Update on the Autoimmune Pathology of Multiple Sclerosis: B-Cells as Disease-Drivers and Therapeutic Targets

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Abstract

Background: Collectively, research on the role of B-cells in the pathogenesis of multiple sclerosis (MS) illustrates how translational medicine has given rise to promising therapeutic approaches for one of the most debilitating chronic neurological diseases in young adults. First described in 1935, the experimental autoimmune/allergic encephalomyelitis model is a key animal model that has provided the foundation for important developments in targeted therapeutics.

Summary: While additional B-cell therapies for MS are presently being developed by the pharmaceutical industry, much remains to be understood about the role played by B-cells in MS. The goal of this review is to summarize how B-cells may contribute to MS pathogenesis and thereby provide a basis for understanding why B-cell depletion is so effective in the treatment of this disease.

Key Messages: B-cells are key players in the pathogenesis of MS, and their depletion via B-cell–targeted therapy ameliorates disease activity.

Clinical Implications: In 2008, data from the first CD20-targeting B-cell depleting therapeutic trials using rituximab in MS were published. Since then, there has been a large body of evidence demonstrating the effectiveness of B-cell depletion mediated via anti-CD20 antibodies. Intense research efforts focusing on the immunopathological relevance of B-cells has gained significant momentum and given rise to a constellation of promising therapeutic agents for this complex B-cell–driven disease, including novel anti-CD20 antibodies, as well as agents targeting CD19 and BAFF-R.

Multiple Sclerosis – A Brief Overview

Multiple sclerosis (MS) is the most common chronic neurological disease in young adults, affecting about 2.5 million people worldwide. In countries populated by Northern Europeans and their descendants, the incidence is about 7/100,000, and the prevalence is about 120/100,000 [1]. The incidence of MS seems to have increased over the last century, particularly in women, leading to a female:malesex ratio of 3:1 [2]. The peak age of onset is between 20 and 40. At disease onset, ~80% of patients are diagnosed with relapsing-remitting MS (RRMS); over time, about 60% of RRMS patients will develop secondary progressive MS. About 25% never experience sustained neurological disability, whereas a smaller percentage become severely disabled a short time after disease diagnosis. Pathologically, MS is characterized by chronic CNS inflammation accompanied by demyelin-
ation, gliosis, and axonal loss. Axonal degeneration is believed to be ultimately responsible for progressive neurological dysfunction. The most widely accepted view of MS pathogenesis includes autoimmune-mediated myelin injury in a susceptible host. MS behaves as a complex genetic trait [3], and exposure to infectious, climatic and other environmental variables likely have a considerable effect on an individual’s risk for developing MS. Disease-specific, immune-modulatory therapies became available in the mid-to-late 1990s; currently, seven substances are approved for the treatment of MS (interferon-ß1, glatiramer acetate, mitoxantrone, natalizumab, fingolimod, dimethyl fumarate, teriflunomide). These compounds have been extensively studied and are discussed elsewhere in the literature. In this review article, we will focus on B-cells, their immunological properties relevant to MS, and how these functions are targeted by the B-cell depleting therapeutic strategies currently in development.

### B-Cells: Drivers of MS

B-cells can exert effector functions as antigen-presenting cells, by cytokine and antibody production, and they participate in the formation of ectopic lymphoid tissues (fig. 1). The strongest evidence to date for B-cells playing a crucial role in MS immune pathology stems from studies evaluating the effect and efficacy of anti-CD20 B-cell-depleting therapies such as rituximab, ocrelizumab, and ofatumumab [4–7]. Interestingly, the initial impetus for B-cell depleting therapy was to remove autoantibody-producing plasma cells after multiple experimental autoimmune encephalitis (EAE) studies had demonstrated the critical roles of antibody responses in the development of CNS demyelination [8–11]. However, since the late 1990s, it has become increasingly appreciated that antigen presentation by B-cells is a necessary step for triggering autoimmunity against the CNS myelin oligodendrocyte glycoprotein [12–14]. B-cells can provide activation/effector mechanisms, and can assume proinflammatory, anti-inflammatory and/or regulatory roles. To date, the exact target antigens of pathogenic B-cell responses in MS remain unknown, despite our knowledge that disease-associated B-cells result from antigen-driven affinity maturation. Needless to say, not all B-cells in MS patients support detrimental autoimmunity. Therefore, being able to clearly differentiate pathologically relevant from normal B-cells in the future will pave the way for treatments with enhanced therapeutic precision and improved safety profiles, thereby bringing us closer to personalized therapy.

