β-Elemene Reverses Chemoresistance of Breast Cancer via Regulating MDR-Related MicroRNA Expression

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Key Words
Breast cancer • Chemoresistance • β-elemene • miRNA

Abstract

Background: Multidrug resistance (MDR) directly contributes to the clinical failure of chemotherapy in breast cancer (BCA). β-elemene is a natural antitumor drug from plants. We previously confirmed that MDR could be reversed by β-elemene. In this study, we intended to investigate the reversal effect of β-elemene on MDR in human BCA adriacin (Adr) - resistant MCF-7 cells (MCF-7/Adr) and docetaxel (Doc) - resistant MCF-7 cells (MCF-7/Doc) through the gene regulatory network. Methods: MTT-cytotoxic, miRNA microarray, Real-time quantitative PCR, Dual Luciferase Activity Asssay, Western blot analysis were performed to investigate the impact of β-elemene on chemo-resistant BCA cell survival, and its impact on the expression of chemo-resistance specific miRNA and the downstream target genes PTEN and Pgp. Results: Compared with the miRNAs expression profiles of MCF-7/Adr and MCF-7/Doc breast cancer cells from our previous studies, there were 322 differentially expressed miRNAs in MCF-7/Adr and MCF-7/Doc breast cancer cells with β-elemene intervention (50μM/L) for 30h, and 6 miRNAs were significantly up-regulated and 12 miRNAs were significantly down-regulated in both MCF-7/Adr and MCF-7/Doc. We have testified that 5 miRNA is related to MDR before, in this study, the expression of miR-34a, miR-222, miR-452 and miR-29a can lead to changes of the characteristics of chemo-resistant MCF-7/Adr and MCF-7/Doc. The PTEN expression under intervention of β-elemene was significantly increased and Pgp expression under β-elemene intervention was significantly decreased in both cell lines. Conclusions: β-elemene could influence MDR related miRNA expression and subsequently regulate the...
expression of the target genes PTEN and Pgp, which may lead to reduction of the viability of the chemo-resistant breast cancer cells.

**Introduction**

Breast cancer is the most common cancer in women and a major cause of cancer mortality. Current treatment strategies combine surgery with adjuvant therapy, but chemoresistance and toxicity are the leading causes that limit the success of treatment towards the aggressive breast cancer cases. Elemene (1-methyl-1-vinyl-2, 4-Diisopropenyl-cyclohexane) isolated from the Chinese medicinal herb Rhizoma Zedoariae, is a novel noncytotoxic anticancer drug [1, 2]. The extract of elemene is a mixture of β, δ and γ-elemene, with β-elemene as the main component, accounting for 60-72% of the three isoforms. Previous studies have provided abundant evidence to reveal that β-elemene might be an effective MDR reversing agent in cancer chemotherapy and mainly via inhibition of the transport activity of Pgp [3, 4]. However the underlinel mechanism has not been fully elucidated.

MicroRNAs (miRNAs) are a new class of small, nonprotein-encoding RNAs that range in size from 19 to 25 nucleotides (nt) and have important roles in a variety of biologic processes [5-7], and also have a very important role in tumorigenesis, development, cellular migration, apoptosis, signal transduction and carcinogenesis. Recently, accumulating evidence is revealing an important role of miRNAs in anticancer drug resistance and miRNAs expression profiling can be correlated with the development of anticancer drug resistance, such as miR-21, miR-22, miR-155, miR-181a, miR-34a, miR-222, etc. There are several mechanisms have been shown to be targeted by miRNAs in drug-resistant breast cancer such as DNA repair [8-11].

There are data which suggest that 90% of patients who died of cancer are connected with chemoresistance. Adriamycin (Adr) and Docetaxel (Doc) are two of the most common chemotherapy drugs. One important reason for the failure of chemotherapy is primary or acquired resistance. Multi-drug resistance (MDR) means that tumor cells with long-term exposure to a single chemotherapy drug may become resistant to a wide range of different structures -- different targets of anticancer drugs. The combined treatment of β-elemene with ADR or DOC at non-effect dosage lead to higher inhibition efficiencies and increased cell death rate, implying the excellent ability of β-elemene in reversing the multi-drug resistance of MCF-7 cells. We can currently propose that β-elemene with anti-cancer agents may be effective in multi-drug resistant breast cancer by down-regulating MDR1 proteins [12].

