Association of MiRNA-146a, MiRNA-499, IRAK1 and PADI4 Polymorphisms with Rheumatoid Arthritis in Egyptian Population

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Key Words
Rheumatoid arthritis • MiRNA-146a • MiRNA-499 • IRAK1 • PADI4

Abstract
Background/Aims: Rheumatoid arthritis (RA) is a systemic autoimmune disease affecting up to 1% of the population worldwide. The aim of the present study was to investigate whether miRNA-146a rs2910164, miRNA-499 rs3746444, IRAK1 rs3027898 and PADI4 rs1748033 polymorphisms are associated with susceptibility to RA in Egyptians and whether they influence disease severity and activity. Methods: The study was performed on 104 unrelated RA patients and 112 healthy subjects. RA patients were further subdivided into active and inactive RA groups. Polymorphisms were genotyped by using real-time polymerase chain reaction with TaqMan allelic discrimination assay. Results: Significant differences in the frequency of miRNA-146a rs2910164, miRNA-499 rs3746444, IRAK1 rs3027898 and PADI4 rs1748033 alleles and genotypes were observed between RA patients and controls. Only CA and AA genotypes of IRAK1 rs3027898 shows a significant difference between active and inactive subgroups. MiRNA-146a rs2910164 and IRAK1 rs3027898 polymorphisms were a risk factor for predisposition to RA in codominant and dominant tested inheritance models, while, the miRNA-499 rs3746444 and PADI4 rs1748033 polymorphisms were a risk factor in codominant and recessive one. CG and GG genotypes of miRNA-146a rs2910164 were associated with positive erosions. CA genotype of IRAK1 rs3027898 was associated with low disease activity and negative erosions, while, the AA genotype was associated with high disease activity. CC genotype of PADI4 rs1748033 was associated with negative rheumatoid factor. Conclusion: The 4 studied SNPs were likely to play an important role in the susceptibility to RA and can influence disease severity and activity in Egyptian population.

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Introduction

Rheumatoid arthritis (RA) is a persistent joint inflammatory disease which can appear in people of any age. The prevalence of RA increases with age affecting approximately 6% of the population over age of 65 [1]. As in other autoimmune diseases (ADs), females are affected more often than males [2]. Both ecological and heredity factors have been involved in RA susceptibility. The principle heredity factor for RA is the HLA-DRB1 gene, however, the HLA genes deem a 33% of the heredity liability to disease [3]. Latterly, many non-HLA genes have been identified to be associated with RA susceptibility [4]. One class of heredity variants that has been the focus of attention recently is the class of DNA polymorphisms that affect microRNA (miRNA) binding to their target messenger RNA (mRNA), leading to translational repression or mRNA degradation [5, 6].

MiRNAs form a class of noncoding RNAs that are assumed to be diagnostic and prognostic biomarkers of several diseases [7]. They are present in many biological fluids, involving saliva, semen, vaginal secretions and menstrual blood [8]. They are contributed in the organization of most biologic processes as chromosome architecture, cell proliferation, apoptosis, stress resistance, and stem cell maintenance [9, 10]. Many studies have announced that polymorphisms in miRNA and its target sites influence the pathogenesis of several human diseases, including RA [11, 12]. MiRNA-146a could control the expression of interleukin (IL)-1 receptor-associated kinase (IRAK1), IRAK2 and targets tumor necrosis factor (TNF) receptor-associated factor 6 (TRAF6), which are regulators of the TNF-α signaling pathway [13]. Increased expression level of miRNA-146a was detected in the synovial fluid and peripheral blood mononuclear cells of RA patients than disease control and healthy subjects. Its expression level is positively correlated with levels of TNF-α and with RA disease activity [5, 14]. On the other hand, miRNA-499 targets IL-17 receptor B (IL-17RB), IL-23a, IL-2Rβ, IL-6, IL-2, IL-18R, IL-21 and peptidyl arginine deiminase type 4 (PADI4). All of these inflammatory cytokines play an important role in the pathogenesis of RA [15-17].

