Chimerism and Tolerance Induction in Kidney Transplantation

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Abstract
Chimerism is a state in which bone marrow hematopoietic stem cells from two genetically different animals coexist. To date, the approach has been used successfully to induce the state of immunologic tolerance in the animal models and is now being evaluated in clinical trials of both HLA-identical and HLA-mismatched living-donor kidney transplant recipients in some transplant centers with varying degrees of success. Although the results are promising, the current conditioning regimens are not optimal and longer-term follow up and multicenter studies are needed to ensure the efficacy and safety of the procedures.

Introduction
The evolution of immunosuppressive therapies in the past two decades has led to lower rejection rates and improved recipient short-term allograft outcomes in kidney transplantation, without long-term improvement in allograft-survival. Chronic alloimmune injuries, as well as long-term toxicity of the current immunosuppressive drug regimens are still major concerns. Recent analysis has shown that chronic antibody-mediated rejection is one of the leading causes of late graft failure \cite{1, 2}. Induction of immunologic tolerance remains the ultimate goal to achieve the best long-term graft outcome and avoid drug toxicity. Though many strategies have been successfully used to induce the immunologic tolerance in rodent models, chimerism was the only strategy capable of inducing the immunologic tolerance in large animals and humans \cite{3}. Here we review the current methods that have been used to induce chimerism state and immunologic tolerance in human kidney transplantation.

Chimerism in Human Kidney Transplantation
Chimerism is a state in which bone marrow hematopoietic stem cells from two genetically different animals coexist. To date, there are only a few centers that have experience in inducing tolerance by using chimerism approach; Stanford University, Massachusetts General Hospital (MGH) and our group at Northwestern University. Each of these centers used their own unique conditioning regimens to induce donor hematopoietic cell engraftment and used different post-conditioning protocols \cite{4–6} (table 1).

The Stanford group used a conditioning regimen that included total lymphoid irradiation (TLI) and rabbit antithymocyte globulin in the recipients of HLA-matched kidney transplant. This regimen has been shown to be safe from graft versus host disease (GVHD) in the preclinical...
models and in humans [7, 8]. From 12 patients reported, 4 patients developed stable mixed chimerism for a follow-up period of more than 18 months, and 4 patients had transient chimerism that declined over 12 months post-transplant [4]. All 8 patients were successfully withdrawn from immunosuppressive drugs. However, 3 patients in the study rapidly lost the chimerism after induction and developed acute cellular rejection (ACR). All patients, including the ones with rejection, had good graft function up to 72 months post-transplant. These results demonstrated that even with successful induction of chimerism, there remains the possibility of losing chimerism and subsequent rejection during withdrawal of immunosuppressive drugs. Furthermore, the requirement of chimerism for successful immunosuppressive drug withdrawal was not proven [4], and TLI alone might be able to induce the immunologic tolerance without the need for donor hematopoietic stem cells [9]. Although there were no major complications, episodes of neutropenic fever, pyelonephritis and viral infections were reported (1 CMV, 1 EBV and 3 varicella zoster). Since the long-term graft survival and overall outcomes of HLA-identical living donor kidney transplants are remarkably good [10], the justification for induction of tolerance in these patients is questionable. Nine patients (90%) experienced acute kidney injury as a part of the ‘engraftment syndrome’ [13], the cytokine syndrome-like manifestations associated with the loss of chimerism and the return of recipient hematopoietic cells. Although 7 patients recovered either spontaneously or with treatment, 2 allografts failed to recover (1 associated with ABMR and 1 progressed to thrombotic microangiopathy associated with high tacrolimus levels) [5]. In all 10 patients, transient mixed chimerism was achieved up to 21 days post-transplant with donor marrow infusion. Despite the 2 graft failure observed in the early phase, the remaining patients were successfully withdrawn from maintenance immunosuppression. One patient then developed ACR 3 months following immunosuppression withdrawal, leading to allograft loss. Although the other patients continued to do well in the early years without immunosuppression, chronic active ABMR was detected in the 5-year protocol biopsy of one

Table 1. Comparison of different conditioning regimen in different transplant centers

<table>
<thead>
<tr>
<th>Centers</th>
<th>Current conditioning regimen</th>
<th>Donor cells</th>
<th>Post-transplant</th>
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<tbody>
<tr>
<td>Stanford group [4]</td>
<td>Total lymphoid irradiation, rabbit antithymocyte globulin, intravenous methylprednisolone, prednisone</td>
<td>Peripheral blood CD34+ hematopoietic progenitor cells</td>
<td>Mycophenolate mofetil for 1 month, cyclosporine tapered at 3 months, discontinued at 6 months</td>
</tr>
<tr>
<td>MGH group [5]</td>
<td>NKD03 protocol: cyclophosphamide, humanized antiCD2 mAB, thymic radiation, intravenous cyclosporine A or tacrolimus Modified NKD03/ITN036 protocol: cyclophosphamide, humanized antiCD2 mAB, thymic radiation, intravenous cyclosporine A or tacrolimus, rituximab, prednisone</td>
<td>Unprocessed donor bone marrow cells</td>
<td>Tacrolimus slowly tapered and discontinued at 8 months</td>
</tr>
<tr>
<td>Northwestern group [6]</td>
<td>Cyclophosphamide, total body irradiation, fludarabine</td>
<td>A bioengineered FDA-regulated hematopoietic stem cells enriched for facilitating cells</td>
<td>Mycophenolate mofetil discontinued at 6 months, tacrolimus tapered at 9 months and discontinued at 1 year if durable chimerism is present</td>
</tr>
</tbody>
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logic tolerance can be achieved by inducing transient mixed chimerism using the current MGH protocol. However, long-term alloimmune injury is present in many of the patients and the conditioning regimen needs adjustments to minimize the engraftment syndrome and avoid chronic antibody-mediated injury. Furthermore, more patients with longer follow up are needed to assess the efficiency and safety of the protocol.