In the following paragraphs, we discuss the B-cell functions that have been demonstrated to be involved in the immune pathogenesis of MS, as well as additional functions that may likely be involved in MS immune pathology. We will focus mainly on data from human studies but will include experimental animal data where appropriate.

### The Peripheral B-Cell Compartment in MS

There is ample evidence that peripheral B-cell responses are closely involved in the immune pathology of MS through proinflammatory mechanisms, bystander activation, or through regulatory functions. Under normal circumstances, B-cell tolerance is necessary to control autoimmunity that can randomly develop during...
early B-cell development [15] or even during targeted immunity against foreign pathogens [16]. Central B-cell tolerance, which reduces the autoimmune potential of B-cells developing in the bone marrow, appears to be intact in the majority of MS patients; conversely, B-cell tolerance mechanisms that regulate the autoimmunity of B-cells circulating between peripheral lymphoid tissues seems to be defective [17]. Cervical lymph nodes have been described as possible sites supporting CNS-directed autoimmunity in both humans [18] and mice [19].

Memory B-cells serve as highly efficient antigen-specific antigen-presenting cells (APC) [20, 21]. In this regard, myelin-reactive memory B-cells can be found in the peripheral blood of MS patients [22]. Memory B-cells express high levels of CD20; they are effectively depleted and repopulate slowly following treatment with anti-CD20 targeting monoclonal antibodies [23], coinciding with sustained suppression of MS disease activity [24]. In MS, B-cells were shown to secrete increased levels of IL-6 compared to healthy controls; this proinflammatory bias was not observed in returning B-cells 12 months after B-cell-depleting therapy [25]. Given the repopulating B-cell compartment is mainly composed of mature naïve and immature B-cells [26], it is likely that the increased IL-6 production in MS patients is a function associated with antigen-experienced memory B-cells. Memory B-cells are also effective supporters of T-cell immune mechanisms; their depletion reduces IL-17 production by peripheral blood lymphocytes, providing further evidence for the role of B-cells in supporting proinflammatory T-helper 17 cell (Th17) responses [13, 25]. Furthermore, B-cells from MS patients were shown to respond to non-specific activating stimuli such as CpG or IFN-γ with an exaggerated proinflammatory cytokine profile [27]. Accordingly, the proinflammatory functions of B-cells in MS can also occur in an antigen-independent fashion, by way of bystander activation of T-cells, a mechanism that explains the association of MS relapses with systemic infections [28, 29].

In humans and mice, IL-10 secreting B-cells (B10 cells) exert regulatory roles by suppressing T-helper 1 (Th1) differentiation [30, 31] and by downregulating TNF-α production in monocytes [32]. Conflicting data exists regarding the B-cell subpopulation responsible for IL-10 secretion. One study found IL-10 expressing B-cells among activated memory B-cells, suggesting that B10 cells perform antigen-specific regulatory functions [32]; increased numbers of IL-10 producing B-cells were reported in patients with autoimmune disease although most of the MS patients in this study had received treatment with immunosuppressive or immune-modulatory therapies [33]. Conversely, another study reported that IL-10 secretion is mainly a function of naïve B-cells [21] and that a switch from regulatory B10 cells to proinflammatory B-cells may occur as B-cells transition from the naïve to the memory phenotypes [21]. In EAE, B10 cells inhibit autoimmune T-cell responses, an effector function that is dependent on IL-21 and CD40-mediated interaction with T-cells [33]. Interestingly, under certain circumstances, therapeutic B-cell depletion in EAE may also eliminate regulatory B-cells and result in exacerbation of disease activity [14]. To date, disease worsening as a direct response to CD20-targeting B-cell–depleting therapy has not been observed in humans; however, an increased proinflammatory monocytic phenotype has been described in some MS patients after treatment with rituximab [34].

We recently demonstrated that in MS, clonally related B-cells provide an antigen-specific and immunologically active link between the periphery and cerebrospinal fluid [35, 36] and that oligoclonal bands (OCB)-producing B-cells are present not only in the CNS but also in peripheral blood [37]. Therefore, antigen-stimulated B-cells provide an active immune axis bridging the CNS and the periphery and they may undergo immune stimulation in both compartments [35–37], further supporting the important pathological role of peripheral B-cell immunity in MS (fig. 2).

