].

On the basis of the above experimental data, clinical trials assessing the safety and efficacy of oral administration of atRA in patients with emphysema have been conducted and although an acceptable safety profile was demonstrated, efficacy results were rather disappointing, as evaluated by radiological and functional parameters [74, 75]. However, efficacy endpoints were assessed 6 months after drug administration, a time point which is too short to anticipate structural changes with clinically apparent functional effects. Other biological agents that have been reported to have a regulatory role in alveolar formation in experimental models, including estrogens, hepatocyte growth factor, granulocyte colony-stimulating factor, fibroblast growth factor-7 and statins, are currently under investigation while results from pilot clinical trials are greatly anticipated [34].

Stem Cell Therapy in CF

CF, the most common life-shortening genetic disorder in Caucasians, affecting approximately 70,000 individuals worldwide, is an autosomal recessive genetic disorder that affects most critically the lungs, but also the pancreas, liver and intestines. It is characterized by abnormal transport of chloride and sodium across the epithelium, lead-
ing to thick, viscous secretions. It is caused by a mutation in the gene for the protein CF transmembrane conductance regulator (CFTR) [76, 77]. This protein is required for the transport of chloride and sodium ions across the epithelial membrane. The most common mutation, ΔF508, is a deletion in 3 nucleotides that results in a loss of the amino acid phenylalanine at the 508th position on the protein [76, 77]. The gradually increasing incidence and the poor survival estimated at 37.5 years with no firmly established treatments apart from lung transplantation have urged chest physicians to rapidly develop personalized therapeutic approaches to alter the basic defect in the disease using gene-class-specific therapy, the so-called CFTR modulators [78]. The latter represent therapies directed towards specific disease-causing mutations and the molecular pathways that underlie their cause. Nevertheless, results are preliminary and treatment approaches are still in the developmental stages.

In this context, stem cell technology has also penetrated into this fascinating therapeutic field leading to a number of ramifications, including the study and potential application of ESCs in patients with CF, as human ΔF508 ESCs have been produced but not extensively studied [79]. So far the only studies reporting a method for producing patient-specific airway epithelial cells for disease modeling and in vitro drug testing were recently published by Wong et al. [80, 81]. In particular, investigators intriguingly demonstrated an in vitro differentiation protocol generating functional CFTR-expressing airway epithelium from human ESCs. A proof of concept analysis was also performed in a CF patient showing enhanced plasma membrane localization of mature CFTR protein. Another recent study reported the generation of disease-specific lung progenitor cells from human CF iPSCs that led to the formation of a fully functional respiratory epithelium, providing us with pivotal data for genetic lung diseases [82].

**Stem Cell Therapy in Asthma**

Asthma is a chronic inflammatory disorder of the airways in which many cells play an essential role. This chronic inflammation is associated with airway hyperresponsiveness that leads to recurrent episodes of variable airway obstruction that is often reversible either spontaneously or with treatment [83, 84]. Current treatment of asthma with inhaled corticosteroids and long-acting β2-agonists is highly effective, well tolerated, safe and relatively inexpensive, however many patients remain poorly controlled. While innumerable lines and funds of research are spent to improve these drug classes, major unmet needs, including better treatment of severe asthma (which shares several characteristics with COPD) as well as curative therapies for mild-to-moderate asthma that do not result in symptom recurrence when treatment is tapered or even discontinued, still represent considerable therapeutic shortcomings for chest physicians [83, 85].

In this context, a steadily increasing number of studies demonstrate the efficacy of either locally or systemically administered MSCs in a rapidly expanding spectrum of lung injury models in asthma (table 3). In specific, several groups of investigators either utilizing ovalbumin or even better airway allergens, including ragweed pollen, to produce a clinical and histological phenotype compatible with human asthma observed a beneficial effect of intra-venously or intratracheally instilled MSCs, which was assessed by a significant decrease in airway hyperresponsiveness, eosinophilic Th2 inflammation, histologically documented injury and mucus metaplasia as well as abrogation of IgE serum concentrations [13, 86–90].

Despite the above promising data rendering MSCs wonderful candidates for fruitful therapeutic interventions in patients with severe persistent asthma nonrespon-sive to conventional treatment, efforts to apply this evidence from bench to bedside have been restrained by considerable safety concerns, including possible malignant transformation of MSCs, on a longitudinal basis coupled with highly effective available current therapeutic regimens.