Therefore, miRNA-146a and miRNA-499 and their genes targets represent a new group of molecules, which seem to be very promising in the discovery of new heredity variants that could predispose to RA susceptibility. The aim of the present study is to investigate the role of certain polymorphisms in miRNA-146a rs2910164 gene, and its target IRAK1 rs3027898 gene, as well as in miRNA-499 rs3746444 gene and its target PADI4 rs1748033 gene in RA susceptibility in a sample of Egyptian people and to examine whether these polymorphisms can influence the severity and activity of the disease.

Materials and Methods

Subjects

One hundred and four unrelated RA patients (88 females and 16 males, mean age 39.5± 10.75 years) were enrolled in the study and diagnosed according to the 1987 American College of Rheumatology criteria for RA [18]. They were recruited from cases admitted and followed up in Rheumatology unit, Kasr Al-Ainy Hospital, Faculty of Medicine, Cairo University, during the period from October 2015 to October 2016. Their ages and general characteristics are summarized in Table 1.

All patients underwent complete history taking especially for disease duration and presenting symptoms or joint affection. Complete blood picture (CBC), alanine aminotransferase (ALT) and creatinine were measured to exclude any patients suffering from kidney or liver diseases. Disease activity was assessed by measuring the erythrocyte sedimentation rate (ESR), anti nuclear antibody (ANA) and 28-joint disease activity score (DAS28). RA patients were split into two subgroups as indicated by their DAS28 into 48 (46%) patients with score >3.2 and they were considered as active RA patients, while 56 (54%) with score ≤3.2 were considered inactive RA patients. Disease severity was assessed by number of patients with erosions and seropositivity for rheumatoid factor (RF). 24 (23%) patients have positive erosions, while 76 (73 %) have positive RF. Plain X-ray films (postero-anterior and oblique views) of the hands/wrist of patients were obtained.
In addition, 112 age and sex matched healthy volunteers were enrolled in the study and were considered as normal control group. They were recruited from healthy subjects admitted to the same hospital for routine checkup. They did not suffer hypertension, liver or renal diseases and no known past or family history of RA. Controls were 86 females and 26 males with a mean age of (41±11.13) years. A written informed consent was obtained from all subjects prior to their inclusion in the study. The research protocol was affirmed by the Ethics Committees of Faculty of Pharmacy, Cairo University.

Sample collection and biochemical tests

Venous blood samples (5ml) were collected and CBC as well as ESR were performed. A portion of blood was admitted to clot and centrifuged at 4°C to separate serum used for assessment of ALT, creatinine, ANA and RF. ALT and creatinine assays were performed using standard diagnostic kits supplied by (Quimica Clinica Aplicada, Spain). ANA and RF were assessed by immunofluoresence technique, following the manufacturer’s instructions. A second portion of blood was assembled in vacutainer tubes containing EDTA and was stored at -80°C until DNA extraction.

DNA extraction and genotyping assay:

