Alagille Syndrome: A New Missense Mutation Detected by Whole-Exome Sequencing in a Case Previously Found to Be Negative by DHPLC and MLPA

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Abstract
Alagille syndrome (ALGS, MIM 118450) is an autosomal dominant, multisystem disorder with high variability. Two genes have been described: \textit{JAG1} and \textit{NOTCH2}. The population prevalence is 1:70,000 based on the presence of neonatal liver disease. The majority of cases (~97%) are caused by haploinsufficiency of the \textit{JAG1} gene on 20p11.2p12, either due to mutations or deletions at the locus. Less than 1% of cases are caused by mutations in \textit{NOTCH2}. The most widely used methods for mutational screening include denaturing high-performance liquid chromatography (DHPLC) and multiplex ligation-dependent probe amplification (MLPA). Very recently, whole-exome sequencing (WES) has become technically feasible due to the recent advances in next-generation sequencing technologies, therefore offering new opportunities for mutations/genes identification. A proband and its family, negative for the presence of mutations in \textit{JAG1} and \textit{NOTCH2} genes by neither DHPLC nor MLPA, were analyzed by WES. A missense mutation, not previously described, in \textit{JAG1} gene was identified. This result shows an improvement in the mutation detection rate due to novel sequencing technology suggesting the strong need to re-analyze all negative cases.

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Alagille syndrome (ALGS, MIM 118450) is an autosomal dominant, multisystem disorder with high variability [Alagille et al., 1975]. Two genes were described as associated to ALGS: \textit{JAG1} [Li et al., 1997] and \textit{NOTCH2} [McDaniell et al., 2006]. Disease general population prevalence is 1:70,000 based on the presence of neonatal liver disease [Danks et al., 1977]. The majority of cases (~97%) are caused by haploinsufficiency of the \textit{JAG1} gene [Li et al., 1997], located on 20p11.2p12, either due to mutations or deletions at the locus. Less than 1% of cases are caused by mutations in \textit{NOTCH2} [McDaniell et al., 2006]. The most widely used methods for mutational screening include denaturing high-performance liquid chromatography (DHPLC) and multiplex ligation-dependent probe amplification (MLPA). Recently, whole-exome sequencing (WES) has become technically feasible due to the cur-
rent advances in sequencing technologies, therefore offering new opportunities for mutation/gene identification in Mendelian disorder studies.

**Materials and Methods**

**Patients**

The proband (III-1) is the first female child of nonconsanguineous Italian parents. At birth, she presented neonatal jaundice, treated with phototherapy for 14 h. At 1 month, poor visual fixation was obvious, and the ophthalmological evaluation revealed retinal hemorrhage with abnormal latency of the visual evoked potentials. Serological examination showed high levels of hepatic enzyme values (AST, ALT and γGT), which remained abnormal during further evaluations. Abdominal ultrasound showed no abnormalities, while cardiac sonography indicated a thoracic aorta hypoplasia. ALGS was suspected, and a liver biopsy confirmed the paucity of intrahepatic bile ducts. At the age of 4, typical facial dysmorphisms of ALGS were clearly evident. Spinal X-rays and ophthalmological examination did not reveal any abnormalities. In the maternal proband’s family, the mother (II-1) presented the same facial dysmorphisms and the coarctation of aorta but with normal liver function. The aunt (II-3) was referred with no clinical symptoms, and the grandfather (I-2), also with facial dysmorphisms, presented unspecified renal failure; however, no further clinical data are available (fig. 1).

**Mutation Analysis**

WES was performed on 3 family members (III-1, II-1 and II-3). Starting from as little as 3 μg of genomic DNA, the exome of each patient was enriched using SureSelect Human All Exon 38Mb Target Enrichment System (Agilent Technologies, Inc.); a whole-exome fragment library was constructed following the manufacturer’s protocols (SureSelect Target Enrichment System for the Applied Biosystems SOLiD System–Version 2.0.1). Paired-end sequencing was performed on the SOLiD4 platform (Life Technologies), and the sequence reads were mapped against the human reference genome (hg19) using SOLiD bioscope software v1.3. Single nucleotide variants (SNVs) were called by SAMTools V0.1.8 [Li et al., 2009] and then filtered according to a dominant inheritance model. Among the SNVs identified, those present in NCBI dbSNP build v132 and in our own database were excluded. Results were confirmed by direct Sanger sequencing.

**Results and Discussion**

The proband (III-1) was referred to be negative at the mutation molecular screening of the entire coding region of JAG1 and NOTCH2 genes; the molecular analysis was done by DHPLC and MLPA. A comprehensive whole-exome sequencing was performed on subjects III-1, II-1 and II-3. The above-described data analysis pipeline allowed the detection of a missense mutation, never described before, in the JAG1 gene (c.1308C>G, p.C436W, NM_00124); this nucleotide variation was confirmed by Sanger sequencing (fig. 2) and has not yet been reported until today in any public databases, such as NCBI dbSNP or in 1000 Genomes Project. The proband (III-1), the mother (II-1) and the grandfather (I-2) were heterozygous (c.1308C>G), while the aunt (II-3) was normal (fig. 2). PolyPhen-2 (http://genetics.bwh.harvard.edu/pph2/), Mutation Tester (http://www.mutationtaster.org/) and Condel (http://bg.upf.edu/condel/home) predict the mutation c.1308C>G, p.C436W, NM_00124 as pathogenic. This mutation affects the EGF-like6 jagged-1 protein region; the replaced cysteine is evolutionarily highly conserved and is involved in disulfide bonds within the EGF-like domain.

A mutation causing a cysteine substitution in the same JAG1 EGF-like6 domain (p.C438F), just 2 amino acids downstream from the one we are now discussing, was already described and its pathogenic role established [Crosnier et al., 1999].

Neither the mutation c.1308C>G nor others were revealed by DHPLC analysis of the entire coding region of JAG1 and NOTCH2 genes; DHPLC mutation discovery rate is lower than 100%, for different reasons, e.g. a large deletion spanning several exons or mutations localized within zones not covered by PCR amplified regions (such as promoters or introns) or, last, it could be due to technical failure [Samejima et al., 2007]. Mutation c.1308C>G was not detected by DHPLC because of technical failure.

Alagille syndrome is a genetic disorder with autosomal dominant inheritance; every affected individual could transmit the mutation and thus, the phenotype to future generations (regardless of the gender of the unborn) with
a risk of 50%. Genetic counseling is very important to inform people of the risks, even though it may encounter difficulties in their ability to define the phenotype. Indeed, no genotype-phenotype correlations exist between clinical manifestations of ALGS and specific JAG1 mutation types or location within the gene [Crosnier et al., 1999]. However, many authors described a specific tissue sensitivity to JAG1 dosage, for example, the heart is more sensitive to JAG1 dosage than the liver [Eldadah et al., 2001; Lu et al., 2003]; this could explain why both, the mother (II-1) and the daughter (III-1), have cardiological involvement, while only the child (III-1) has liver affection.

The improvement of the mutation detection rate observed using WES strategy compared to DHPLC and MLPA approach strongly suggests reanalyzing all negative cases so far reported at mutational screenings (done with lower-resolution techniques). This example highlights the huge impact of new sequencing technologies, including targeted resequencing, in reducing false negative mutation detection rate and offering better clinical management of patients and their families, leading to remarkable improvements of counseling and of diagnostic algorithms.

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**References**


