Acute Promyelocytic Leukemia with $\text{Flt3-TKD}$ and $\text{WT1}$ Mutations Relapsing in a Testicle and Followed by Systemic Relapse

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Abstract
Extramedullary relapse is a rare phenomenon in patients with acute promyelocytic leukemia (APL), especially that derived from urogenital systems like the testicles. In this report, we describe an APL patient who had received standard induction/maintenance therapy resulting in durable remission for 4.5 years, when he presented with a unilateral testicular mass confirmed as myeloid sarcoma; this was followed by systemic relapse of APL. Retrospective analysis of the involved blood and bone marrow samples at the time of the initial diagnosis revealed a rare point mutation of \text{Flt3-TKD} and a novel mutation of \text{WT1}. These mutations were detected recurrently throughout the course of the disease. After reinduction therapy with arsenic trioxide and all-trans retinoic acid combined with daunorubicin, complete hematological remission was achieved for the ensuing salvage allogeneic hematopoietic stem cell transplant.

Introduction
Acute promyelocytic leukemia (APL) is a distinctive subtype of heterogeneous myeloid malignancies with specific cytogenetic features, characterized by reciprocal translocation between chromosomes 15 and 17, which leads to generation of the $\text{PML-RARA}$ fusion gene and product interfering with the maturation of myeloid cells. Patients with APL often have an outstanding response to all-trans retinoic acid (ATRA)/arsenic trioxide (ATO)-based therapy, with a reported high 5-year disease-free survival rate of 74% [1]. However, some patients inevitably relapse even after adequate consolidation therapies. Similar to other types of myeloid leukemia, APL commonly relapses in the bone marrow; however, an increasing number of extramedullary (EM) recurrences have also been reported. Here we present an APL patient who underwent standard chemotherapy to achieve complete remission followed by routine monitoring of minimal residual disease for 4.5 years, when he presented with soli-
tary testicular involvement prior to systemic relapse. In addition, we also retrospectively screened the preserved pathological samples for several predictive genetic markers that had been well established in acute myeloid leukemia (AML). A rare mutation (p.Asp839Gly, c.2516A>G) in the tyrosine kinase domain (TKD) of the FMS-like tyrosine kinase 3 gene (FLT3-TKD) and a novel Wilms’ tumor 1 gene (WT1) mutation (p.Arg458Pro, c.1373G>C) were detected and found to be correlated with the disease biology. To our knowledge, this is the first report of testicular involvement prior to systemic recurrence of APL, indicating the importance of any EM abnormalities during follow-up and the genetic prognostic factors seldom detected in APL patients.

Case Report

A 32-year-old Chinese man presented to the hematology clinic because of bone pain and gingival bleeding. He was found to have a leukocytosis of 124.40 × 10^9/l consisting of 97% circulating promyelocytes, hemoglobin (Hb) of 10.9 g/dl, and a platelet (FLT) count of 50 × 10^9/l. The coagulation profile showed a prothrombin time (PT) of 15.8 s (normal range 11.8–14.8 s), an international normalization ratio (INR) of 1.31, an activated partial thromboplastin time (APTT) of 49.7 s (normal range 28–45 s), a thrombin time (TT) of 19.6 s (normal range 14–22 s), and fibrinogen (FIB) 1.74 g/l (normal range 2.0–4.0 g/l). Bone marrow smear and biopsy showed neoplastic infiltration of promyelocytes containing bundles of Auer rods, and flow cytometric analysis revealed an aberrant myelogenous phenotype. The ATRA therapy was initiated immediately at a dose of 20 mg, twice a day, and the diagnosis of APL was confirmed by detection of the PML-RARA fusion gene in a bone marrow aspirate sample. At the beginning of induction therapy, daunorubicin (DNR; 60 mg/day for 3 consecutive days) was added to the regimen, and ATO (10 mg/day for 22 days) was administered to replace ATRA. After nearly 40 days of induction therapy, the patient’s bone marrow aspiration indicated morphologic complete remission, but the PML-RARA fusion gene remained detectable. After another 3 cycles of anthracycline-based consolidation chemotherapy (DA regimen, DNR 60 mg/day on days 1–3 plus ATRA 20 mg, twice a day, on days 1–15), the patient achieved complete molecular remission. Thereafter, different regimens including ATO (10 mg/day for 14 consecutive days), ATRA (10 mg, thrice a day, for 28 days), and 6-mercaptopurine (50 mg, twice a day, for 28 days) plus methotrexate (MTX; 10 mg/week, for 4 weeks) were given in succession as maintenance therapy for a total of 8 cycles over the next 2 years. The patient achieved durable remission verified by morphological and molecular examination of the bone marrow every 3 months. In addition, examination of cerebrospinal fluid and intrathecal injection of MTX or arabinosylcytosine were performed regularly, and no evident APL relapse was found during the 2-year maintenance therapy.