DNA was extracted from entire blood utilizing QIA-amplification extraction kit (Qiagen, Venlo, Limburg, Netherlands). DNA samples were quantitated utilizing the NanoDrop1-1000 spectrophotometer (NanoDrop innovations, Inc., Wilmington, USA). Genotyping was performed using real-time polymerase chain reaction with TaqMan allelic discrimination assay (Applied Biosystems, USA). Single nucleotide polymorphisms (SNPs) of miRNA-146a rs2910164, IRAK1 rs3027898, miRNA-499 rs3746444 and PADI4 rs1748033 were analyzed in the extracted DNA by using specific primers and Taqman probes (Taqman genotyping assays, Applied Biosystems, USA). The following PCR primers were used for miRNA-146a rs2910164: 5’CATGGCTTGTGTCAGTGACACCT 3’(forward) and 5’TGAATTACGTTCTTCCAAGCTGGAT3’(reverse); for miRNA-499 rs3746444: 5’ATGTTTAACTCCTCTCCAGTGAAAC3’(forward) and 5’TCACAGCAAGCTGTGCTTCCC3’(reverse); for IRAK1 rs3027898: 5’ACAAGACCCCTGACACCTAAGACAC3’(forward) and 5’TGAAGTGTAAGGCATCAGGAAAGCT3’(reverse); for PADI4 rs1748033: 5’ACTTGATGGGATTTCAGAAATCTC3’(forward) and 5’TGAAATTCAGTTCTTCAGCTGGGAT3’(reverse); for miRNA-146a rs2910164. DNA samples were genotyped utilizing the NanoDrop1-1000 spectrophotometer (NanoDrop innovations, Inc., Wilmington, USA). Genotyping was performed using real-time polymerase chain reaction with TaqMan allelic discrimination assay (Applied Biosystems, USA). Single nucleotide polymorphisms (SNPs) of miRNA-146a rs2910164, IRAK1 rs3027898, miRNA-499 rs3746444 and PADI4 rs1748033 were analyzed in the extracted DNA by using specific primers and Taqman probes (Taqman genotyping assays, Applied Biosystems, USA). The following PCR primers were used for miRNA-146a rs2910164: 5’CATGGCTTGTGTCAGTGACACCT 3’(forward) and 5’TGAATTACGTTCTTCCAAGCTGGAT3’(reverse); for miRNA-499 rs3746444: 5’ATGTTTAACTCCTCTCCAGTGAAAC3’(forward) and 5’TCACAGCAAGCTGTGCTTCCC3’(reverse); for IRAK1 rs3027898: 5’ACAAGACCCCTGACACCTAAGACAC3’(forward) and 5’TGAAGTGTAAGGCATCAGGAAAGCT3’(reverse); for PADI4 rs1748033: 5’ACTTGATGGGATTTCAGAAATCTC3’(forward) and 5’TGAAATTCAGTTCTTCAGCTGGGAT3’(reverse).

Probes were synthesized with reporter dye FAM or VIC covalently linked at the 5’ end and a quencher dye MGB linked to the 3’ end of the probe (Applied Biosystems, USA). Extracted genomic DNA was amplified in a 50 μl solution. After a denaturation time of 10 min at 95°C, 45 cycles at 92°C for 15s then 60°C for 90s for annealing and extension were carried out and fluorescence was measured at the end of every cycle and at the endpoint. PCR products were checked by a Rotor gene Q Real Time PCR System (Qiagen, Valencia, CA, and USA) (Fig. 1). Homology searches using BLAST was carried out to confirm the nucleotide sequence [19].

Table 1. General characteristic of RA patients. Quantitative data were presented as M±SD; qualitative data were presented as number and percent n (%).

<table>
<thead>
<tr>
<th>Variable</th>
<th>RA Patients</th>
</tr>
</thead>
<tbody>
<tr>
<td>n</td>
<td>104</td>
</tr>
<tr>
<td>Age, years</td>
<td>39.5±10.75</td>
</tr>
<tr>
<td>Sex</td>
<td></td>
</tr>
<tr>
<td>Male</td>
<td>16 (15.38)</td>
</tr>
<tr>
<td>Female</td>
<td>88 (84.61)</td>
</tr>
<tr>
<td>Disease duration, years</td>
<td>7±5.04</td>
</tr>
<tr>
<td>CBC</td>
<td></td>
</tr>
<tr>
<td>Hb (gm/dl)</td>
<td>11.71±1.54</td>
</tr>
<tr>
<td>TLCx (10^3/cell/liter)</td>
<td>7.67±2.34</td>
</tr>
<tr>
<td>PLC (x10^3/palatlet/liter)</td>
<td>322±200.34</td>
</tr>
<tr>
<td>ALT (U/L)</td>
<td>29.77±22.88</td>
</tr>
<tr>
<td>Cr (mg/dl)</td>
<td>0.93±0.81</td>
</tr>
<tr>
<td>Disease activity</td>
<td></td>
</tr>
<tr>
<td>- ESR (mm/hr)</td>
<td>41.79±23.67</td>
</tr>
<tr>
<td>- ANA</td>
<td></td>
</tr>
<tr>
<td>ANA+</td>
<td>14 (13.5)</td>
</tr>
<tr>
<td>ANA-</td>
<td>84 (80.8)</td>
</tr>
<tr>
<td>-DAS 28</td>
<td></td>
</tr>
<tr>
<td>Low disease activity</td>
<td>56 (54)</td>
</tr>
<tr>
<td>Moderate to high disease activity (&gt;3.2)</td>
<td>48 (46)</td>
</tr>
<tr>
<td>Disease severity</td>
<td></td>
</tr>
<tr>
<td>-Erosions</td>
<td></td>
</tr>
<tr>
<td>Erosions+</td>
<td>24 (23)</td>
</tr>
<tr>
<td>Erosions-</td>
<td>80 (77)</td>
</tr>
<tr>
<td>-RF</td>
<td></td>
</tr>
<tr>
<td>RF+</td>
<td>76 (73)</td>
</tr>
<tr>
<td>RF-</td>
<td>28 (27)</td>
</tr>
</tbody>
</table>