**Transient vs. Durable Chimerism**

Our group at Northwestern Memorial Hospital (NMH) has developed bioengineered hematopoietic stem cells enriched for facilitating cells (FCRx). These bone marrow-derived facilitating cells are predominantly composed of a plasmacytoid precursor dendritic cell subpopulation and CD8+ T cells that do not express T cell receptors, which potently enhance engraftment of allogeneic hematopoietic stem cells [18] and limited numbers of syngeneic hematopoietic stem cells in conditioned recipients [19]. Facilitating cells induce the generation of antigen-specific, regulatory T cells in vitro and in vivo and potently prevent GVHD in the mice [20–22]. The conditioning regimen at NMH consisted of pre-transplant fludarabine, pre- and post-transplant cyclophosphamide, and 200 cGy total body irradiation. Eight of 15 HLA mismatched patients who underwent this protocol developed high-level (>90%) durable donor chimerism beginning 1 month after transplantation [23]. One patient developed atypical viral infection and bone marrow failure at 3 months post-transplant, which was rectified by infusion of autologous stem cells. However, he later developed sepsis and allograft loss and was subsequently re-transplanted with a living donor kidney. Six of 8 patients who had full chimerism were successfully withdrawn from immunosuppression and the other 2 patients were successfully weaned to tacrolimus monotherapy. Durable mixed chimerism was noted in two patients. Three patients in the protocol had only transient donor chimerism lasting several months (2 had reduced doses of FCRx and 1 was highly sensitized with high PRA at the time of transplantation) but in vitro proliferative assay by MLR and CML also demonstrated DSH despite the loss of chimerism [23]. One of them was subsequently withdrawn from MMF and later Tacrolimus based on the in vitro testing, but later developed subclinical Banff 1A rejection in the 12-month protocol biopsy, which was treated successfully with intravenous corticosteroids and by increasing tacrolimus dosage to therapeutic range. A second patient with transient mixed chimerism also demonstrated similar subclinical rejection in his 6-month pro-

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protocol biopsy despite stable renal function and DSH by in vitro testing. Complete failure of donor engraftment occurred in one subject who was highly sensitized at the time of transplantation, and developed tacrolimus-induced hemolytic uremic syndrome following transplantation. No patients in the protocol experienced GVHD or engraftment syndrome. Contrary to the tolerance observed in the MGH group after transient mixed chimerism, subclinical rejection was detected early in our two subjects with transient mixed chimerism upon tapering of immunosuppression despite the presence of DSH demonstrated by in vitro assays. In contrast, the patients who achieved durable chimerism were completely free of cellular infiltrates or manifestations of chronic rejection. Notably, a study from the MGH group in which there was solely transient mixed chimerism for a period less than 21 days, reported DSH by in vitro testing in their nonchimeric subjects, some of whom went on to experience rejection episodes [24]. This evidence suggests that, in the absence of durable chimerism, the in vitro MLR and CML assay, which mainly tested the direct donor alloreactivity, is not always predictive of successful immunosuppression without rejection. Based on these findings, nonchimeric recipients are maintained on low-dose immunosuppression rather than complete withdrawal. The patients need close monitoring for the development of chronic GVHD as well as the status of their chimerism. In conclusion, transient or durable chimerism was achieved in the majority of subjects treated with the target dose of FCRx and conditioning without GVHD or engraftment syndrome. Only durable chimerism allowed safe withdrawal of maintenance immunosuppression without the risk of rejection.

Conclusions

Immunologic tolerance to allogeneic kidney transplant in humans is possible by using a chimerism approach. Significant progress has been made in the past decade; however, important issues still need to be investigated:

1. The current conditioning regimen still needs to be modified to achieve a higher rate of successful chimerism induction and reduce the risk of toxicity that potentially outweighs the therapeutic benefits. This is valid for protocols used in HLA-identical as well as in HLA mismatch kidney transplant recipients.

2. Longer-term follow up and a larger sample of patients are needed to ensure the efficiency and safety of these approaches compared to the conventional treatment.

3. If these protocols will be proven to be safe and effective in the long term, similar approaches to induce tolerance should be investigated for recipients of cadaveric renal transplants.

References


