Acetaminophen Attenuates Obesity-Related Renal Injury Through ER-Mediated Stress Mechanisms

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Abstract

Background/Aims: Obesity is an independent risk factor for the development of kidney disease. The purpose of this study was to determine how obesity may contribute to renal damage and whether acetaminophen ingestion can diminish obesity-associated renal cell injury in the obese Zucker rat model. Methods: Male obese Zucker rats (4 weeks old, \textit{n}=6) were treated with acetaminophen (30 mg / kg body weight / day) for 26 weeks. Age matched obese control (OC), obese vehicle (OV, 0.073 mL DMSO/kg/d), and lean Zucker rats (LC) were used to determine the effects of treatment and obesity. Results: Compared to lean control rats, renal lipid deposition, expression of the endoplasmic reticulum (ER) stress protein GRP78 and activation of the ER stress-related eIF2\textalpha-ATF4-C/CHOP, caspase 12, and JNK-MAPK signaling pathways were increased in the obese control and obese vehicle rats. These alterations were associated with the elevated renal cell apoptosis and urinary albumin excretion. Acetaminophen treatment decreased renal lipid deposition, ER-stress related signaling, apoptosis and albuminuria. Conclusion: These data suggest that the protective effects of low dose acetaminophen on renal injury are mediated, at least in part, through attenuation of ER stress.

Introduction

It is estimated that more than two thirds of adults in the United States are overweight or obese [1]. Paralleling this rise in obesity is an increase in the incidence of chronic kidney
disease (CKD) [2-4]. How obesity might increase CKD is not well understood. However, it is well accepted that increased renal lipid levels may play a pivotal role in the pathogenesis of obesity-associated nephropathy. Indeed, recent researches have demonstrated that lipid accumulation can induce functional impairment (lipotoxicity) and cell death (lipoapoptosis) [5, 6]. The mechanism(s) regulating renal lipoapoptosis have not yet been elucidated; however, it has been suggested that the abnormal lipid accumulation can induce endoplasmic reticulum (ER) stress and cellular apoptosis in several tissue types [7-9]. As endoplasmic reticulum plays a critical role in regulating metabolic homeostasis and cellular functions, perturbations of ER homeostasis in pathological conditions such as excess lipid accumulation can lead to abnormal protein translation and folding, also known as unfolded protein response, which if allowed to proceed unchecked can activate apoptotic pathway and result in cell death [8, 9].

Acetaminophen (N-acetyl-p-aminophenol, APAP) is a widely used over-the-counter analgesic and antipyretic drug. Previous data has demonstrated that acetaminophen treatment can be effective in attenuating the increases in body fat deposition subsequent to the initiation of a high-fat diet in mice [10-12]. Whether chronic acetaminophen ingestion can function to reduce renal lipid accumulation and lipotoxicity during the development of obesity, as to our knowledge, not been investigated. To address this gap in our understanding, we examined whether acetaminophen ingestion can reduce renal lipid accumulation and lipoapoptosis in the obese Zucker rats and the potential role that ER stress may play in these processes. Our data suggest that acetaminophen administration at 30mg/kg/d can attenuate albuminuria and renal lipotoxicity in the obese Zucker rat, and that these effects are mediated, at least in part, by diminished ER-stress related signaling.

**Materials and Methods**

**Animal study**

Male obese and lean Zucker rats (3 weeks old) were purchased from Charles River Laboratories and housed two per cage under controlled temperature (~22°C) and humidity (~50%) with a 12:12 h light:dark cycle. Food and water were available ad libitum. After a week for environmental acclimation, the obese Zucker rats were randomly assigned into one of three groups (n = 6 each): obese control (OC), obese vehicle (OV, 0.073 mL DMSO/kg/d), or obese treated with acetaminophen (OT). Acetaminophen (30mg/kg/d) was dissolved in DMSO and then provided to animals via drinking water for 26 weeks as described previously [13]. Age-matched lean Zucker rats were used as controls (LC, n = 6). Animals were euthanized at 30 weeks for the collection of tissues and urine. Urine specimens were centrifuged at 2000 x g for 5 min, and the supernatant was stored at -80 °C. After removal, kidneys were snap frozen in liquid nitrogen and stored at -80 °C. All animal procedures were conducted under the Animal Use Review Board of Marshall University using the criteria outlined by the Association for Assessment and Accreditation of Laboratory Animal Care (AAALAC) as described in the Animal Welfare Act (PL89-544, PL91-979, and PL94-279).