**Future Challenges and Limitations**

For these cell-based therapies to become truly evolutionary and be in the same trajectory as bronchodilators in the therapeutic field of chronic lung diseases, there is a number of challenges and limitations that should be addressed properly.

**Lack of Stem Cell Clinical Trials in Patients with Chronic Lung Diseases**

Although abundant lines of both human and experimental data have provided us with encouraging safety and efficacy data, unfortunately pulmonary and critical care medicine have traditionally lagged behind other fields, including hematology, cardiology and gastroenterology, in translational studies of potential new therapies, e.g. the use of reparative cells [13].
More specifically, two published phase-I clinical trials of EPCs in primary pulmonary hypertension demonstrate both safety and efficacy \cite{11, 91} while a third study exploiting the potential usefulness of autologous progenitor cells to act as vehicles of drug delivery, namely human endothelial nitric oxide synthase in patients with severe pulmonary artery hypertension, is currently under investigation. First anecdotal safety results from the latter study seem promising, and investigators are still recruiting eligible patients. On the other hand, evidence from a fourth study that has completed patient recruitment is still unknown.

Furthermore, the use of stem cell therapy is now being established in patients suffering from complications following acute myocardial infarction. In particular, six randomized placebo-controlled clinical trials estimating safety and efficacy of BM-derived stem cell therapy either locally or systemically administered to patients that have experienced acute myocardial infarction are now currently available in the medical literature. Although efficacy results arising from these studies seem rather confusing and conflicting with three trials reporting negative results and the remaining three demonstrating the effectiveness of stem cell treatment, which was assessed by the improvement in parameters such as left-ventricular function, quality of life and ventricular remodeling, nevertheless all of them were characterized by encouraging safety data \cite{92–97}.

The latter studies offered pivotal clinical insights that helped clinicians to overcome potential safety concerns emerging from the local or systematic delivery route of MSCs. Furthermore, in one of the aforementioned studies, authors came up with an exploratory finding as they reported lung function improvement (FEV\textsubscript{1}) in the majority of patients, indicating either a hemodynamic effect on ejection fraction or alternatively homing of a large proportion of infused cells to the lungs and induction of anti-inflammatory and regenerative processes. The latter observation attracted the interest of chest physicians and triggered the launch of two clinical trials assessing the safety and efficacy of intravenous infusion of BM-derived MSCs in patients with moderate and severe COPD.

The first study was recently published by Ribeiro-Paes et al. \cite{98} who reported a marginal statistically significant improvement in functional parameters as well as in exercise capacity in patients with severe COPD after an intravenous administration of autologous BM-derived MSCs. However, this trial was severely hampered by the limited number of patients enrolled (only 3 patients eligible for analysis) posing major limitations to the data presented. Regarding the second phase-II clinical trial, sponsored by

Table 3. Experimental data of stem cell therapy in animal models of asthma

<table>
<thead>
<tr>
<th>Study year</th>
<th>Type of stem cells</th>
<th>Animal model</th>
<th>Route of administration</th>
<th>Outcome</th>
<th>Potential mechanisms of action</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bonfield et al. 2010</td>
<td>Human BM-MSCs Plastic adherent</td>
<td>OVA-induced AR</td>
<td>IV</td>
<td>Improved histologic injury Decreased BALF inflammatory cytokines (IL-5; IL-13, IFN-γ), serum IgE, BALF iNOS</td>
<td>None specific Potentially soluble mediators</td>
</tr>
<tr>
<td>Cho and Roh 2010</td>
<td>Mouse BM-MSCs</td>
<td>OVA-induced AR</td>
<td>IV</td>
<td>Decreased nasal inflammation, OVA-specific IgE, IgG1, IL-4, IL-5 Increased IFN-γ in cultured splenocytes</td>
<td>None specific Potentially soluble mediators</td>
</tr>
<tr>
<td>Nemeth et al. 2010</td>
<td>Mouse BM-MSCs</td>
<td>Ragweed-induced AAI</td>
<td>1st IV 2nd IT</td>
<td>Decrease in histologic injury, BALF inflammatory cytokines, serum IgE</td>
<td>TGF-β1 secretion</td>
</tr>
<tr>
<td>Park et al. 2010</td>
<td>Mouse ADSCs</td>
<td>OVA-induced AAI</td>
<td>IV</td>
<td>Diminished airway hyperresponsiveness Decreased eosinophilic Th2 lung inflammation Th2 switching to Th1 immune response</td>
<td>None specific Potentially soluble mediators</td>
</tr>
<tr>
<td>Goodwin et al. 2011</td>
<td>Mouse BM-MSCs</td>
<td>OVA-induced AAI</td>
<td>IV</td>
<td>Diminished airway hyperresponsiveness Decreased eosinophilic Th2 lung inflammation Th2 switching to Th1 immune response</td>
<td>None specific Potentially soluble mediators</td>
</tr>
</tbody>
</table>

