Effects of an Oral CRTh2 Antagonist (AZD1981) on Eosinophil Activity and Symptoms in Chronic Spontaneous Urticaria

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Abstract

Background: Approximately 50% of patients with chronic spontaneous urticaria (CSU) experience symptoms that are not fully controlled by antihistamines, indicating an unmet clinical need. **Objective:** To evaluate the effects of the selective CRTh2 antagonist AZD1981 on symptoms and targeted leukocytes in adults with persistent CSU despite treatment with H₁-antihistamines. **Methods:** We performed a single-center, randomized, placebo-controlled study involving adult CSU subjects with symptoms despite daily antihistamines. The subjects underwent a 2-week placebo run-in and 4 weeks of double-blinded therapy with either AZD1981 40 mg TID or placebo, followed by a 2-week placebo washout. The primary objective was to assess the effect of AZD1981 on CSU signs and symptoms. Secondary objectives included the effects of AZD1981 on prostaglandin D₂ (PGD₂)-induced eosinophil shape change, circulating leukocyte subsets, CRTh2

expression on blood leukocytes, and total blood leukocyte histamine content. **Results:** Twenty-eight subjects were randomized to AZD1981 or placebo, with 26 subjects completing the study. The urticaria activity scores declined during the treatment phase in both groups, and they were significantly reduced in the AZD1981 group at the end of washout. AZD1981 treatment increased circulating eosinophils and significantly impaired PGD₂-mediated eosinophil shape change. CRTh2 surface expression rose significantly on blood basophils during active treatment. No serious adverse events were observed. **Conclusions:** This is the first study to examine the efficacy of a CRTh2 antagonist in antihistamine-refractory CSU. AZD1981 treatment was well tolerated, effectively inhibited PGD₂-mediated eosinophil shape change, shifted numbers of circulating eosinophils, and reduced weekly itch scores more than hives during treatment and into washout. Further studies are needed to determine whether inhibition of the PGD₂/CRTh2 pathway will be an effective treatment for CSU.

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Introduction

Chronic spontaneous urticaria (CSU) is defined as pruritus and hives occurring daily or almost daily for at least 6 weeks without an underlying etiology [1]. Although second-generation H₁-antihistamines are a first-line therapy for CSU, 30–50% of patients fail to achieve symptom control with antihistamines alone [2–4]. Cyclosporine, sulfasalazine, dapsone, Plaquenil, and mycophenolate mofetil have been used in more severe cases of CSU, but each has the potential for significant toxicities that require frequent laboratory monitoring [4, 5]. Omalizumab has been approved for antihistamine-refractory CSU by the US Food and Drug Administration (FDA) at 150 or 300 mg every 4 weeks, and by the European Medicines Agency (EMA) at 300 mg every 4 weeks. The optimal dose of omalizumab is 300 mg every 4 weeks [6, 7]; yet 47–66% of patients treated fail to achieve complete symptom control (urticaria activity score over 7 days [UAS7] = 0) at this dosage [4, 6–8]. The disadvantages of omalizumab treatment include the high cost of a biologic agent, the requirement for subcutaneous injection in a physician's office, and the need for an epinephrine auto-injector prescription due to the risk of drug-related anaphylaxis of up to 0.2% [9, 10]. Thus, there remains a need for safe, affordable, and convenient therapies for antihistamine-refractory CSU.

While the pathogenesis of CSU remains unknown, biopsies of CSU skin lesions reveal evidence of mast cell degranulation and infiltration by CRTh2-bearing leukocytes which include basophils, eosinophils, and Th2 lymphocytes [11]. The endogenous agonist for CRTh2 is prostaglandin D₂ (PGD₂) [12, 13], which is synthesized and released by activated mast cells [14–18]. We recently reported that CRTh2 levels were notably decreased on blood basophils and eosinophils from CSU subjects as compared to healthy controls [19]. This finding is consistent with ongoing *in vivo* receptor-ligand engagement, leading to CRTh2 receptor internalization [20, 21].

The cellular effects of CRTh2 activation include shape change, chemotaxis, upregulation of CD11b, Th2 cytokine production by Th2 cells and group 2 innate lymphoid cells (ILC2s), eosinophil cationic protein (ECP) release by eosinophils, and enhancement of allergen-induced basophil histamine release [21–28]. Animal studies have shown that PGD₂ propagates eosinophil mobilization from bone marrow [29] and mediates eosinophil infiltration into the airway and skin [30–33]. PGD₂ injection into healthy human skin has been shown to trigger wheal and flare responses [34, 35]. While the functional role of the CRTh2 activation

in CSU is unknown, two single nucleotide polymorphisms in the CRTh2 promoter region are associated with antihistamine resistance in CSU [36].