Statistical analysis

Statistical Package for the Social Science (IBM SPSS) version 22 was used for statistical calculations. Results were expressed as mean ± standard deviation (M±SD) or frequencies (number of cases) and percentages when appropriate. Association of different genotypes with Hardy–Weinberg equilibrium (HWE), the genotypes distribution and allele’s frequencies of each SNP in RA patients and controls, or
in active RA and inactive RA patient groups were analyzed using the Chi-square test. Kruskal Wallis test with Mann Whitney as post-hoc pairwise comparisons were used to compare numerical variables between groups. Multivariate logistic regression analysis models, odds ratios (ORs) and 95% confidence intervals (CIs) of crude ORs and of age and sex adjusted ORs were used to test for the preferential effect of each gene mutation on the occurrence of cases. P values less than 0.05 was considered statistically significant.

Results

The observed genotype frequencies for the polymorphism in RA patients and control subjects were in HWE for miRNA-146a (P=0.42 and 0.12, respectively) and miRNA-499 (P=0.25 and 0.8, respectively), while did not conform to HWE for IRAK1 (P=0.04 and 0.001, respectively) and PADI4 (P=0.001 and 0.001, respectively).

The genotype distributions of the four studied SNPs in all subjects are illustrated in Table 2. MiRNA-146a rs2910164 showed a highly statistically significant difference in the G allele frequency (P=0.001) and GG genotype distribution (P=0.03). The G allele and the homozygote genotype GG were more frequent in RA patients than controls (48% vs 32.1%, respectively, for the G allele and 21.2% vs 7.14%, respectively, for the GG genotype). IRAK1 rs3027898 revealed a statistically significant difference in the A allele frequency (P=0.01) and CA genotype distribution (P=0.001). Where, the A allele and the heterozygote genotype CA were more frequent in the RA patients than controls (40.38% vs 29.46%, respectively, for the A allele and 38.46% vs 8.9%, respectively, for the CA genotype). In the codominant and dominant models when CC homozygote genotype of miRNA-146a and of IRAK1 were used as reference group, the CG, GG and CG+GG genotypes of miRNA-146a and the CA, AA and CA+AA genotypes of IRAK1 were associated with a significant increased risk of RA. Beside, miRNA-499 rs3746444 and PADI4 rs1748033 exhibited a highly statistically significant difference in
the C allele frequency (P=0.003 and 0.002, respectively) and in the CC genotype distribution (P=0.003 and 0.05, respectively). For miRNA-499, the C allele and the homozygote genotype CC were more frequent in the RA patients than controls (52.89% vs 38.39%, respectively, for the C allele and 30.8% vs 14.3%, respectively, for the CC genotype). For PADI4, the C allele and the homozygote genotype CC were more frequent in the RA patients than controls (31.73% vs 29.58%, respectively, for the C allele and 30.77% vs 16.07% respectively for the CC genotype). In the codominant and recessive models when TT or TT+TC genotypes of miRNA-499 and of PADI4 were used as reference group, only the CC genotype of miRNA-499 and PADI4 were associated with a significant increased risk of RA.

