Monkeys and Rats Are Not Susceptible to Ferret Hepatitis E Virus Infection

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Key Words
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Abstract
Ferret hepatitis E virus (HEV), a novel hepatitis E-like virus, has been identified in ferrets in the Netherlands, Japan, and the US. To determine whether ferret HEV transmits to other animals, we inoculated laboratory rats (Wistar), nude rats (Long-Evans-rnu/rnu), and cynomolgus monkeys with ferret HEV (F4351) by intravenous injection. None of the animals demonstrated a positive sign for virus replication, indicating that rats and monkeys are not susceptible to ferret HEV.

Hepatitis E virus (HEV), a small round nonenveloped single-strand-positive RNA virus, is the sole member of the genus Hepevirus in the family Hepeviridae [1]. Hepatitis E virus includes four genotypes of HEV that have been isolated from humans (G1, G2, G3, and G4 HEV) and cause acute hepatitis E [2]. Swine, wild boars, deer, and mongooses are the reservoirs for G3 and G4 HEV, and these HEVs can be transmitted to humans, suggesting that hepatitis E is a zoonotic disease [3]. In addition to G1–G4 HEV, numerous HEVs or HEV-like viruses have been isolated from rabbits, bats, birds, wild rats, red foxes, minks, moose, and ferrets [4–10], and Hepevirus may include at least four species of HEV: G1–G4/rabbit/unclassified wild boar HEV, avian HEV, bat HEV, and rat/ferret HEV [11].

The genome structure of the ferret HEV is similar to that of the other HEVs, containing three open reading frames (ORFs): ORF1 encodes a nonstructural protein, ORF2 encodes a capsid protein, and ORF3 encodes a phosphoprotein with multifunction. The serotype of ferret HEV is different from that of G1–G4 HEV [12]. After ferret HEV was first identified in ferrets in the Netherlands, it was also detected in laboratory ferrets in the US and in imported ferrets in Japan [12]. Expression of ferret HEV ORF2 allowed the partial capsid protein to assemble into virus-like particles, and an enzyme-linked immunosorbent assay (ELISA) for the detection of anti-ferret HEV IgG and IgM antibodies using the virus-like particles as the antigen has been established [12]. However, the pathogenicity and epidemiology of ferret HEV are largely unknown.

Because ferrets have been used not only as a small animal model for virus infections but also kept as pets, studies to determine whether ferret HEV transmits to humans...
or other animals are urgently needed. In the present study, we inoculated SPF rats, nude rats, and cynomolgus monkeys with ferret HEV, and we monitored the virus replication to determine the susceptibility of rats and monkeys to ferret HEV infection.

We used the ferret HEV strain F4351 (GenBank accession No. AB890001) derived from stool specimens from a laboratory ferret (Macaca fascicularis furo) for the infection experiments [13]. The stool specimen containing F4351 was diluted with 10 mM phosphate-buffered saline to prepare a 10% suspension, shaken at 4°C for 1 h, and clarified by centrifugation at 10,000 g for 30 min. The supernatant was passed through a 0.45-μm membrane filter (Millipore, Bedford, Mass., USA) and stored at −80°C until use. The RNA copy number of F4351 was detected as 2 × 10⁵ copies/ml by real-time reverse transcription-polymerase chain reaction (RT-PCR).

Three 15-week-old SPF rats (Wistar, Japan SLC, Hamamatsu, Japan), three 5-week-old nude rats (Long-Evans-rnu/rnu, Japan SLC), three 11-year-old cynomolgus monkeys, three 22-week-old ferrets, and one 2-year-old ferret (Japan SLC) were used. All of the animals were shown to be negative for ferret and rat HEV RNA and anti-G1, G3, G4, ferret, and rat HEV antibodies by a nested broad-spectrum RT-PCR and ELISA, respectively.

To examine the susceptibility of the laboratory animals to ferret HEV, the SPF and nude rats were intravenously inoculated with 0.5 ml of F4351 through the tail vein, whereas the monkeys were intravenously inoculated with 1.0 ml of F4351 through a femoral vein. Three ferrets (FS-1, FS-2, and FS-3) were orally inoculated with 2.0 ml of F4351 mixed with 10 ml of milk for 3 consecutive days, and one ferret (2-year-old, FS-4) was inoculated with 1.0 ml of F4351 through the cranial vena cava.

The serum samples were collected weekly to examine ferret HEV RNA as well as ferret HEV-specific IgG and IgM antibodies. Sera were also used to determine the animal’s alanine aminotransferase (ALT) levels. Stool samples were collected every 3 days after inoculation to detect HEV RNA. The animals were monitored for 3 months after inoculation, with the exception of ferret FS-4, which was monitored until day 63 after inoculation. All of the animal experiments were reviewed and carried out according to the ‘Guidelines for Animal Experiments Per...
formed at the National Institute of Infectious Diseases' under codes 513005 and 614006. The animals were individually housed in BSL-2 facilities.