From recent studies, β-elemene has revealed to have an apparent synergistic effect over chemotherapeutic agents in cancer cells. However, there has been no report to demonstrate the mechanisms of β-elemene to reverse MDR in breast cancer from miRNA levels. Our team conducted the profiling of miRNAs expression in MCF-7/Adr and MCF-7/Doc cell lines, we are the first to report the findings and testify that 5 miRNA is related to MDR. We also revealed that β-elemene modulated the expression of MDR-related miRNAs and proteins, which may contribute to reversing the BCA chemo-resistance. We propose a logical hypothesis: β-elemene could mediate the MDR specific miRNA, which could then regulate the downstream target and corresponding target genes through the gene regulatory network to interrupt the development process of drug-resistance in cancer cells, hence to improve the treatment efficacy.

**Materials and Methods**

**Cell culture**

Human breast cancer cell line MCF-7 was purchased from ATCC (Rockville, MD). The resistant sublines, selected at 100nm docetaxel (MCF-7/Doc) or at 500nm Adriamycin (MCF-7/Adr), were successfully
established from human breast cancer parental cell line MCF-7 by exposing MCF-7 to gradually increasing concentrations of Doc or Adr in vitro in our laboratory. The IC50 (inhibitory concentration to produce 50% cell death) values of Adr were 403.56 and 0.66μM for MCF-7/Adr and MCF-7/S cells, respectively. The IC50 values of Doc in MCF-7/Doc and MCF-7/S cells were 68.31 and 3.08μM, respectively. All cell lines were cultured in DMEM high glucose (HyClone), supplemented with 10% fetal bovine serum (Gibco) in a humidified atmosphere containing 5% CO₂ at 37°C.

**MTT-cytotoxic**

Cells were seeded into 96-well plates (6×10³ cells/well), treated with different concentrations of β-elemene and incubated for 48 hours. Then 20µl of MTT solution (5mg/ml) was added to each well and the cells were maintained in a humidified atmosphere for 3-4 hours at 37°C. The MTT-containing medium was removed and 150µL of DMSO (AMRESCO, America) was added to each well; each experiment was performed in quadruplicate. The absorbance was measured at 570nm using CliniBio128 (ASYS-Hitech, Austria).

**Total RNA extraction and miRNA microarray**

Total RNA including miRNAs was extracted using MirVana miRNA Isolation Kit (Ambion, AM1560). The concentration and quality of the RNA were measured by the UV absorbance at 260 and 280 nm (260/280 nm) on Nanodrop 2000 spectrophotometry (Thermo Scientific) and by formaldehyde denaturing gel electrophoresis. The RNA was labeled using the FlashTag RNA Labeling Kit (Genishere), according to Affymetrix manufacturer’s recommendations. First, poly(A) tailing was carried out at 37 °C for 15 min in a volume of 15 μl reaction mix, which contains 1× Reaction Buffer, 1.5 μl 25mM MnCl2, 1 μl 1:500 diluted ATP Mix and 1 μl PAP enzyme. Second, FlashTag Ligation was performed at room temperature for 30 min by adding 4 μl of 5× FlashTag Ligation Mix Biotin and 2 μl T4 DNA Ligase into the 15 μl of reaction mix. 2.5 μl of Stop Solution was added to stop the reaction. Hybridization and washing were performed using the Affymetrix Fluidics Station 450 and Hybridization Oven 640 under standard conditions. Image processing was conducted using the Affymetrix GeneChip 2.0 Array contains 15,644 probe sets including 1105 human mature miRNAs. The raw data was treated using miRNA QC tool software (Affymetrix). The data output was received in Excel spreadsheets containing the normalized micro-RNA expression profiles. Differentially expressed miRNAs were filtered to exclude those changes less than 2.0-fold compared with MCF-7/S.

