Kidney Regeneration in Mammals

Hai-Chun Yang    Shao-Jun Liu    Agnes B. Fogo
Department of Pathology, Microbiology and Immunology, Vanderbilt University Medical Center, Nashville, Tenn., USA

Introduction

The principles of strategies for progressive kidney diseases focus on control of the blood pressure, decreasing proteinuria, and immunosuppression. However, in most cases, these approaches only delay the progression of end-stage renal disease and do not achieve efficient long-term stabilization, let alone improvement, of renal function. In mammals, several organs such as the skin and liver have a great capacity for regeneration. The kidney also has an innate capacity for regeneration, especially after tubular necrosis. However, this capacity is limited, particularly for glomerular remodeling after injury, perhaps due to its complex cells and architecture. We will discuss potential mechanisms of kidney regeneration and whether this goal can be achieved in mammals.

What Is Kidney Regeneration?

In biology, regeneration is the process of renewal, restoration, and growth that results in cells, tissues, and organs being restored to their natural morphology and function. Regeneration can be either complete, when the new tissue is the same as the lost tissue, or incomplete, as
for instance when necrotic tissue becomes fibrotic. Regression or remission of disease is on the opposite end of the spectrum of disease outcomes in relation to progression, the characteristic phenomenon of chronic kidney disease. Regeneration is the process of tissue repair, including regression of the disease, structure remodeling, and new organogenesis.

Attenuation of the regenerative capacity during evolution is a general phenomenon, presumably due to increased organism complexity during evolutionary adaptations [1]. In fish and other lower branches of the animal kingdom, nephron progenitors are retained throughout life, and these progenitors can replace damaged or lost nephrons and thereby restore the adult nephron (so-called nephron neogenesis). In contrast, in mammals nephron progenitors are lost at the end of kidney organogenesis once the adult number of nephrons has been reached. During postnatal growth, increases in renal demand are met by concomitant increases in nephron size (hyperplasia and hypertrophy) and filtration [2]. Thus, kidney regeneration in mammals can only modify the nephron structure and function but not increase the nephron number.

Different parts of the nephron have different capacities for regeneration. Tubules, especially the proximal tubule, can regenerate after an acute injury. In contrast, the complex architecture of the glomerulus makes its regeneration the most difficult challenge for organ regenerative therapy. In this review, we will focus on the regeneration of injured glomeruli.

### Evidence of Kidney Regeneration

Several clinical observations support the potential of kidney regeneration. The precedent that some early-to-moderate glomerular lesions can regress comes from biopsy analysis of diabetic patients who received pancreas transplants [3, 4]. In the REIN core study of nondiabetic nephrotic patients, ramipril treatment for 2 years resulted in amelioration of the glomerular filtration rate (GFR) decline to a yearly loss similar to that which occurs with normal aging. Further, 10 of 78 treated patients in this study showed improvement of the GFR and never reached end-stage renal disease [5]. In another long-term follow-up study on diabetic nephropathy, 7 of 42 patients on captopril had full remission of proteinuria and stabilized renal function. After an 8-year extended follow-up period, the GFR remained stable in 6 patients who still had less than 1 g/24 h of proteinuria [6]. These findings show that remission and even regression of the functional parameters of chronic kidney disease can occur in humans. Whether these functional improvements were contributed to in part by any regression of structural injury remains unknown.

Animal studies may help to clarify the significance of findings in humans and support the possibility of structural lesion regression. A recent study showed that replacing leptin reversed the diabetic nephropathy lesions modeled by the leptin-deficient BTBR ob/ob mouse [7]. We and other groups have proven that high-dose ACE inhibitor angiotensin receptor blocker (ARB) or inhibition of aldosterone can reverse proteinuria and glomerulosclerosis in models of chronic puromycin aminonucleoside nephopathy, hypertensive injury, and subtotal nephrectomy and in aging rats [8–12]. The high-dose ACE inhibitor remodeled the glomerular architecture by reducing the sclerosis volume and enlarging the volume of intact capillaries by up to 40% [13]. Mesangial and endothelial cell proliferation was reversed, but the number of podocytes per glomerulus was not changed by ACE inhibitor treatment [14]. We also found that glomeruli in rats with progressive sclerosis were enlarged but had a reduced number of capillary segments and capillary branch points and a decreased complexity of the glomerular network compared to normal glomeruli. In contrast, in rats with regression of sclerosis induced by ARB, glomerular enlargement was due to an increased number of glomerular capillary segments and capillary branch points and restored complexity of the capillary network [15]. These structural changes support that regeneration of portions of glomeruli with segmental sclerosis may occur.

### Mechanisms of Kidney Regeneration

Kidney regeneration may occur through various steps, including cell reconstitution, matrix remodeling, and tissue reorganization. Cell reconstitution involves the proliferation and transdifferentiation of remnant healthy cells, renal progenitor cells, and/or stem cells. After acute kidney injury, surviving resident tubular epithelial cells dedifferentiate and subsequently reenter the cell cycle and replace the injured necrotic tubular epithelium, which may be driven by cells surrounding the tubular compartment through secreted ligands [16]. In glomeruli, mesangial cells and endothelial cells readily proliferate and can be replenished after injury, but podocytes do not, due to a high expression of the cyclin-dependent kinase.
inhibitor p27kip1 [17]. Recently, reversal of glomerulosclerosis in aged MWF rats was observed after high-dose ACE inhibitor, linked to podocyte proliferation, regenerated from the parietal epithelium [18]. Cells of the renin lineage may also enhance glomerular regeneration by serving as progenitors for glomerular epithelial cells and podocytes [19].