At 28 months from the last consolidation treatment, the patient observed slight enlargement of the right testicle. However, it was not thought to be significant at that time because of its asymptomatic progression and normal routine monitoring of minimal leukemias. Subsequently, at the 6-month follow-up, the right testicle became progressively more swollen and then the patient was referred to the Department of Urinary Surgery. A right testicular mass with abnormal density (5.6 × 4.0 cm in size) was detected by computed tomography (CT) (fig. 1), indicative of secondary or therapy-related neoplasm. Restaging scans of the brain, chest, or abdomen did not show any evidence of relapse involvement. The patient’s blood and coagulation tests were completely normal. The involved testicle was subsequently removed and the cut surface of the mass was gray-green. Corresponding pathological examination revealed that the testicle and ipsilateral spermatic cord were infiltrated by small malignant cells with granular cytoplasm (fig. 2a), which appeared to be homogeneously expressing myeloperoxidase (MPO) (fig. 2b) and lacking CD34 (fig. 2c) or CD117 (fig. 2d) by immunohistochemical (IHC) staining. Further detection of the PML-RARA fusion gene by fluorescence in situ hybridization (FISH) confirmed the testicular relapse of APL (fig. 3). However, consecutive evaluation of bone marrow morphology and corresponding PML-RARA fusion gene detection were negative. Therefore, systemic therapy for APL relapse was not administered. At 1 month after orchiectomy, the patient presented to the hospital for spontaneous gingival bleeding unrelated to tooth brushing. He reported no concomitant fever, headache, or vomiting. A physical exam did not reveal any cutaneous petechiae. The laboratory tests showed Hb 14.4 g/dl, PLT 92 × 10^9/l, and WBC 2.56 × 10^9/l, with 3/50 circulating blasts; coagulation function showed PT 15 s, INR 1.34, APTT 16.2 s, TT 27.4 s, and FIB 0.5 g/l. Bone marrow examination found 51% promyelocytes and 11% blasts; flow cytometric analysis of the bone marrow showed 78% blasts positive for CD117 (dim), CD123 (dim), CD13 (dim), and CD33 (dim), which partially expressed CD34 and HLA-DR. The PML-RARA fusion gene was detected at a level of 9.5 × 10^5 copies/ml (PML-RARA/ABL ratio 26.39%) in the bone marrow by quantitative PCR.

In order to understand the unusual relapse pattern of testicular involvement, we performed a series of mutational analyses aimed
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at 11 prognosis-associated genes: \(\text{CEBPA, DNAT3A, FLT3-ITD/TKD, IDH1, IDH2, KIT, K-RAS, NPM1, N-RAS, TET2, and WT1}\), which ultimately lead to the identification of rare \(\text{FLT3-TKD}\) (fig. 4a) and novel \(\text{WT1}\) (fig. 4b) point mutations. The mutations in \(\text{FLT3-TKD}\) and \(\text{WT1}\) loci usually suggest opposite therapeutic responses and prognoses for general non-M3 AML but are seldom detected or interpreted for APL.