**Measurement of urinary albumin content**

Urinary albumin concentration was determined using an enzyme immunoassay assay kit purchased from Cayman Chemical (Ann Arbor, Michigan), while urinary creatinine content was measured using a colorimetric assay kit (Cayman Chemical; Ann Arbor, Michigan). Urinary albumin was normalized to creatinine concentration and expressed as micrograms albumin per milligrams creatinine.

**Detection of renal lipid accumulation**

Kidneys were sectioned (8 μm) using a IEC Minotome Cryostat, fixed with 4% formaldehyde, and stained with an oil red O staining kit (Poly Scientific R&D Corp, Bay Shore, NY). Images were captured in a blinded manner using an Olympus BX51 light microscope (Olympus America, Melville, NY, USA) at 200× and 400× magnification.

**Determination of renal nuclear DNA fragmentation**

Apoptosis was assessed using a transferase-mediated dUTP nick-end labeling (TUNEL) kit (Roche Applied Science, Indianapolis, IN) as described previously [13, 14]. Images were captured using an Olympus
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**Fig. 1.** Chronic acetaminophen treatment is associated with reduced lipid deposition in the kidney of obese Zucker rat. Renal tissue sections were stained with oil red O. Representative micrographs were captured at 200× magnification (Scale bar = 100 μm) and 400× (Scale bar = 50 μm). LC: lean Zucker control rats; OC: obese Zucker control rats; OV: obese Zucker rats with vehicle (DMSO); OT: obese Zucker rats treated with acetaminophen (30 mg/kg/d) for 26 weeks.

B5X1 fluorescence microscope (Olympus America, Melville, NY) at 400× magnification. Apoptotic index was expressed as the number of TUNEL-positive nuclei per 100 nuclei (TUNEL+/100 Nuclei).

**Immunofluorescence**

Immunofluorescence staining was used to detect the expression and spatial distribution of GRP78 (an ER stress master regulator [15]) and CCAAT-enhancer-binding protein homologous protein (CHOP, a mediator of ER stress-induced apoptosis [16]). Frozen sections were fixed with 4% formaldehyde, blocked with 5% BSA, and then incubated with anti-GRP78 (#ab21685; Abcam, Cambridge, MA) or anti-CHOP antibody (#sc-575; Santa Cruz Biotechnology, Dallas, TX) overnight at 4°C. After washing, sections were incubated with secondary antibody (fluorescein-labeled goat anti-rabbit IgG; Vector Laboratories, Burlingame, CA) for 60 minutes at room temperature. Images were captured using an Olympus BX51 fluorescence microscope (Olympus America, Melville, NY).

**Immunoblotting**

Total protein extracts were prepared and immunoblotting was performed as previously described [13, 17, 18]. Primary antibodies included anti-GRP78 (#ab21685), anti-CHOP (#sc-575), anti-phospho-eIF2α (#3597S; Cell Signaling Technology (CST), Danvers, MA), anti-eIF2α (#9722; CST), anti-ATF4 (#ab1371; Abcam), anti-caspase 12 (#ab62484; Abcam), anti-c-Jun N-terminal kinase (JNK, #9252; CST), anti-phospho-JNK (#9251S; CST), and anti-glyceroldehyde-3-phosphate dehydrogenase (GAPDH, #2118; CST). The secondary antibody was horseradish peroxidase-linked anti-rabbit IgG (CST). Protein level was quantified by AlphaView image analysis software and normalized to the amount of GAPDH of the same animal.

**Statistical analysis**

Quantitative results were presented as mean ± standard error of mean (SEM). Statistical significance was determined by analysis of variance followed by the Tukey’s test using SigmaStat 3.5. Differences were considered to be significant at \( P \leq 0.05 \).

**Results**

*Acetaminophen reduces lipid deposition in the obese kidney*

Oil red O staining suggested that lipid deposition was higher in the glomeruli and tubulointerstitium of the obese control (OC) and vehicle treated (OV) animals than that observed in the lean control (LC), while these obesity-associated changes were markedly reduced in acetaminophen-treated (OT) animals (Fig. 1).
Acetaminophen decreases GRP78 expression in situ in the obese kidney

Elevations in ER stress have been implicated in tissues with excess lipid accumulation [8, 9] and in the pathogenesis of kidney diseases [19, 20]. To determine whether the association between changes in renal lipid deposition and acetaminophen treatment were associated with the alteration in renal ER stress, we examined the expression of the ER stress regulator, GRP78 [15]. Compared to that observed in the lean controls, the expression of GRP78 was significantly elevated in both the glomeruli and tubulointerstitium from obese control and vehicle rats and, importantly, attenuated by acetaminophen treatment (Fig. 2A and B). Taken together, these data suggest that acetaminophen may suppress the activation of renal ER stress in the obese Zucker rat.