Cellular regeneration alone appears to be ineffective in the face of a chronic insult, when increased matrix accumulation modulates cell migration and function. Matrix accumulation is modulated by proteases and their inhibitors, such as the matrix metalloproteases (MMPs) and tissue inhibitors of MMPs (TIMPs), and by factors that promote matrix synthesis, including transforming growth factor-β1 (TGF-β1). In our studies of regression in the remnant kidney model, high-dose angiotensin inhibition did not increase MMP mRNA or activity or reduce TGF-β1 mRNA. However, regression by ARB was tightly linked to decreases in plasminogen activator inhibitor 1 (PAI-1) and TIMP-1 [11, 20]. We also observed that thymosin-β4 is increased in early glomerulosclerosis and is required for angiotensin II-induced PAI-1 expression in glomerular endothelial cells [21]. Thymosin-β4 is degraded via a 2-step process to the antifibrotic tetrapeptide N-acetyl-seryl-aspartyl-lysyl-proline (Ac-SDKP), with the initial step mediated by unknown protease(s) and then by prolyl oligopeptidase. Ac-SDKP is degraded by ACE and thus increased by the ACE inhibitor. The regression induced by the ACE inhibitor may be, at least partially, related to the effects of thymosin-β4 and Ac-SDKP.

Chronic kidney disease is characterized by a progressive loss of glomerular capillaries. The podocyte normally secretes specific growth factors, including vascular endothelial growth factor-A (VEGF-A) and angiopoietin-1, which are key for maintaining normal glomerular endothelial function and fenestration. Glomerular VEGF was decreased in the remnant kidney model and, conversely, treatment with VEGF ameliorated the development of glomerulosclerosis and tubulointerstitial fibrosis [22]. In addition to its effects on blood pressure, extracellular matrix, and other growth factors, ARB also promotes direct beneficial effects on podocytes and, in turn, glomerular endothelial cell growth [23]. Elegant studies in a conditional transgenic mouse model demonstrated that a sequential reduction in podocyte VEGF-A levels led to a loss of endothelial fenestrations or failure of fenestrations to form [24]. Our ongoing study supports that over-expression of VEGF in podocytes after glomerular injury stimulates tip cell formation and may accelerate angiogenesis. Hence, it is proposed that revascularization might be a key process to restore kidney function and regenerate the glomerulus.

**Current Approaches and Future Directions**

Although the regression of glomerulosclerosis was achieved in animal studies via angiotensin inhibition, it did not occur in all animals and was not sustained in the long term. We have previously shown that ARB induces the regression of existing glomerulosclerosis in about two thirds of rats during a relatively short period after 5/6 nephrectomy (week 8 to week 12). However, long-term follow-up (from week 8 to week 30) showed only a survival benefit of ARB but no regression of glomerulosclerosis. Further, the glomerular structure was not completely returned to normal by using the ACE inhibitor or ARB, suggesting that additional injury mechanisms such as reactive oxygen species and abnormal cell growth promoting sclerosis were still active. We and others have found that the combination of an aldosterone synthesis inhibitor and ARB, or the combination of PPAR-γ agonists and ARB, induces more regression of glomerulosclerosis in the 5/6 nephrectomy model. These additional effects were related to less inflammation and more podocyte protection [25]. Combination therapy with an ACEI, ARB, and statin achieved even better results in sclerosis than monotherapy with any one of these classes of drug [26]. Thus, a multiphasmaceutical strategy involving novel anti-inflammatory and antifibrotic molecules should be considered in order to potentiate kidney regeneration.

Regeneration does not uniformly occur even with multimodal therapeutic interventions. This may be dependent not only on the disease model but also on the disease stage. Our long-term study in 5/6 nephrectomy rats showed that earlier stages of sclerosis responded more to ARB than later stages. Our mathematical modeling indicated that individual glomerular tufts with sclerosis occupying >50% of the capillaries are doomed to progression. Conversely, glomeruli with <50% sclerosis of the tuft are capable of growing new capillary loops. Elegant morphometric studies of CKD in children and rats have demonstrated that both lengthening and branching of glomerular capillaries occurs after injury [27].

Tissue engineering, cell-based therapies and novel agents beyond renin-angiotensin-aldosterone system blockade are among the major fields of investigation in kidney regeneration. The creation of a biocompatible kidney using scaffolds as a skeleton in which to seed renal
cells or stem cells would be an ideal solution for organ shortages. However, the complexity of the kidney structure is a formidable obstacle. Alternatively, the application of stem cells, direct lineage conversion, and reprogramming of existing cells to kidney progenitor cells may be feasible approaches to slowing the progression of chronic kidney disease or even reversing it.

In summary, kidney regeneration in mammals is feasible but limited. Cell reconstitution, matrix remodeling, and tissue reorganization are major mechanisms for regeneration. Multipronged and early interventions are future choices for the induction of kidney regeneration.

References
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