AZD1981 is an oral, potent, selective, reversible antagonist of CRTh2 [37]. Preclinical studies have shown that AZD1981 blocks CRTh2-mediated shape change in human eosinophils and basophils *in vitro*, inhibits CRTh2-mediated chemotaxis of human Th2 cells and eosinophils, and blocks PGD₂-mediated eosinophil release from bone marrow in guinea pigs [37]. In clinical trials, oral CRTh2 antagonists have reduced esophageal eosinophils in eosinophilic esophagitis [38] and provided an improvement in airway function and protection against allergen-induced symptoms in patients with allergic rhinoconjunctivitis and asthma [39–43]. Here we report the results of the first study to examine the safety and efficacy of the selective oral CRTh2 antagonist AZD1981 as add-on therapy to standard antihistamines in CSU.

Subjects and Methods

Study Design and Participants

This was a phase II, single-center, randomized, double-blind, placebo-controlled, parallel group study which recruited patients with CSU, aged 18–65 years, who remained symptomatic despite treatment with standard doses of non-sedating H₁-antihistamines (for details on the inclusion and exclusion criteria, see the online Supplementary Material section; for all online suppl. material, see www.karger.com/doi/10.1159/000496162). This study involved a 1-week screening period (days –7 to 0), a 2-week single-blind placebo run-in (days 1–14), a 4-week randomized double-blind treatment period (days 15–42), and a 2-week single-blind placebo washout (days 43–56) (Fig. 1).

During the screening and placebo run-in periods, the subjects were required to have an *in-clinic* physician-assessed UAS of ≥ 2 (range 0–6) and demonstrate a UAS7 of ≥ 8 . The UAS is a validated measure of CSU disease activity which scores the intensity of pruritus (0–3) and the number of hives (0–3) with a maximum value of 6 [44]. The UAS7 is the sum of the daily average UAS (average of a.m. and p.m.) for 7 days with a maximum value of 42 [45]. The participants were required to show compliance with twice-daily symptom recording in eDiaries throughout the study.

Subjects who met the screening and run-in criteria for symptom activity were randomized on day 15 in a 1:1 ratio to receive either AZD1981 40 mg TID or matched placebo administered orally as 10-mg tablets. This dosage was selected based on preclinical pharmacokinetic and pharmacodynamic studies comparing BID to TID dosing as well as safety considerations. Throughout the study, the participants were required to maintain stable doses of their pre-randomization H₁-antihistamine treatment. They could take diphenhydramine 25 mg up to TID as a rescue medication. Subjects who required therapies other than diphenhydramine (e.g., prednisone) to treat disease symptoms were removed from the study.

Fig. 1. Study design.

Fig. 2. a, b Effect of AZD1981 therapy and washout on the urticaria activity score over 7 days (UAS7) (**a**) and weekly itch scores (**b**). **c, d** Effect of placebo on UAS7 (**c**) and weekly itch scores (**d**). Values are shown as mean $\pm$ SEM. * $p < 0.05$. AZD1981 group: $n = 12$; placebo group: $n = 11$.

All research team members were blinded to the treatment assignments – with the exception of the site pharmacists who dispensed the study drug, but they did not interact with the participants. The pharmacists randomized the subjects in blocks of 4 to maintain a balanced randomization.

Study Endpoints

The primary efficacy endpoint was the change in diary-based clinical symptoms as measured by the UAS7 from baseline (placebo run-in, days 8–14) to the last week in the treatment period (days 36–42).

Secondary endpoints included the following: the safety of AZD1981; the pharmacodynamics of AZD1981 through an ex vivo

eosinophil shape change assay; the pharmacokinetics of AZD1981 through a serum bioassay; and the quality of life benefit as measured by the Dermatology Life Quality Index (DLQI) questionnaires. Additional efficacy endpoints included the change from baseline to the last week of treatment of the following: weekly itch severity scores (ISS); the weekly score for the number of hives; the weekly score for sleep interference; rescue medication use; and the presence of angioedema. Safety analyses included adverse events (AEs), severe AEs (SAEs), laboratory abnormalities, and physical examination abnormalities.

Exploratory endpoints were as follows: the change in circulating leukocyte population numbers that are targeted by CRTh2, such as blood basophils and eosinophil and lymphocyte numbers,

by automated analysis; the change in basophil histamine release and whole blood histamine content; and pruritus-free and hive-free days during the treatment period. For details on the laboratory experiments, please see the Methods section in the online supplementary materials.