B-Cells in the CNS

A growing body of evidence has not only established the CNS as the target tissue of autoimmunity in MS but has identified sub-compartments (i.e., brain parenchyma, cerebrospinal fluid, meningeal tissue) that reflect immunological activity, supporting B-cell affinity-maturation, proliferation, and terminal differentiation to antibody-producing plasma cells. B-cells are commonly found in MS lesions, albeit predominantly in active lesions and at significantly lower numbers compared to T-cells [38]. Lymphoid B-cell follicle-like structures featuring characteristics of germinal centers have been observed in the cerebral meninges of secondary progressive MS patients [39] and are associated with cortical neuronal loss and demyelination [40, 41]. While the pathological importance of such ectopic B-cell follicles as drivers of CNS-targeted autoimmunity remains to be fully understood, the presence of CD35+ follicular dendritic cells and proliferating B-cells, together with expression of the B-cell-attracting chemokine CXCL13 and of the B-cell-activation factor (BAFF) suggest that active immune responses occur in meningeal tertiary lymphoid tissues in second-
ary progressive MS patients [39, 42]. Furthermore, CXCL13 and BAFF have also been described in MS lesions [43, 44] where these factors could mediate local B-cell recruitment and maturation at sites of active demyelination. B-cells present in MS CNS and CSF are clonally expanded [45–47] and IgG class-switched [48, 49], and their immunoglobulin genes are somatically hypermutated and appear to be subject to intrathecal affinity-maturation [36, 47, 50–52], which is an additional evidence to support the involvement of active B-cell immune mechanisms. The fact that overlapping B-cell repertoires expressing related Ig-VH sequences were found in MS brain parenchyma, meningeal lymphoid follicles, CSF, and the periphery [35, 36, 53, 54], suggests an intrathecal immunological continuum that may be exposed to immune-stimulation on both sides of the blood-brain barrier (fig. 2).

Memory B-cells in the CSF display upregulation of costimulatory molecules [55] suggesting active B- and T-cell interactions. Different stages of B-cell development are present in CSF [55] and CSF plasma cells are producers of soluble clonal IgG [56, 57], further supporting antigen-driven B-cell immune responses to be active intrathecally. It has been repeatedly shown that antigen-experienced B-cell subsets predominate in the CSF and CNS. Accordingly, >90% of B-cells in the CSF carry the memory B-cell marker CD27 and a fraction of CSF B-cells express CD138 and/or CD38, supporting the presence of mechanisms that stimulate the maturation of clonally activated memory B-cells into antibody-producing plasmablasts and plasma cells [58]; the levels of CD27-IgD+ naïve B-cells are significantly lower in the CSF compared to blood [59].

To date, the pathological relevance of antibodies in MS remains unclear despite the intrathecal presence of clonal IgG (OCB) and IgM [60–62] and significant IgG deposits in some demyelinating MS lesions [63]. The rapid response to B-cell depleting therapy, leaving antibody levels nearly unchanged [5, 6], has led to the speculation that antibodies play a less important pathogenic role. However, CNS-directed autoantibodies require a permissible inflammatory environment [8] or at the very least a functional complement system to exert their pathogenic func-
tion [64]. Eliminating the T-cell activating, antigen-presenting functions of B-cells by way of B-cell depletion, likely reduces intraparenchymal inflammatory effectors to a degree that will render autoantibodies ineffective promoters of tissue damage. Overall, memory B-cells and plasmablastsplasma cells are the most abundant B-cell subsets in the CNS and CSF of MS patients. However, these B-cells do not represent a static immune response but rather engage in active affinity maturation with help from other immune cell types, cytokines, and survival factors.

**B-Cell–Depleting Therapy in MS**

CD20-targeting lymphocyte-depleting therapy was shown to effectively suppress MS disease activity measures, including the development of new enhancing lesions and relapse rates [4–7]. Beginning with the first studies that described the successful treatment of MS with the anti-CD20 antibody rituximab [4, 5], further efforts were made to explore B-cell depletion as therapeutic paradigm in MS. In the following paragraphs, we briefly discuss emerging therapies that were developed to directly target B-cells: anti-CD19, anti-CD20, and anti-BAFF-R (table 1). All are IgG1 antibodies and can mediate complement-dependent (CDC) and antibody-dependent cell-mediated (ADCC) cytotoxic effects on their target cells.

**Anti-CD20 Therapy**

Three monoclonal anti-CD20 antibodies have been or are currently being studied for the treatment of MS: rituximab (chimeric human/mouse IgG1), ocrelizumab (humanized IgG1), and ofatumumab (fully human IgG1); they differ in their recognition of CD20 epitopes and in the intensity of CDC or ADCC elicited, but all mediate near-complete depletion of CD20+ B-cells in peripheral blood (reviewed in [65]). Very low numbers of B-cells remain in the circulation following CD20-targeted depletion [66], and certain B-cell populations resident in lymphoid tissues may also display resistance to B-cell depleting therapies [67]. CD20 is expressed on a wide range of B-cell subsets starting at the pre-B-cell stage and extending through to memory B-cells (fig. 3). Accordingly, pro-B-cells and antibody-producing plasmablasts/plasma cells are not primarily affected by anti-CD20 therapy; levels of soluble immunoglobulins in serum [5, 6] remain mostly unchanged, at least in the short-term. Rituximab was shown to reduce CSF B-cell counts, but at 6 months the OCB and CSF IgG-Index remained unchanged [68].