The extraction of RNA was carried out using the MagNA Pure LC Total Nucleic Acid Isolation Kit (Roche Applied Science, Mannheim, Germany) according to the manufacturer’s recommendations. Reverse transcription was performed with a high-capacity cDNA reverse transcription kit (ABI Applied Biosystems, Carlsbad, Calif., USA). A nested broad-spectrum RT-PCR targeting a portion of the ORF1 genome was performed as described with a slight modification [14]. The nested PCR was carried out with a forward primer, HEV-cs, and an internal reverse primer, HEV-casn. The ferret HEV RNA titer was determined by one-step quantitative RT-PCR with forward (5'-TGAGTGGTATTTATGCGGA-3', nt 4006–4026) and reverse (5’-CAAACACAAAATCTTTCTC- AAAGC-3’, nt 4082–4109) primers, and a probe (5’-6FAM-GCATGCAGGTGTAGGCGC-CAAACTCAGAAAAATCATTCT- TAMRA-3’, nt 4034–4059). Anti-rat, monkey, and ferret HEV IgG and IgM antibodies were detected by ELISA as described [12, 15]. Horseradish peroxidase (HRP)-conjugated goat anti-rat IgG (H+L; Zymed Laboratories, San Francisco, Calif., USA), IgM (Jackson Immunoresearch Laboratories, West Grove, Pa., USA), HRP-conjugated goat anti-human IgG (Cappel, West Chester, Pa., USA), IgM (Cappel), HRP-conjugated goat anti-ferret IgG (H+L; KPL, Gaithersburg, Md., USA), and IgM (Rockland Antibodies and Assays, Gilbertsville, Pa., USA) were used to detect rat, monkey, and ferret antibodies, respectively. Serum ALT levels were monitored weekly using the Fuji Dri-Chem Slide GPT/ALT-PIII kit (Fujifilm, Saitama, Japan).

To examine the infectivity of ferret HEV F4351, we orally inoculated three ferrets with 2 ml of 10% stool suspension. As shown in figure 1, ferret HEV RNA was detected in stools of all three ferrets from 19 to 26 days after inoculation. However, the RNA titer was measurable only in ferret FS-2 at 18, 21, and 24 days after inoculation. The highest ALT level during the whole period is shown. a The highest ALT level during the whole period of the experiment is shown.

All of the SPF rats, nude rats, and monkey serum samples collected from 1 to 13 weeks after inoculation were negative for ferret HEV RNA, or IgG and IgM. The positive period is shown in fig. 1. a The positive period is shown in fig. 1. The highest ALT level during the whole period of the experiment is shown.

<table>
<thead>
<tr>
<th>Animals</th>
<th>Sex</th>
<th>Virus inoculum (in genome equivalent copies)</th>
<th>Ferret HEV RNA</th>
<th>IgG and IgM (IU/L)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Rat-1 (Wistar) M</td>
<td>10^5</td>
<td>Neg</td>
<td>Neg</td>
<td></td>
</tr>
<tr>
<td>Rat-2 (Wistar) M</td>
<td>10^5</td>
<td>Neg</td>
<td>Neg</td>
<td></td>
</tr>
<tr>
<td>Rat-3 (Wistar) M</td>
<td>10^5</td>
<td>Neg</td>
<td>Neg</td>
<td></td>
</tr>
<tr>
<td>Long-Evans-rnu/rnu-1 F</td>
<td>10^5</td>
<td>Neg</td>
<td>Neg</td>
<td></td>
</tr>
<tr>
<td>Long-Evans-rnu/rnu-2 F</td>
<td>10^5</td>
<td>Neg</td>
<td>Neg</td>
<td></td>
</tr>
<tr>
<td>Long-Evans-rnu/rnu-3 F</td>
<td>10^5</td>
<td>Neg</td>
<td>Neg</td>
<td></td>
</tr>
<tr>
<td>Monkey 4824 F</td>
<td>2 x 10^5</td>
<td>Neg</td>
<td>Neg</td>
<td></td>
</tr>
<tr>
<td>Monkey 4826 M</td>
<td>2 x 10^5</td>
<td>Neg</td>
<td>Neg</td>
<td></td>
</tr>
<tr>
<td>Monkey 4830 M</td>
<td>2 x 10^5</td>
<td>Neg</td>
<td>Neg</td>
<td></td>
</tr>
<tr>
<td>Ferret-FS4 M</td>
<td>2 x 10^5</td>
<td>Pos</td>
<td>Pos</td>
<td></td>
</tr>
</tbody>
</table>

Neg = Negative: during the whole period of the experiment the sera were negative for ferret HEV RNA, or IgG and IgM; Pos = positive for ferret HEV RNA, or IgG and IgM. The positive period is shown in fig. 1. a The highest ALT level during the whole period of the experiment is shown.

ferret HEV F4351 is infectious and that ferret HEV was transmitted through the fecal-oral route. When ferret FS-4 was inoculated with F4351 through the cranial vena cava, ferret HEV RNA was detected in stools from 21 to 27 days after inoculation, and anti-ferret HEV IgG and IgM antibodies were detected in the sera at 3 weeks and reached their peak at 6 weeks after inoculation. A significant elevation of the ALT level was not observed. This result indicates that ferret HEV is also transmitted through intravenous inoculation.

Two new proposals for an HEV taxonomy have been published. One proposal suggests that rat and ferret HEVs belong to the same species Orthohepevirus [16], and the other suggests that these two HEVs are different species in the genus Rochehepevirus [17]. However, the nucleotide sequence analyses indicated that the ferret HEV genome...
shared 60.8–68.8% identity with rat HEVs, which is higher than those with G1–G4, rabbit, bird, and bat HEVs (46.6–55.2%), and this is the reason why we selected rats to examine whether ferret HEV was transmissible to other animals. However, our findings even with nude rats did not support any evidence of ferret HEV replication. Here we selected a nonhuman primate (cynomolgus monkey) and inoculated three monkeys with ferret HEV; we found no evidence that cynomolgus monkeys were susceptible to ferret HEV. Our results demonstrated that rats and cynomolgus monkeys are not susceptible to ferret HEV. However, it is necessary to clarify whether ferret HEV is transmissible to humans and causes a zoonotic disease, especially in the ferret feeders.

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References