Gastrointestinal Stromal Tumor: A Rare Abdominal Tumor

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Abstract
Gastrointestinal stromal tumors (GISTs) are rare abdominal tumors which arise from the interstitial cells of Cajal in the gastrointestinal tract. Gastric GISTs are the most commonly seen GIST tumors and may grow to a very large size. They are often associated with abdominal pain, anorexia and weight loss. Most of them can be detected by CT. These tumors have been found to harbor mutations in CD117 which causes constitutional activation of the tyrosine kinase signaling pathway and is considered to be pathognomic. Tyrosine kinase inhibitors such as imatinib have revolutionized the treatment of these tumors, which are otherwise resistant to conventional chemotherapy and radiotherapy. Although surgical resection is the mainstay of treatment, tyrosine kinase inhibitors have been useful in prolonging the recurrence-free survival of these patients. Resistance to imatinib has been reported in GISTs with specific mutations. We present a case of gastric GIST which grew to a very large size and was associated with abdominal pain and weight loss. It was successfully resected and the patient was commenced on imatinib therapy.

Introduction

Gastrointestinal stromal tumors (GISTs) are mesenchymal tumors that may arise from any part of the gastrointestinal tract. GISTs have been shown to originate from the interstitial cells of Cajal, which are located in the submucosal and myenteric plexus of the gastrointestinal tract [1–3]. GISTs are rare tumors and comprise less than 1% of all gastrointestinal tumors, and it is estimated that up to 6,000 new cases are diagnosed in the US every year [4, 5]. This likely represents an underestimation as many smaller-sized GISTs may go undetected [6, 7]. They are rarely detected in children and present with metastasis in adults over the
age of 50 years [8]. An equal distribution in gender has been observed though there have been reports of a male preponderance [9]. GISTs present as round masses with clearly defined borders arising from the submucosal layer. Based on their histology, they are divided into eight subtypes, but all of them stain for the stem cell factor receptor (CD117/KIT) [10]. GISTs which do not express KIT may express alpha-type platelet-derived growth factor receptor (PDGFRA), protein kinase C theta (PKC-θ) and discovered on GIST-1 (DOG-1) [11–13]. Malignancies such as melanoma, mastocytoma, Ewing’s sarcoma, lung small cell carcinoma, etc. may also express KIT, and their presence in the gastrointestinal tract likely represents metastases and they need to be distinguished from GISTS [14–16]. An autosomal dominant pattern of inheritance has been described in familial GIST [17–19]. Mutations involving succinate dehydrogenase subunits B, C and D have been observed in the familial form of GIST [20]. GISTs have been observed in the small intestine of patients with neurofibromatosis type-I [21, 22]. A separate inherited syndrome consisting of paragangliomas and GISTs has also been described [23].

Gastric GISTs are known to reach sizes exceeding 40 cm and have a better prognosis than intestinal GISTs of similar size and mitotic rate [10]. Extraintestinal GISTs (EGISTs) have been reported in the gall bladder, urinary bladder and rectovaginal septum [24–26]. EGISTs are considered to be metastases but paradoxically exhibit a better overall prognosis [27]. CT scans and endoscopic ultrasound are commonly used diagnostic modalities.

**Case Summary**

The patient is a 41-year-old male with a past medical history significant for type 1 diabetes, gastroesophageal reflux disease and hypertension. He presented in our gastroenterology clinic with abdominal discomfort, distention and unintentional weight loss of 24 pounds over the past 2 months. The patient underwent a CT scan of the abdomen which revealed a 20-cm epigastric mass with several areas of necrosis suggestive of hemangioma of the liver. The patient was scheduled to be seen in the general surgery clinic, but his symptoms worsened and he presented to our hospital with a fever of 102°F and worsening abdominal pain. A repeated CT scan of the abdomen revealed a large heterogeneous mass measuring 31 × 10 × 26 cm, predominantly solid and containing small cystic areas in the right abdomen and pelvis. The patient was admitted to the medical service where he became hypotensive and anemic with a significant hematocrit drop. The patient was hemodynamically stabilized and underwent an arterial embolization of the gastroduodenal artery to control the hemorrhage. Thereafter, the patient underwent resection of the tumor en bloc with the stomach, gastrectomy, Billroth II reconstruction and wedge biopsy of segment six of the liver. The patient tolerated the procedure well, with no significant post-operative complications. On gross examination, the tumor measured 34 cm in its greatest dimension, with notable hemorrhage and necrosis (fig. 1). Microscopic examination and immunohistochemical staining confirmed the tumor to be a GIST with tumor cells staining positive for CD117 (fig. 2a, b). Segment 6 of the liver was reported as metastatic GIST. The patient was started on chemotherapy with imatinib.

**Discussion**

The most common site of occurrence for GISTs is the stomach (60%) followed by the small intestine (30%) [28]. It is difficult to distinguish extraintestinal GISTs from metastases.
The mainstay of management of GISTs is surgical resection when possible. Regional lymphadenectomy is not advocated as GISTs have not been observed to metastasize to lymph nodes [29, 30]. However, there is a very high risk of recurrence if there is intraperitoneal rupture or spillage during surgery [31]. The 5-year overall survival in patients with complete resection has been estimated to be superior compared to that of patients with incomplete resection (42 vs. 95%) [32]. Tyrosine kinase inhibitors such as imatinib have been successfully used to treat GISTs [33]. KIT mutations cause constitutive activation of tyrosine kinase and the most common mutations are duplications in the 3’ region of exon 11 of the KIT gene. Mutations involving codons 557–558 and point deletions carry a worse prognosis [34]. Imatinib resistance has been noted in patients with PDGFRA mutations in the absence of KIT mutations [35]. Chromosomal losses at 1p, 14q (gastric GISTs), 15q (intestinal GISTs) and 22q have been observed, and the type of mutations have been noted to correlate with the response to imatinib [36, 37]. Mitotic rate, tumor size and location have been utilized for predicting the risk of progression in completely resected primary GISTs [38, 39].

Considering that GISTs may reach large sizes and therefore making surgical resection risky, neoadjuvant therapy with tyrosine kinase inhibitors has been utilized to reduce their size. This strategy has been used to reduce surgical morbidity and improve the operability of these tumors [40, 41]. However, the mutational status (KIT vs. PDGFRA) of the tumor largely determines the effect of preoperative tyrosine kinase inhibitor therapy and monitoring of response by PET scan is advised. A duration of 4–6 months of neoadjuvant therapy with continuous monitoring has been recommended [42, 43]. Up to 66% of patients with high-risk GISTs who have undergone resection experience recurrence [44]. The American College of Surgeons Oncology Group (ACOSOG) Z9000 clinical trial showed that patients with PDGFRA mutations had the best outcomes with 90% recurrence-free survival at 3 years. The ACOSOG Z9001 trial involved adjuvant therapy with 400 mg of imatinib and resulted in 98% recurrence-free survival [45]. The dosing and the type of tyrosine kinase inhibitors that are optimal for the patient’s treatment depend on the type of mutation [46–48]. Abdominal CT, MRI and PET scans have been used for follow-up and for monitoring the progression of the disease [49]. The follow-up frequency has ranged from months to years and depends on the risk profile of the initial disease [50].

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Disclosure Statement

The authors declare no conflict of interest.

References


Fig. 1. Excised tumor with a part of the stomach. The gastric mucosa can be seen in the lower part of the specimen.

Fig. 2. a Hematoxylin and eosin staining of the tumor shows a mildly eosinophilic cytoplasm. b CD117 staining with a membranous distribution is shown.