**Real-time quantitative PCR**

Total RNA was extracted using TRIzol® Reagent (Invitrogen, Carlsbad, CA); afterwards a reverse transcription was done using TaqMan® MicroRNA Reverse Transcription Kit (Applied Biosystems, Foster City, CA); mature miRNA was spotted using TaqMan® MicroRNA Assay (Applied Biosystems, Foster City, CA); all procedures were done according to manufacturer’s instructions. Relative expression levers were calculated using the ΔΔCt method, normalized with endogenous control and was presented along with negative control. Clustal X software was used to analyze measured miRNA sequences; the sequences were similarity not high in a reverse transcription system. All reverse transcriptions and PCR assays were presented in triplicate.

**Dual Luciferase Activity Assay**

In order to facilitate observation, a recombinant lentiviral vector stable expression of green fluorescent protein was used in breast cancer cell lines. MCF-7/Doc and MCF-7 cells in logarithmic growth phase were seeded in 24-well plates (3×10⁴ cells/well) after digestion until cell fusion becomes 50% to 60%; which was carried out in accordance with reagent instruction lentivirus infections. After 72 hours, the collected fluorescence was stronger in each well, which resurfaced after digestion was covered with 50% to 60%; added 2μg/ml puromycin to screen. After one week, 1μg/ml puromycin was added to maintain the pressure; three generations were continued to be cultured to observe the expression of the green fluorescence. Then, the green fluorescence MCF-7 cells, MCF-7/Adr cells and MCF-7/Doc cells were inoculated for 24 hours in equal amounts with β-elemene intervention; then afterwards intervened with Doc and Adr for 24 hours. Luciferase activities were measured using a Dual Luciferase Reporter Assay System (Promega, USA) according to the manufacturer’s instruction and the renilla luciferase activity was normalized.
Western blot

Total protein was extracted and lysed in the RIPA buffer (Beyotime, Jiangsu, China). Equal amounts of proteins were separated by 10% SDS-PAGE and transferred to the polyvinylidene difluoride membranes (Sigma, Germany). After blocking with 5% skim milk, the membranes were incubated with primary antibodies against human PTEN and Pgp (1:100, Abcam, America) overnight at 4°C; after washing with TBS, the horseradish peroxidase-conjugated secondary antibody (Kangwei Ltd., Beijing, China) was further incubated; the protein band was visualized by Chemiluminescence with Pierce ECL kits (Millipore, Billerica, MA). β-actin (1:4000, Bioworld, MN) was used as an internal load to normalize the expression patterns of each sample. Three separate experiments were performed to show the protein expression.

Statistical analysis

All experiments were performed in triplicate and a representative data was shown from three separate experiments. A statistical analysis was performed using a t-test or One-way ANOVA and Spearman rank test with a SPSS 16.0 statistic. All experiments were performed in triplicate; p<0.05 was considered statistically significant.

Results

The viability of chemo-resistant breast cancer cell lines after intervention with different concentrations of β-elemene

Some studies show that β-elemene treatment effect human breast cancer cells. In this study, we utilized different β-elemene concentrations and exposure times to determine its impact on cell proliferation. As shown in Fig. 1, the viability of MCF-7/Doc (Fig. 1A), MCF-
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Expression profile of miRNAs in MCF-7/Doc and MCF-7/Adr cells with β-elemene intervention

In our previous studies, we tested the expression profile of miRNAs in MCF-7/Adr, MCF-7/Doc and MCF-7/S cells. Compared with MCF-7/S cell line, there were 183 differentially expressed miRNAs (at least 2.0-fold changes) in MCF-7/Adr and MCF-7/Doc cells. Among the 183 miRNAs, 10 miRNAs were up-regulated, while 26 miRNAs were down-regulated in both MCF-7/Adr and MCF-7/Doc cells [10].