*Table 1. Biologics targeting B-cells or B-cell-activating factors*

<table>
<thead>
<tr>
<th>Biologic</th>
<th>Molecular characteristics</th>
<th>Targets</th>
<th>Effects in MS</th>
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<tr>
<td>Rituximab</td>
<td>murine/human chimeric monoclonal IgG1</td>
<td>CD20 B cells: see fig. 3 CD20+ T cells</td>
<td>reduced MRI measures of disease activity [4, 5] reduced relapse rate [4, 5] reduced intrathecal B-cells</td>
</tr>
<tr>
<td>Ocrelizumab</td>
<td>humanized (90%) monoclonal IgG1</td>
<td>CD20 B cells: see fig. 3 Effect on CD20+ T cells not yet known</td>
<td>reduced MRI measures of disease activity [6] reduced relapse rate [6]</td>
</tr>
<tr>
<td>Ofatumumab</td>
<td>fully human monoclonal IgG1</td>
<td>CD20 B cells: see fig. 3 Effect on CD20+ T cells not yet known</td>
<td>reduced MRI measures of disease activity [7]</td>
</tr>
<tr>
<td>MEDI-551</td>
<td>humanized monoclonal IgG1</td>
<td>CD19 B cells: see fig. 3</td>
<td>unknown</td>
</tr>
<tr>
<td>VAY736</td>
<td>fully human monoclonal IgG1</td>
<td>BAFF-R B cells: see fig. 3 BAFF-R+ T cells?</td>
<td>unknown</td>
</tr>
<tr>
<td>Atacicept</td>
<td>recombinant fusion protein with extracellular domain of TACI receptor and Fc domain of human Ig</td>
<td>BAFF APRIL</td>
<td>unexpected inflammatory effects [81]</td>
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Anti-CD20-mediated B-cell depletion using rituximab represents the first effort at directly targeting B-cells in MS [4, 5]. In the 48-week phase II trial [5], patients who received rituximab had a highly significant >90% reduction in total gadolinium-enhancing lesions on brain MRI over the course of the study, beginning at week 12; clinically, a significant relapse-rate reduction of about 50% was observed at 24 and 48 weeks [5]. At 48 weeks, 24.1% of patients tested for the presence of human anti-chimeric antibodies (HACA, i.e., antibodies against rituximab) had developed HACA without apparent association with efficacy measures. The phase II trials with ocrelizumab and ofatumumab cite similar highly statistically significant reduction in the numbers of new and total gadolinium-enhancing lesions [6, 7]; ocrelizumab resulted in relapse rate reductions between 73 and 80% [6]. Ocrelizu- 
mab is currently in phase III clinical development for RRMS and primary progressive MS (ClinicalTrials.gov Identifiers: NCT01247324, NCT01412333, and NCT01194570).

As detailed earlier in this article, multiple functions of B-cells can be affected by anti-CD20-mediated B-cell deple-
tion. The long-term effects of anti-CD20 therapy in MS have yet to be fully understood, both in terms of ef-
icacy and safety. Ocrelizumab was reported to result in sustained suppression of clinical MRI disease-activity 72 weeks after the last of 4 dosages applied at 24-week intervals [24]. Over the short-term, anti-CD20 therapy can result in mostly mildly reduced serum immunoglobulin levels; however, prolonged exposure to anti-CD20 antibodies may induce delayed depletion of the plasma cell population and reduction of soluble immunoglobulins due to reduced memory B-cell formation and terminal differentiation into antibody-producing subsets [69].

Taken together, the clinical evidence demonstrates that CD20-targeting therapy is highly effective in reducing MRI disease-activity outcome measures and phase III clinical trials are expected to fully reveal the therapeutic potential of this approach.

Interestingly, CD20 was also reported to be expressed at low levels in a small subset of T-cells [66, 70] in healthy donors and patients with rheumatoid arthritis (RA); rituximab effectively depletes these T-cells from the peripheral blood of RA patients [71]. CD20-expressing T-cells are likely functionally diverse but harbor proinflammatory Th17 properties in RA [72]. Our own work confirms the depletion of CD3+CD20dim T-cells from the peripheral blood of MS patients treated with rituximab [66]. Clearly, further work is necessary to determine whether CD20-expressing T-cells contribute to MS immune pathology.