Table 3 compared the genotype distributions of the four studied SNPs in active to inactive RA patients. There was no significant difference in the distribution of genotypes and the allelic frequency of miRNA-146a rs2910164, miRNA-499 rs3746444, and PADI4 rs1748033, neither in the allelic frequency of IRAK1 rs3027898 C/A, 50% vs 25%, respectively, while the homozygous AA genotype was more frequent in the active RA relative to the inactive RA (29.16 % vs 14.3%, respectively). But this significant difference in genotypes was disappeared when adjusting the age and sex of patients.

As IRAK1 gene is coded on the x chromosome, we evaluate the effect of IRAK1 rs3027898 genotypes on RA risk according to different gender (Table 4). A significant increased risk of
RA associated with the IRAK1 rs3027898 CA genotype (P=0.00) and A allele (P=0.00) were more evident among females compared to males.

Assessment of the 4 SNPs genotypes associations with the disease severity and activity parameters in RA patients were established in Table 5. Significant association was observed between miRNA-146a rs2910164 and IRAK1 rs3027898 polymorphisms with one of the disease severity parameters, erosions, (P=0.05 and 0.009, respectively). The percent of patients having positive erosions were significantly increased among patients carrying CG or GG genotypes in miRNA-146a, while significantly decreased among patients carrying CA genotype in IRAK1 when compared to CC genotype. In addition, IRAK1 showed a significant association with one of the activity parameters, DAS28, where the percent of patients having moderate to high disease activity were significantly decreased among patients carrying CA compared to CC genotypes (P=0.02). MiRNA-499 rs3746444 and PADI4 rs1748033 showed a significant association with one of the activity parameters, positive ANA, (P=0.002 and 0.001, respectively). Positive ANA were significantly increased among patients carrying TT or TC genotypes compared to CC genotype of miRNA-499 and among patients carrying TC or CC genotypes compared to TT genotype of PADI4. Additionally, PADI4 shows significant association with ESR levels (P=0.05), where ESR values were significantly elevated among patients carrying TT and CC genotypes when compared to TC genotypes. Beside, PADI4 shows a significant association with two of the severity parameters positive erosions and RF (P=0.02, each one). Where positive erosions were significantly increased in TC genotype and positive RF were significantly decreased among CC genotype compared to TT genotype.

### Discussion

Genetic polymorphisms of miRNA-146a rs2910164, miRNA-499 rs3746444, IRAK1 rs3027898 and PADI4 rs1748033 were analyzed with susceptibility of RA in a sample of Egyptian population. A significant difference was noticed in the distribution of the 4 studied SNPs genotypes and their minor alleles between RA patients and controls. The prevalence of miRNA-146a rs2910164 GG genotype and G allele, IRAK1 rs3027898 CA genotype and A allele, miRNA-499 rs3746444 and PADI4 rs1748033 CC genotype and their C alleles were identified as significantly higher in RA patients. By comparing the active and inactive RA groups only the CA and AA genotypes of IRAK1 rs3027898 showed a significant difference. Where, the rs3027898 CA genotype was more frequent in inactive RA and the rs3027898 AA genotype was more frequent in active RA group. No significant difference was observed in the distribution of miRNA-499 rs3746444 genotypes with the disease severity and activity parameters in RA patients, while, the CG and GG genotypes of miRNA-146a were associated with positive erosions. IRAK1 CA genotype was associated with low disease activity and negative erosions as well as AA genotype were associated with high disease activity. CC genotype of PADI4 was associated with negative RF. Finally, polymorphisms of miRNA-146a and IRAK1 were a risk factor for predisposition to RA in codominant and dominant models while, polymorphisms of miRNA-499 and PADI4 were a risk factor in codominant and recessive models.

Formerly, variants of miRNA-146a rs2910164 and its target gene IRAK1 rs3027898 were proved to be associated with susceptibility to RA [20, 21]. For miRNA-146a rs2910164, 

### Table 4. Stratified analysis between IRAK1 rs3027898 C/A polymorphism and risk of RA according to gender. Data are reported as number. rs: reference single-nucleotide polymorphism (SNP) ID. P values less than 0.05 are statistically significant.