In this study, the expression profile of miRNAs in MCF-7/Doc cells and MCF-7/Adr cells with β-elemene (50μM/L) intervention for 30h were evaluated using an Affymetrix GeneChip miRNA 2.0 Array; screened differentially expressed miRNA and validated through real-time quantitative PCR (primer stem-loop RT-PCR method). Compared with MCF-7/Doc and MCF-7/Adr cells without β-elemene intervention, there were 322 differentially expressed miRNAs among 1,200 miRNAs (criteria differences for Ratio > 2.0 or < 0.5, compared to MCF-7). Among the 322 miRNAs, 65 miRNAs were correlated with the constant changes of the MDR in two cell lines, 18 miRNAs were up-regulated, and 47 miRNAs were down-regulated in both MCF-7/Adr and MCF-7/Doc. 89 miRNAs were up-regulated and 56 miRNAs were down-regulated in MCF-7/Doc only, 109 miRNAs were up-regulated and 68 miRNAs were down-regulated in MCF-7/Adr only. There were 25 miRNAs up-regulated in MCF-7/Doc but down-regulated in MCF-7/Adr; and 21 miRNAs down-regulated in MCF-7/Doc but up-regulated in MCF-7/Adr (Fig. 3).

Among the 322 miRNAs, 6 miRNAs were up-regulated, and 12 miRNAs were down-regulated in both MCF-7/Adr and MCF-7/Doc cells significantly (criteria differences for Ratio > 4.0 or < 0.2, compared to MCF-7) (Table 1).

Table 1. Significantly changed miRNAs by miRNA microarray of the expression profile in MCF-7/Doc and MCF-7/Adr cells with β-elemene intervention (50μM/L) for 30h compared with the expression of miRNAs in MCF-7/Doc and MCF-7/Adr cells we tested before and confirmed targets or pathway, (Criteria differences for Ratio > 2.0, compared to MCF-7)

<table>
<thead>
<tr>
<th>miRNA</th>
<th>FC</th>
<th>Confirmed target or pathway</th>
<th>miRNA</th>
<th>FC</th>
<th>Confirmed targets or pathway</th>
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<td>miR-155</td>
<td>miR-503</td>
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<td>miR-29a</td>
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<td>miR-34a</td>
<td>6.83</td>
<td>NOTCH1, BIRC3, miR-145</td>
<td>miR-424</td>
<td>3.24</td>
<td>PTEN</td>
</tr>
<tr>
<td>miR-4284</td>
<td>2.29</td>
<td>miR-455</td>
<td>miR-503</td>
<td>2.21</td>
<td>P73</td>
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<td>miR-503</td>
<td>miR-503</td>
<td>2.21</td>
<td>BCL-2</td>
</tr>
<tr>
<td>miR-193b</td>
<td>2.99</td>
<td>miR-29a, PTEN</td>
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<td>PTEN</td>
</tr>
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<td></td>
<td>Mir-101a</td>
<td>6.63</td>
<td>BCRP/ABCG2</td>
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</table>
β-elemene can reverse chemo-resistance

After MCF-7/Doc and MCF-7/Adr cells with β-elemene intervention (50μM/L) for 30h, divided into two groups, the Doc treated and Doc untreated group, and the Adr treated and Adr untreated group. By Doc and Adr, remnants from green fluorescent cells had significant differences between MCF-7/Doc, MCF-7/Adr and MCF-7 cells; proving the existence of MCF-7/Doc and MCF-7/Adr drug resistance. However, as seen in Figure 4C, 4D, after treated with 50nm Doc or 250nm Adr for 30h, consistent with a decrease of the residual GFP number, MCF-7/Doc or MCF-7/Adr cells compared to MCF-7/Doc or MCF-7/Adr cells with β-elemene intervention, we can show that co-culture with β-elemene with MCF-7/Doc or MCF-7/Adr may significantly promote apoptosis induced by toxic insult. These suggested that MCF-7/Doc and MCF-7/Adr with β-elemene intervention, could potentially reverse chemoresistance to recipient cells.

β-elemene reverses breast cancer cell resistance by mediating related-miRNA

In previous study, we also testified that 5 miRNA is related to MDR (miR-34a↓, miR-130a↑, miR-29a↑, miR-222↑ and miR-452↑). The MCF-7/Adr and MCF-7/Doc cells after β-elemene intervention (50μM/L) for 30h; and the expression of five drug-specific miRNAs that were compared to MCF-7/Adr and MCF-7/Doc drug resistance. However, as seen in Figure 4C, 4D, after treated with 50nm Doc or 250nm Adr for 30h, Consistent with a decrease of the residual GFP number, MCF-7/Doc or MCF-7/Adr cells compared to MCF-7/Doc or MCF-7/Adr cells with β-elemene intervention, we can show that co-culture with β-elemene with MCF-7/Doc or MCF-7/Adr may significantly promote apoptosis induced by toxic insult. These suggested that MCF-7/Doc and MCF-7/Adr with β-elemene intervention, could potentially reverse chemoresistance to recipient cells.