Acetaminophen decreases renal eIF2α-ATF4-CHOP pathway in the obese Zucker rat

It has been suggested that increased ER stress can stimulate CHOP (also known as DNA damage-inducible transcript 3) expression, which plays an important role in inducing apoptosis [21]. Immunofluorescence staining showed that CHOP protein content in both glomeruli and tubulointerstitium was higher in the obese control and vehicle treated rats when compared to that seen in the lean controls while it was visibly decreased following chronic low dosage acetaminophen ingestion (Fig. 3A and B). Immunoblotting analysis further confirmed the diminished expression of CHOP in obese kidney by acetaminophen (P ≤ 0.05; Fig. 3C).

To further identify the molecular mechanism responsible for altered CHOP expression in obese kidney and the protective effect of acetaminophen, we next examined the regulation of eIF2α and ATF-4, which are thought to function as the upstream activators of CHOP expression [22]. Compared to lean control animals, the amount of phospho-eIF2α (p-eIF2α) and ATF-4 expression were significantly elevated in obese control and obese vehicle rats (P ≤ 0.05; Figs 3D and E). Consistent with diminished expression of CHOP, the amount of both upstream regulators were attenuated with acetaminophen treatment (P ≤ 0.05; Fig. 3D and 3E). There was no difference in the amount of total eIF2α between the four groups (P = 0.28; Fig. 3D).

Fig. 2. Acetaminophen treatment decreases renal GRP78 protein levels. A and B. Representative images of glomerular (A) and tubulointerstitial (B) GRP78 immunofluorescence (green signal) were captured at 400× magnification (Scale bar = 50 μm, n = 6). C. GRP78 protein expression was analyzed by immunoblotting and normalized to GAPDH of each animal. Data are expressed as mean ± SEM (n = 6). a: Groups without the same letter are significantly different (P ≤ 0.05).
Acetaminophen reduces renal JNK phosphorylation in the obese Zucker rat

Like that observed for elf2α-ATF4-CHOP pathway, the activation of JNK-MAPK is also thought to be involved in ER stress-induced cell death [16]. Compared with the lean controls, the phosphorylation of JNK (p-JNK) was increased in obese control and vehicle rats (P ≤ 0.05), while it was decreased after acetaminophen ingestion (P ≤ 0.05; Fig. 4).
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Acetaminophen diminishes renal caspase-12 activation and cell apoptosis in the obese Zucker rat

Caspase-12, an ER-resident cysteine protease, is known to be essential for ER stress-induced apoptosis [23]. Compared to the lean control animals, the cleaved product of caspase-12 (active form, 42kDa) was significantly increased in obese control and obese vehicle animals (P ≤ 0.05), while it was diminished following acetaminophen treatment (P ≤ 0.05; Fig. 5A). Paralleling this finding, the amount of full length caspase-12 (inactive form, 55kDa) in both OC and OV rats was significantly less than that of lean control and acetaminophen-treated animals (P ≤ 0.05; Fig. 5A).

To examine whether alterations of eIF2α-ATF4-CHOP, JNK-MAPK and caspase-12 were associated with renal apoptosis and whether increased apoptosis, if present, can be decreased with acetaminophen treatment, TUNEL staining was used to determine cell apoptosis. The number of TUNEL-positive cells in the glomeruli was higher in the obese control and vehicle animals than that in the lean controls (P ≤ 0.05), while it was attenuated with chronic acetaminophen ingestion (P ≤ 0.05; Fig. 5B). Similar results of altered TUNEL-positive cells were found in the tubulointerstitium of obese rats as reported previously [13].

Acetaminophen decreases albuminuria in the obese Zucker rat

Given our findings of diminished renal cell apoptosis with acetaminophen ingestion, we next examined whether treatment was also associated decreased urinary albumin excretion (albuminuria), an early sign of renal damage [24]. As an estimate of 24-hour urine albumin excretion and to minimize potential differences in urine concentration between animals,
urine albumin content was normalized to urine creatinine concentration and expressed as the ratio of albumin / creatinine, as suggested elsewhere [25]. Compared to the lean controls, urine albumin / creatinine ratio was significantly higher in obese animals (OC and OV), while it was markedly reduced in acetaminophen-treated animals (Fig. 6; $P \leq 0.05$).