<table>
<thead>
<tr>
<th>Variable</th>
<th>IRAK1 rs3027898 C/A (case/control)</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Genotypes</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Male</td>
<td>CC 6/12, CA 6/10, AA 4/4</td>
<td>0.72</td>
</tr>
<tr>
<td>Female</td>
<td>CC 36/62, CA 34/0, AA 18/24</td>
<td>0.00</td>
</tr>
<tr>
<td>Alleles</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Male</td>
<td>C 18/34, A 14/18</td>
<td>0.402</td>
</tr>
<tr>
<td>Female</td>
<td>C 106/124, A 70/48</td>
<td>0.00</td>
</tr>
</tbody>
</table>
similar recent study found a significant association between miRNA-146a genotype GG and the increase in risk of RA in Chinese female patients [20]. It was evidenced that estrogen could regulates miRNA-146a expression in immune cells, and miRNA-146a plays an important role in estrogen-mediated immune regulation [22]. While in contrast, other two studies observed no significant differences in miRNA-146a genotype distribution in Chinese female RA patients compared to male RA patients [23] and in Egyptian female patients compared to controls [24]. Moreover, the later study of Egyptian female RA patients reported no significant differences in all disease severity and activity parameters among different miRNA-146a genotype [24]. Studies specified that miRNA-146a is a critical manager in RA evolution as it can drive a negative feedback mechanism to prevent excess inflammation [25, 26]. The expression of miRNA-146a was believed to be increased in RA but it is unable to properly function, leading to prolonged TNF-α production [27]. The elevated expression of miRNA-146a might be due to the polymorphism of miRNA-146a rs2910164. This polymorphism involves a G>C nucleotide replacement that causes change from a C:U pair to a G:U mismatch in the stem structure of the miRNA-146a precursor, which affects the specificity of mature miRNA-146a in binding to its targets [6, 28].

Our results of IRAK1 rs3027898 were in agreement with numerous previous studies in different population. Significant difference in the IRAK1 rs3027898 A>C polymorphism distribution was reported between RA patients and controls in Greece [29]. AA genotype was associated with a significantly increased risk of RA in a Chinese population [30]. IRAK1 rs3027898 was marginally associated with RA susceptibility among Koreans in an allelic model [28]. However, in disagreement with present discoveries, IRAK1 rs3027898 C allele was reported to be associated with an increased risk of RA in China [31]. An association between inflammatory arthritis and rs3027898 CC genotype was revealed in Song et al. meta-analysis [32]. Finally, a study on Iranian female patient observed no association between the IRAK1 rs3027898 AA genotype and the risk of RA, and concluded that the CA genotype and the A allele were not a risk factor for predisposing women to RA [33]. IRAK1 is one of the serine/threonine protein kinases which play a significant role in Toll interleukin receptor (TIR) activation of NF-κB. Upon stimulation by TIR-specific ligands, IRAK1 is activated the transcription factor NF-κB [34]. Activated NF-κB increases the expression of proinflammatory cytokines such as IL-6, tissue destructive matrix metalloproteinases, TNF-α, IL-8 and chemokines [35, 36]. TNF-α and IL-1β were reported to target miRNA-146a

Table 5. Associations of miRNA-146a rs2910164 C/G, IRAK1 rs3027898 A/C, miRNA-499 rs3746444 T/C and PADI4 rs1748033 T/C gene polymorphisms with RA clinicopathological variables. Quantitative data were presented as mean ±SD; qualitative data were presented as number and percent n (%). ESR: erythrocyte sedimentation rate, ANA: anti nuclear antibody; DAS28: disease activity score; RF: rheumatoid factor, VAS: Visual analogue scale. * Significant difference from CC, TT. † Significant difference from CA, TC. P values less than 0.05 are statistically significant.
expression [11]. Then miRNA-146a targets the expression of the following genes; IRAK1, TRAF6, IRF5, STAT1 and consequently acts as a negative regulator in Toll-like receptors (TLRs) and INF-α signaling pathways [24, 37].