β-elemene treatment alters the expression of PTEN and Pgp protein in breast cancer cells

In order to further verify if β-elemene can reverse breast cancer cell resistance to chemo-agents, we utilized western blot to detect whether the potential expression changes of PTEN and Pgp. As we know, the expression of the PTEN and PGP has an important role in BCA drug resistance [13, 14].
Fig. 4. The green fluorescent cells expression by dual luciferase activity assay in MCF-7 cells, MCF-7/Doc cells, MCF-7/Adr cells, MCF-7/Doc and MCF-7/Adr cells with β-elemene intervention (50μM/L) for 30h(Fig 4A, 4B). As shown in Fig 4C, 4D, between the untreated group and the Doc and Adr treated group, it shows clearly, after the MCF-7/Doc and MCF-7/Adr cells with β-elemene intervention, coped with the drug, the residual green fluorescent cells was significantly reduced more than MCF-7/Doc and MCF-7/Adr cells. Apoptotic rate of GFP-S was determined after cell mixture was treated with 50nm Doc or 250nm Adr for 30h. **P<0.01, MCF-7/Adr or MCF-7/Doc vs MCF-7/Adr +β-elemene.

Fig. 5. The four miRNAs with consistent expression changes in MCF-7/Adr and MCF-7/Doc cells after β-elemene intervention (50μM/L) for 30h. Compare with MCF-7/Adr and MCF-7/Doc without intervention, miR-29a, miR-222 and miR-452 levels were significantly lower, miR-34a levels were significantly higher, ##**P<0.01, MCF-7/Adr or MCF-7/Doc vs MCF-7/Adr +β-elemene.

As shown in Fig. 6A, the PTEN expression in MCF-7/Doc and MCF-7/Adr cells was significantly decreased, compared with MCF-7 cells. However, the PTEN expressions in...
MCF-7/Doc treated with β-elemene (50μM/L) and MCF-7/Adr cells treated with β-elemene (50μM/L) were significantly increased when compared with the untreated ones. Also in Fig. 6B, the results showed that the Pgp expression in MCF-7/Doc and MCF-7/Adr cells was significantly increased, compared with MCF-7 cells; however, the Pgp expression was significantly decreased in MCF-7/Doc with the intervention of β-elemene and MCF-7/Adr cells with the intervention of β-elemene compared with the untreated MCF-7/Doc and MCF-7/Adr cells.

Discussion

Breast cancer is the most common cancer for women all over the world, Adr and Doc are two chemotherapeutic agents commonly used in the treatment of breast cancer, especially in recurrent or metastatic patients. One of the most important factors for the limited advances applied in cancer treatment is acquired drug resistance. In this study, we proved β-elemene could mediate the MDR specific miRNA, then regulate the corresponding target genes PTEN and Pgp -- reversing the drug-resistant of BCA cells.

Recently, the role of miRNAs in regulating drug resistance is reported. MiRNAs are a class of small non-coding RNAs with 18–25 nucleotides in length, which have been associated with every aspect of tumor biology, including acquisition of resistance to various chemotherapeutic agents. There are several mechanisms have recently been shown to be targeted by miRNAs in drug-resistant breast cancer, including: decreased intracellular drug concentrations; mediated by drug transporters and metabolic enzymes; impaired cellular responses that affect cell cycle arrest, apoptosis; DNA repair and alterations in the availability of drug targets.
We previously showed that β-elemene significantly suppresses breast cancer cells growth and proliferation [15, 16], β-elemene has a wide range of applications in traditional medicine, it has been studied as an agent capable of reversing resistance to chemotherapy [17, 18]. Previous Chinese publication has shown that by up-regulating the expression of c-Cbl and Cbl-b, which leads to inhibition of PI3K/Akt signaling and down-regulation of Pgp expression [19]. β-elemene enhanced the sensitivity of A549/DDP cells to cisplatin and reversed the drug resistance of A549/DDP cells, it enhances susceptibility to cisplatin in resistant ovarian carcinoma cells via downregulation of ERCC-1 and XIAP and inactivation of JNK. [20, 21]. This recent discovery shows that elemene-induced reversal of tamoxifen resistance in MCF-7 cells through oestrogen receptor α (ERα) re-expression and it also shows that estrogen receptors can combine with the primary transcript of miRNAs to modify its biogenesis process [22].