Considering that the patient may be at high risk of developing refractory APL with undefined genetic mutations, he was enrolled into the clinical trial of tamibarotene, a synthetic retinoid first approved to be used in relapsed/refractory APL in Japan. However, the patient was randomized to the control group receiving ATRA (20 mg twice a day) combined with ATO (10 mg/day). With the conventional regimen used as reinduction therapy, the patient’s hypofibrinogenemia improved gradually and the differentiation of his aberrant promyelocytes to mature myeloid cells was demonstrated in peripheral blood examination. Although he had developed serious fungal pneumonia during the induction therapy, bone marrow examination at day 40 indicated hematologic complete remission. The patient is planned to undergo contralateral testicle radiotherapy followed by allogeneic stem cell transplant (allo-HSCT) during the second remission for a better outcome.

### Discussion

ATRA/ATO combined with anthracycline is a first-line regimen resulting in long-term survival for most APL patients. Nevertheless, relapse still occurs in approximately 20% of cases [2]. The majority of postremission relapses occur in the bone marrow, but approximately 3% could be present at EM sites [3, 4]. The most frequently

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**Fig. 2.** Hematoxylin and eosin and IHC staining of the specimens from the involved testicle, supporting the EM myelogenous neoplasm.  
**a** H&E staining. ×200.  
**b** Cytoplasmic expression of MPO. ×400.  
**c** IHC staining negative for CD34. ×400.  
**d** IHC staining negative for CD117. ×400.
**Fig. 3.** PML-RARα fusion gene detected by FISH in sample sections of the myeloid sarcoma. PML-RARα dual-color fusion probes were used. PML-RARα positive for 1R2G1F (arrows), an atypical signal pattern for the fusion gene distinguished from the typical 1R1G2F, was detected in 47% of the counted cells, indicating translocation of RARα on chromosome 17 into PML on chromosome 15.

**Fig. 4.** FLT3-TKD and WT1 point mutations detected in this case. A rare mutation of FLT3-TKD (Asp839Gly) and a novel mutation of WT1 (Arg458Pro) were recurrently detected during the treatment. Specimens for mutational analysis were from the preserved bone marrow film at the initial diagnosis, paraffin-coated testicular tissue with myeloid sarcoma, and the peripheral blood sample at EM relapse, respectively. The double peaks (arrows) demonstrate the point mutations. WT = Wild type; M = mutation; JM = juxtamembrane.
documented sites of EM disease (EMD) are the central nervous system and skin [4], but other unusual places have also been reported [3, 5–8]. In total, only 3 patients have been reported with APL-associated testicular involvement to date (table 1). One patient manifested a solitary testicular mass at presentation [9], and the other two suffered from testicular relapse even after successful allo-HSCT at 125 [10] and 69 months [11] from diagnosis, respectively, but no mutational analysis was performed during the treatment in these cases. By contrast, our case did not undertake HSCT and relapsed in a unilateral testicle long after the end of consolidation therapy. Moreover, this patient had an asymptomatic and painless swollen testicle more than half a year before systemic relapse, indicating that isolated EM abnormality may be the earliest clinical sign of systemic relapse of APL, in addition to prognostic factors that may be detected by mutational analysis. For this reason, any EM manifestations during therapy may be noteworthy for leukemic relapse, and risk-adapted therapy should be initiated in due time to block disease progression.