AAI = Allergic airway inflammation; AR = allergic rhinitis; BALF = bronchoalveolar lavage fluid; IFN = interferon; iNOS = inducible nitric oxide synthase; IV = intravenous; OVA = ovalbumin.
Osiris Pharmaceuticals, recruitment has been completed and a total of 62 patients with a diagnosis of moderate (n = 23) or severe (n = 39) COPD, based on the recent GOLD functional criteria [99], were enrolled and being followed for a period of 2 years in a placebo-controlled study. Despite the great hype that was generated, first anecdotal results are rather disappointing, highlighting the need for a careful study design before rigid conclusions can be drawn. Official findings and publication are greatly anticipated.

On the other hand, safety concerns based on IPF pathogenesis, which is still elusive and controversial, coupled with issues reflecting the origin and the potential of MSCs to differentiate into fibroblasts have severely hampered clinicians’ efforts to apply, so far, cell-based therapies in the treatment of this dismal disease. To address the above concerns and to establish a rigid basis for future efficacy trials, we have conducted a nonrandomized unicentric, dose-ranging safety study of endobronchial infusion of autologous ADSCs (stromal vascular fraction) in IPF patients with moderate disease (FVC >50%, DLCO >35%) pattern. As secondary exploratory endpoints, parameters reflecting functional and radiological disease status have been assessed. A total of 15 patients have been enrolled so far, and although interim analysis [100] in the first 12 recruited patients showed improvement in indicators of exercise capacity and quality of life, final evaluation failed to show such a benefit. Nonetheless, the safety data reported raised clinicians’ hopes and helped them to overcome their fears and concerns accelerating the conduction of well-designed randomized controlled clinical trials in the near future (table 4).

### Setting Realistic Endpoints to Assess the Effectiveness of Stem Cells in Clinical Trials

Based on the above data from pilot studies estimating the safety and efficacy of the intravenous administration of either allogeneic or autologous BM-derived MSCs, it is essential to underline the need for a careful study design by setting feasible primary endpoints in order to reliably assess efficacy. In other words, it seems illusory to anticipate that stem cells promote lung renewal and completely reverse a disease phenotype that advances through years assessed by ameliorations in functional and radiological parameters, exercise capacity or quality of life within a period of 1 year or after 1 single MSC administration. It is far more reasonable to evaluate the effectiveness of cell-based therapies using biomarkers that reflect changes in the molecular phenotype of the airway epithelium and endothelium and estimate whether these alterations produce beneficial effects that are clinically occult but may be of primary clinical importance ultimately.

### Precautions of Stem Cell Treatment

Currently, despite sound information supporting the application of stem cells either as first-line or adjuvant

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Table 4: Human data of stem cell therapy in patients with chronic lung diseases

<table>
<thead>
<tr>
<th>Study year</th>
<th>Disease</th>
<th>Type of stem cells</th>
<th>Route of administration</th>
<th>Patients n</th>
<th>Outcome</th>
<th>Potential mechanisms of action</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ribeiro-Paes et al. [98] 2011</td>
<td>COPD</td>
<td>BM-MSCs</td>
<td>Intravenous</td>
<td>3</td>
<td>Marginal improvement in FEV₁</td>
<td>None specific</td>
</tr>
<tr>
<td>Osiris Pharmaceuticals, 2012</td>
<td>COPD</td>
<td>BM-MSCs</td>
<td>Intravenous</td>
<td>62</td>
<td>No improvement in FEV₁</td>
<td>None specific</td>
</tr>
<tr>
<td>Tzouvelekis et al. [100] 2011</td>
<td>IPF</td>
<td>ADSCs-SVF</td>
<td>Endobronchial</td>
<td>15</td>
<td>Disease stabilization 1 year after first infusion</td>
<td>None specific</td>
</tr>
<tr>
<td>Wong et al. [81] 2012</td>
<td>CF</td>
<td>ESCs</td>
<td>Intravenous</td>
<td>1</td>
<td>Enhanced plasma membrane localization of mature functional CFTR</td>
<td>Construction of mature functional CFTR</td>
</tr>
</tbody>
</table>

6MWD = 6-Minute walking distance; DLCO = diffusing lung capacity for carbon monoxide; FEV₁ = forced expiratory volume in 1 s; FVC = forced vital capacity; QoL = quality of life; SVF = stromal vascular fraction.
treatment for chronic lung diseases, there are several safety precautions precluding their global applicability in the everyday clinical practice that should be addressed properly.