In the present work, the efficacy of β-elemene in reversing the MDR of Doc and Adr cells was evaluated first via the MTT approach. The results demonstrated that β-elemene alone ranging from 10 to 30μM/L did not display a significant anti-proliferative effect on Adr and Doc cells, while it at 50μM/L enhanced the cytotoxicity toward the two cells. After the two cells were exposed to 50μM/L β-elemene for 30h, there was a pronounced increase in the apoptosis rate. Furthermore, it shows time dependence and concentration dependence. The MCF-7/Doc and MCF-7/Adr cells with β-elemene intervention coped with the drug was significantly reduced more than MCF-7/Doc and MCF-7/Adr cells; suggesting that β-elemene can reverse drug resistance by synergistic action.

Secondly, in order to verify another molecular pathway revolved on the impact of β-elemene in anticancer; we proceeded with miRNA expression profiling analysis, which aims to test the specific regulators of β-elemene-mediated anti-cancer properties in MCF-7 cells. From the miRNA expression profiles, we recognized differentially expressed miRNAs and consistent expression changes miRNAs in MCF-7/Adr and MCF-7/Doc cells with β-elemene intervention. This not only shows that resistant breast cancer cells with β-elemene intervention may have characteristics that can significantly change the miRNA expression, but also imply that the cells had mutual pathways along drug resistant specific pathways in selected MCF-7/Adr and MCF-7/Doc cells. In order to verify whether the differential miRNA expression has a major influence in preventing the process of acquiring drug resistance with β-elemene intervention, we conducted related experiments. The results showed that miR-34, miR-222, miR-452 and miR-29a can changed the characteristics of drug resistant BCA cells to Doc and Adr. We therefore conclude that β-elemene could mediate MDR related miRNA expression to reduce the drug resistance of breast cancer. This study could contribute to understanding of the miRNAs roles in reversing drug resistance in BCA.

MDR of tumor cells is often associated with overexpression of Pgp and lower expression PTEN, finally leads to chemotherapeutic failure [13, 14]. To explore a possible role of Pgp and PTEN in the effect of β-elemene on reversing drug resistance, we assessed PTEN and Pgp expression in breast cancer cells treated with β-elemene. Taken together, our results clearly indicate that β-elemene effectively sensitized drug resistant BCA cells to Doc and Adr through a signaling pathway involving regulation of PTEN and Pgp.

In summary, several studies have shown that β-elemene agent enhances sensitivity to chemotherapy in human breast cancer cell lines, it was approved as a national first-class new agent and phase II clinical trials are currently underway. We were able to establish that β-elemene not only causes a strong anticancer effect via activation of the apoptotic pathway to induce BCA cell apoptosis, but also the anticancer effect and reverse the drug resistance of β-elemene have a synergistic effect, β-elemene can effect target gene expression through transcriptional pathway; therefore, we conclude that miRNA plays a major influence in the β-elemene-mediated effect of MCF-7/Adr and MCF-7/Doc cells, particularly, the four specified drug resistant miRNAs. Moreover, this study provides a novel insight of the molecular mechanisms of β-elemene reversing tumor resistance by using the latest technology to detect miRNA communication mechanisms, The strategic in-depth exploration of MDR reducing expression mechanisms could help establish an alternative way of improving chemotherapy
treatments. However, this study still has some shortcomings that need to be discussed. If animal models are embraced into our study — through pharmaceutical interventions on the above subjects — we could further explore how Chinese medicine can inverse and inhibit chemotherapy resistance in BCA from a clinical perspective; which could make our conclusions more persuasive.

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Disclosure Statement

The authors declare no conflicts of interest.

References