Some clinical characteristics have also been considered to correlate with EMD. According to a study involving 806 APL patients, EM relapse occurred more frequently in patients with high WBC counts (above 10,000/mm³) at diagnosis [4]. In other studies, treatment of APL with ATRA [12] and the occurrence of retinoic acid syndrome [13] were considered to predispose patients to EM relapse, but this was not proved by others [3]. Additionally, insufficient doses of cytotoxic drugs in ATRA-based regimens may fail to yield an effective concentration in the EM sites, which is also a possible contributor to subsequent relapse [14]. With respect to our case, although the remarkable leukocytosis at presentation might be responsible for EM recurrence in the patient, two genetic mutations have also been identified with a potential impact on the clinicobiological characteristics of APL. Two major forms of gain-of-function mutations of FLT3, internal tandem duplication (ITD) and point mutations within the activation loop of the TKD were found in 21–38% and 9–20% of APL cases, respectively [15–20], among which Asp835Tyr (D835Y) is the most frequently detected FLT3-TKD (13.2% in M5, 11.8% in M3v) mutation [21], resulting in constitutive tyrosine phosphorylation boosting neoplastic proliferation. Particularly in this patient, we detected a rare mutation, Arg458Pro (c.1373G>C), resulting in constitutive tyrosine phosphorylation boosting neoplastic proliferation. Additionally, a novel WT1 mutation, Arg458Pro (c.1373G>C), was also detected in our patient, but little is known about the prevalence of WT1 mutations and their significance in APL; the limited information was found in a recent report in which 4 out of 103 APL patients carrying WT1 mutations were predisposed to leukemia relapse [23]. Although WT1 mutations act as well-recognized indicators of a poor prognosis independently of chromosomal aberrations in AML [24], it is not the case for FLT3-TKD, and their prognostic significance in APL is still controversial [17, 20, 25, 26]. Most of the reports showed that some of these mutations are associated with inferior traits of APL. For example, FLT3-ITD mutations, with a higher incidence in M3v and the PML-RARA bcr3 isoform, are associated with a higher WBC count at diagnosis, resulting in a lower remission rate.

### Table 1. APL patients with testicular relapse

<table>
<thead>
<tr>
<th>First author, year</th>
<th>Age, years</th>
<th>Regimens before relapse</th>
<th>Treatment of EMD</th>
<th>Prognosis</th>
</tr>
</thead>
<tbody>
<tr>
<td>Forrest, 1997 [10]</td>
<td>34</td>
<td>HiAra-C+DNR, Ara-C+AMSA, Cis-RA, Allo-HSCT</td>
<td>Orchietomy, irradiation</td>
<td>Died in 2 years</td>
</tr>
<tr>
<td>Gopal, 2005 [9]</td>
<td>27</td>
<td>(Primary myeloid sarcoma)</td>
<td>Orchietomy</td>
<td>Contralateral relapse in 1 year</td>
</tr>
<tr>
<td>Present study</td>
<td>32</td>
<td>ATRA, ATO+DNR, 6-MP, MTX</td>
<td>Orchietomy</td>
<td>Second remission</td>
</tr>
</tbody>
</table>

6-MP = 6-mercaptopurine; AMSA = amsacrine; Ara-C = arabinosylcytosine; Cis-RA = cis-retinoic acid; DLI = donor lymphocyte infusion; HiAra-C = high-dose arabinosylcytosine; IDA = idarubicin; MIT = mitoxantrone; VP-16 = etoposide.
higher induction-related mortality, and a lower 5-year overall survival rate [15, 19, 21, 25, 26]. However, there is little information about the correlation between genetic mutations and EMD in APL [27]; we could not exclude the possibility that the rare genetic mutations observed in this patient are relevant to his peculiar form of relapse and preferential site of myeloid sarcoma development.

With the diagnosis of testicular relapse, there is still a lack of standard therapy for this cohort of patients. Limited experience regarding treatment was acquired from several published case reports, in which patients received diverse or palliative approaches, including focal excision, radiotherapy on the involved field, and systemic administration of conventional agents combined with autologous or allo-HSCT. Novel agents such as gemtuzumab ozogamicin [8] and tamibarotene [7] may effectively induce a second remission. Unfortunately, our patient was randomly assigned to receive conventional therapy in the clinical trial of tamibarotene and missed a chance to verify this therapeutic option. However, cases of successful management of EMD with ATRA [28] and ATO [29] have also been previously published, and we thus still administered ATRA/ATO plus anthracyclines as the reinduction regimen after the clinical trial, resulting in a significant response of hematologic remission appropriate for HSCT.

In summary, we reported a case of unilateral testicle involvement as the sentinel manifestation of systemic relapse in a nontransplant APL patient with recurrent gene mutations, highlighting the importance of carefully monitoring EMD during follow-up, which might be a clinical marker of impending systemic relapse, and the risk stratification with mutational analysis for APL. However, further preclinical studies on these novel mutations and the interpretation of their different mutation statuses during APL progression are still warranted.

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**References**

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