Firstly and most importantly, a still opening question is related to the yet unknown fate and potential mechanisms of actions of these cells once engrafted into a highly inflammatory and potentially dysplastic microenvironment. A panel of experts strongly believes on the fibrogenic and/or tumorigenic capacity of these cells on a longitudinal basis given the close association of chronic lung injury with malignant transformation. Since the risk of malignancy or even ectopic tissue formation is present with any cell type that is propagated ex vivo, experts in the field support the use of minimally manipulated cells, such as unfractonated stromal vascular fraction cells, versus more than minimally manipulated MSCs, such as culture-expanded BM-derived MSCs [3, 7, 101]. At this point, it is crucial to highlight that the use of either allogeneic or autologous MSCs is safe and well tolerable since extensive human and experimental data have starkly demonstrated that MSCs derived either from BM or adipose tissue do not express HLA-DR and seem to be less immunogenic than other cell types since they exert powerful immunosuppressive properties [3, 7, 101]. Furthermore, the therapeutic strategy of re-activating endogenous signals to induce lung regeneration may harbor detrimental effects leading from cell senescence to cell immortality and carcinogenesis.

Secondly, faulty or aberrant engraftment of these systemically administered cells may pose other still underestimated potential risks, including lethal pulmonary emboli or differentiation into bone or other inappropriate cellular structures.

Conclusions

Based on the above data, evidence indicates that an increasing, although not uniform, body of data supports to move forward from animal studies to clinical trials. Salient information arising from seminal clinical observations gives credence to the view that cell-based therapies may be a fruitful therapeutic strategy for lung repair and remodeling after injury. In the past 5 years, we witnessed major advances that increased our current state of knowledge from theoretical discussions to practical considerations. It is anticipated that the few ‘brave’ pilot investigations of the safety of stem cell treatment in chronic lung diseases will excite new fields of research to improve our current understanding of the mechanisms orchestrating lung renewal and sparking the design of large multicenter clinical trials, as it happens in other fields of medicine. Lastly, but most importantly, we should always bear in mind that separating the hope from the hype when informing end-stage lung disease patients represents the most crucial step for the moment.

Acknowledgments

Authors have no competing interests related to this article to declare.
Stem Cell Treatment for Chronic Lung Diseases

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Respiration 2013; 85:179–192


Erratum

The authors of the article entitled ‘Stem Cell Treatment for Chronic Lung Diseases’ [Respiration 2013;85:179–192] wish to publish the following corrections.

On page 182, right column, first paragraph, the following text should be published after the last sentence:
In particular, Germano et al. [50] intratracheally injected, in the BLM-model of lung fibrosis, BM-derived pulmonary progenitor cells that were different from MSCs since they expressed Prominin-1/CD133 and hematopoietic CD45 marker, whereas no expression of typical mesenchymal markers was observed. Intriguingly a reduction of histologic lesions, collagen deposition and BALF inflammatory cells was reported. The latter therapeutic effect was mainly attributed to the production of nitric oxide and was associated with engraftment and differentiation into alveolar type II epithelial cells.

On page 183, table 1, the third line should read as follows:

<table>
<thead>
<tr>
<th>Study year</th>
<th>Type of stem cells</th>
<th>Animal model</th>
<th>Route of administration</th>
<th>Outcome</th>
<th>Potential mechanisms of action</th>
</tr>
</thead>
<tbody>
<tr>
<td>Germano et al. [50] 2009</td>
<td>Mouse Lung BM-derived Prominin-1/CD133+ cells</td>
<td>BLM</td>
<td>Intratracheal</td>
<td>Decreased systemic inflammatory cytokines (IL-1β, IFN-γ, IL-6, IL-8, MIP-1α), BALF inflammatory cells and histologic lesions</td>
<td>Nitric oxide production; alveolar epithelium differentiation</td>
</tr>
</tbody>
</table>