Another polymorphisms rs3746444 of miRNA-499 and its target gene PADI4 were also reported to be involved in RA risk [22, 38]. MiRNA-499 gene polymorphisms were investigated to be associated with ADs, however, the results are still debated. The homozygous genotype CC and allele C were found to be risk factors for predisposition to RA in Iranians [39]. The miRNA-499 polymorphism was discovered to be an independent factor of RA and was associated with RA risk, severity, and activity in Egyptian female patients [23]. Our overall results were compatible with these two studies. However, no significant difference in miRNA-499 SNPs was found between RA patients and healthy controls and the miRNA-499 SNP association with anti-CCP antibody production was firstly investigated in a Han Chinese population [22]. Similarly, other study showed that miRNA-499 SNPs were not significantly associated with the risk for RA in Chinese people [30]. Therefore, a meta-analysis concluded that, miRNA-499 SNPs may be associated with susceptibility to RA in Mediterranean populations, but not in East Asians [40].

Present PADI4 results were concordant with previous report that determined an increased risk of RA in Indian population possessing rs1748033 C allele. The report also ascertained an association between anti-CCP antibody levels with high DAS28 values in RA patients [41]. Similarly, earlier studies suggested that the anti-CCP antibodies and the susceptible haplotype of PADI4 serve as markers for susceptibility and severity of the disease [42, 43]. Likewise, inconsistent results have produced from the association studies between PADI4 SNPs and RA in different populations. PADI4 rs1748033 SNP was found to be associated with RA in Korean [44], Japanese population [38, 45], and Chinese Han population [46], while this variant was not associated with RA in UK Caucasian population [47], German [48] and Spanish population [49]. PADI4 rs1748033 SNP has been shown to be associated with greater risk of RA in men than in women and in ever-smokers than in never-smokers [50].

PADI4 gene is a vital target of miRNA-499 as it encodes PADI4 enzyme, which produces the citrullinated peptides identified by anti-CCP [51] through post-translational alteration of arginine residues to citrullines [52]. The role played by the citrullination in a number of proteins, is not known, but it has been related to some apoptosis function [53]. From the immunological view, it can be stated that post-translationally changed self-antigens are more likely to break tolerance towards self. Anti-CCP antibody can early diagnose and predict joint damage in RA [54]. In this way miRNA-499 can control the production of anti-CCP antibody by controlling PADI4 gene expression. SNP rs3746444 is located at the premiRNA regions of miRNA-499 and may influence both the binding of target miRNAs to 3p mature miRNAs and premiRNA maturation of 5p and 3p miRNAs. Researchers observed that RA patients with miRNA-499 CT genotype had a higher level of anti-CCP antibody more than homozygote [22, 23].

In spite of the considerable efforts to explore the possible relationship between the 4 SNPs and RA risk, some limitations should be considered. First, as this is a hospital-based, case-control study, selection bias is unavoidable, and the subjects are not fully representative of the general population. Second, we have no data regarding anti-CCP antibodies; HLA-DRB1 shared epitope and smoking history. Thus, we could not determine the association between these 4 SNPs and these factors; however, this is the first study that addresses association of IRAK1 rs3027898 and PADI4 rs1748033 with susceptibility of RA in Egyptians. Third, we are aware that a single case-control study may not be sufficient to fully interpret the relationship between them and susceptibility for RA in Egypt because of the relatively small number of patients involved. Nevertheless, we believe that the results of this study provide an important input into the debate concerning the clinical relevance of studied variant.

In conclusion, our study provides strong evidence that miRNA-146a rs2910164, miRNA-499 rs3746444, IRAK1 rs3027898 and PADI4 rs1748033 polymorphisms may
contribute to the risk of RA in Egyptian population. However, further studies in other ethnic groups and with larger sample sizes are required to confirm our findings. Consistent with growing evidence, the present study has demonstrated that miRNA polymorphisms may be suitable for use as diagnostic biomarkers for RA in future. Nevertheless, by identifying new genetic predisposing factors to immune diseases, even though overlapping, we highlight new pathways in disease susceptibility and possibly in its therapeutic intervention.

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Disclosure Statement

No conflict of interests exists.

References