**Discussion**

Increased obesity prevalence and excessive tissue lipid accumulation in obesity have been shown to contribute to the increased kidney dysfunction and renal disease. Using the obese Zucker rat model, we demonstrate that increased renal lipid deposition was associated with increased endoplasmic reticulum stress, cellular apoptosis, and renal damage (albuminuria). In addition, our data suggest that acetaminophen ingestion (30mg/kg/d for 26 weeks) can attenuate renal lipid accumulation (Fig. 1), renal cell apoptosis (Fig. 5B) and albuminuria (Fig. 6) in the young obese Zucker rat. The renoprotective effects of acetaminophen were associated with the suppression of renal ER stress as suggested by the reduction of GRP78 level (Fig. 2), and the inhibition of apoptotic signaling pathway (diminished eIF2α-ATF4-CHOP pathway, and decreased caspase 12 cleavage and JNK phosphorylation, Fig. 3-5A).

Recent studies have demonstrated that obesity is an independent risk factor for both CKD and end-stage renal disease (ESRD) [2-4]. Indeed, it is thought that excess intracellular non-esterified free fatty acid is a contributor to the progression of the renal disease oftentimes associated with obesity [26, 27]. Moreover, FFA overload can also stimulate the synthesis of triglycerides (TGs), very low density lipoprotein (VLDL), and low density lipoprotein (LDL), which may be lipotoxic, especially if the LDL becomes oxidized LDL [2]. In the present study, our analysis of the oil red O staining suggested that lipids are dramatically increased in the glomeruli and moderately elevated in the tubulointerstitium of the obese animals (Fig. 1). Oil red O is a fat-soluble diazo dye that can be used to qualitatively detect the accumulation of not only TGs but also FFA and cholesterol in tissue [28]. Consistent with recent work demonstrating that acetaminophen attenuates high fat diet-induced tissue fat mass [12], here we demonstrated that acetaminophen administration at a low dose (30 mg/kg/day) can reduce lipid deposition in the young obese Zucker kidneys. Whether these decreases in lipid deposition are solely responsible for the reduced renal apoptosis seen in the present study or if acetaminophen plays a direct role in preventing cell death is currently unclear and will require additional experimentation.

Previous data has suggested that proteinuria is often associated with increased obesity in humans and animals [29-31]. Consistent with these findings, we also found evidence of significant albuminuria in the young obese Zucker rats used in the present study. In addition, we also noted that acetaminophen treatment can attenuate the extent of albuminuria (Fig. 6). Paralleling these decreases in urine albumin levels we also found that the number of TUNEL-positive (apoptotic) cells was increased in the kidneys from both obese control and vehicle animals but diminished with acetaminophen treatment (Fig. 5B). Taken together, these
findings suggest that reduced kidney lipid deposition by acetaminophen might function to attenuate obesity-associated renal injuries.

Although the mechanisms involved in renal lipotoxicity have not been fully elucidated, accumulated experimental evidence has suggested that elevations in ER stress may be involved [32]. The ER is a principal site for protein and lipid biosynthesis, protein folding, and calcium storage and signaling. Under conditions of nutrient excess, insulin resistance and diabetes, tissue lipid metabolism can become abnormal causing elevations in ER stress [9, 33]. To examine the possible mechanism of acetaminophen with renoprotective effect, we investigated the expression and spatial distribution of GRP78, a master regulator of ER stress [15]. Consistent with our findings of reduced renal lipid accumulation and cell injury, we found that renal GRP78 was significantly increased with obesity but diminished with acetaminophen treatment (Fig. 2), suggesting that the renoprotective effect of acetaminophen may be related to its ability to attenuate obesity-associated renal ER stress.

Emerging evidences have suggested that the eIF2α-ATF4-CHOP signaling pathway is involved in FFA-induced apoptosis in human renal proximal tubular cells [8, 34]. Supporting this contention, we found increased levels of eIF2α phosphorylation (activation) and elevations in the expression of ATF4 and CHOP in the obese kidney, and importantly, that these changes were diminished subsequent to acetaminophen treatment (Fig. 3). Consistent with these data, we also observed other important ER-stress mediators, including JNK phosphorylation and caspase-12 cleavage (activation) [16, 23], were increased in the obese kidneys but diminished following acetaminophen treatment (Fig. 4 and 5).

In summary, the data from this study demonstrate that low dose acetaminophen ingestion (30 mg/kg/d) can reduce renal lipid accumulation and the degree of cell injury observed during obesity progression in the young obese Zucker rat. This dosage, if normalized on the basis of body surface area [35], is roughly equivalent to a human dose of approximately 4.9 mg/kg/d (or 367.5 mg/d for a 75-kg person) which suggests that it may be potentially useful for clinical application if long term treatment efficacy and safety can be confirmed. We also demonstrate for the first time that the renoprotective effects of acetaminophen may be mediated, at least in part, by suppressing ER stress and ER stress related apoptosis (Fig. 7).

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Disclosure Statement

All the authors declare that they have no competing interests.

References