**Anti-CD19 Therapy**

More recently, a humanized anti-CD19 IgG1 antibody [73] MEDI-551, entered phase II clinical development for the treatment of MS (ClinicalTrials.gov Identifier: NCT01585766). At this point, no data for MEDI-551 in MS have been reported. CD19 is expressed on B-cells starting with pro-B-cells through the antibody-producing plasmablast stage (fig. 3), and is gradually lost during terminal differentiation to plasma cells [74]. In an animal model with humanized CD19 and CD20, MEDI-551 induced longer-lasting B-cell depletion compared to rituximab due to significant effects on early B-cells (pro-B-
cells) in the bone marrow [73]. Also in animals, it was
shown that anti-CD19 therapy reduces levels of serum immunoglobulins including autoantibodies due to plasmablast/plasma cell depletion [75]. It is unclear whether autoantibodies play a significant role in MS immune pathology and whether the enhanced antibody-reducing functionality of MEDI-551 will provide increased efficacy and/or risk of infection compared to anti-CD20 therapy. Aside from the effect on plasmablasts and plasma cells and slower B-cell repopulation after treatment, MEDI-551 is expected to affect the same B-cell functions as CD20-targeting strategies. Anti-CD19 therapy will not deplete CD3+CD20

dim T-cells; however, a small number of reports describe CD19-expressing dendritic cells [74, 76], but their existence in humans has not been diligently studied and the potential effect of anti-CD19 therapy on dendritic cells is merely speculative at this point.

**Anti-BAFF-R Therapy**

The most recent addition to the armamentarium of biologics directly targeting B-cells is VAY736, a fully human IgG1 antibody against the B-cell activating factor-receptor (BAFF-R), currently in phase II clinical development for MS (ClinicalTrials.gov Identifier: NCT02038049). In healthy humans, BAFF-R is expressed on naïve B-cells and can be found through post-germinal center memory B-cell and plasma blast stages (fig. 3) [77]. Low levels of BAFF-R have also been described in human central and effector T-cells [78]. BAFF-R is the receptor for BAFF (also known as BLyS, B lymphocyte stimulator); BAFF promotes B-cell survival at numerous stages throughout B-cell development. In MS, BAFF was found to be elevated in demyelinating lesions [44], and has also been suggested to be involved in the formation of lymphoid follicle-like meningeal structures [42]. Interestingly, therapeutic targeting and neutralization of soluble BAFF and APRIL (A Proliferation-Inducing Ligand), which is another B-cell stimulator, by atacicept lead to increased inflammatory activity in MS [79]. No data has been reported regarding the biological effects of VAY736 in humans. However, given the fact that its IgG1 isotype can induce CDC and ADCC, and that BAFF-R is expressed on a wide range of B-cell subsets, VAY736 may likely show similar effects on the B-cell compartment as anti-CD20 antibodies but may also lead to reduced serum immunoglobulin levels. Like anti-CD20 antibodies, VAY736 may induce depletion of a small portion of T-cells, which may or may not be of therapeutic relevance. Theoretically, VAY736 could exert additional effects by blocking BAFF binding to BAFF-R on B-cells that escape depletion in peripheral blood and lymphoid tissues, and may therefore interfere with a similar immunological pathway as atacicept, that is, BAFF/BAFF-R interaction. However, atacicept was designed to target and neutralize soluble B-cell activation factors, while VAY736 appears to have been developed to primarily target B-cells for depletion, illustrating a different mode of action. In this context, it is interesting to note that BAFF-R-deficient mice have fewer mature B-cells [80] and develop increased EAE severity [81], a scenario that probably mirrors the effects of atacicept in humans.

**Summary**

B-cells play important roles in the initiation and perpetuation of CNS-targeting inflammation in MS. There are overlapping B-cell repertoires on both sides of the blood-brain barrier, suggesting that disease-driving immunological stimuli are active not only in the CNS but also in the periphery. Multiple B-cell mediated mechanisms are likely involved in MS immune pathology, with antigen-presentation by B-cells occupying a central role. B-cell depletion is a highly effective and promising therapeutic approach for MS. Four different biologic agents that directly target B-cells for depletion are currently in clinical development, two targeting CD20, one targeting CD19, and one targeting the BAFF-R. The common denominator for each of these therapeutic approaches is the depletion of the B-cell compartment (although this remains to be formally demonstrated for the anti-BAFF-R). However, each strategy also has its unique features, which may or may not contribute to differences in therapeutic efficacy and/or safety profiles. Additional work is required to further delineate the features of pathologically relevant B-cells and their target antigens in MS.
References