Data Analysis

The analysis of the primary endpoint (change in UAS7 from baseline to week 4) was made using pairwise comparisons. The UAS7 obtained 1 week prior to randomization was used as the baseline score. The minimally important difference of the UAS7 for patients with CSU is estimated to be 9.5–10.5 [45]. The Wilcoxon matched-pairs signed-rank test (Prism 7.00) was used for intragroup comparisons between UAS7 values at baseline, the end of treatment, and the end of placebo washout. The secondary endpoints were analyzed using pairwise comparisons between the two study groups. Treatment comparisons of change from baseline to week 6 in the weekly ISS and the number-of-hives score were made using the Wilcoxon signed-rank test. For missing patient-entered data (ISS and number of hives), the preceding recorded observation was carried forward. Patient characteristics were analyzed using the χ^2 test and the two-sample t test for categorical and continuous variables, respectively. Any AEs or SAEs were tabulated by system organ class and preferred term, as well as by seriousness, severity, and treatment relatedness, and reported by frequency. p values ≤ 0.05 were considered significant.

Results

Subject Characteristics

A total of 38 subjects consented, and 28 subjects were randomized to treatment with AZD1981 or placebo. Following randomization, 1 subject withdrew due to CSU disease progression and 1 subject withdrew due to inability to comply with study visits, leaving 26 subjects (93%) who completed the study (online suppl. Fig. S1). The baseline characteristics were similar across the treatment groups (Table 1). All CSU subjects were taking approved doses of H_1 -receptor antagonists. No subjects were taking leukotriene receptor antagonists, aspirin, or NSAIDs (nonsteroidal anti-inflammatory drugs) during the study.

Patient-Reported Outcomes

During the course of the study, the UAS7 decreased during the treatment phase in both the active and the placebo group, but they reached a significant reduction only in the AZD1981 group at the end of washout (Fig. 2). Active treatment had a greater effect on weekly ISS (35.9% reduction) than on hives (20.5% reduction; not shown), but only itch was significantly reduced at the end of washout compared to baseline ($p = 0.001$ vs. $p = 0.33$).

This delayed effect raised the possibility of a carryover effect, as has been seen in prior studies with CRTh2 an-

Table 1. Characteristics of the subjects who completed the study

	Active (n = 14)	Placebo (n = 12)	p value
Age, years	41.85 (23–65)	45.17 (23–64)	0.54
Female	10 (71.4)	10 (83.3)	0.47
Race/ethnicity			
Caucasian	6 (42.9)	4 (33.3)	0.62
African American	4 (28.6)	4 (33.3)	0.79
Asian	3 (21.4)	2 (16.7)	0.61
Pacific Islander	0	1 (8.3)	0.31
More than 1 race	1 (7.1)	1 (8.3)	0.91
Hispanic	0	0	–
Baseline symptom scores (days 8–14)			
UAS7	20.61	22.79	0.83
Weekly itch scores	9.71	10.17	0.81
Weekly hives scores	11.11	12.46	0.93
Aspirin/NSAID sensitivity ¹			
Sensitive	6 (42.9)	3 (25)	0.34
Tolerant	6 (42.9)	9 (75)	0.10
Unknown	2 (14.3)	0 (0)	0.17
Current atopy			
Allergic rhinitis	5 (35.7)	2 (16.7)	0.19
Asthma	3 (21.4)	0	0.06
Atopic dermatitis	1 (7.1)	2 (16.7)	0.45

Values are expressed as n (%) or median (range). p values are based on a χ^2 test for the categorical variables, and on a two-sample t test for the continuous variables. UAS7, urticaria activity score over 7 days. ¹ Aspirin/NSAID sensitivity was determined (by patient interview) if a subject reported increased symptoms within 24 h of taking either an aspirin or NSAIDs, which led to avoidance.

tagonists [39]. The UAS7 also decreased in the placebo group from baseline to the end of treatment, but this did not reach statistical significance. There was no difference in symptom response to AZD1981 based on aspirin/NSAID sensitivity or tolerance, baseline laboratory assessments (CRTh2 expression, blood eosinophil percentage, and eosinophil shape change response), baseline UAS, or medication compliance. There was no significant change in diphenhydramine use, DLQI, angioedema episodes, sleep disturbance, emergency room visits, or hive-free days with AZD1981 treatment relative to placebo.

Compliance with eDiary entry was high (>90%) throughout the study, and it was similar in the two groups. Of the 14 subjects who completed the study in the AZD1981 arm, 1 patient was excluded from the final data analysis due to a dispensing error which resulted in the patient receiving AZD1981 during the washout period. No other dispensing errors occurred. One active patient and 1 placebo patient were excluded from symptom anal-

Fig. 3. Effect of AZD1981 therapy (a) and placebo (b) on ex vivo eosinophil shape change shown as net change (baseline value – no PGD₂) for the subjects at randomization, the end of treatment, and the end of washout. Values are shown as mean $\pm$ SEM. PGD₂, prostaglandin D₂.

Fig. 4. Circulating numbers of CRTh2⁺ leukocytes based on complete blood count (CBC). Shown is the percentage of peripheral blood eosinophils (a) and basophils (b) at baseline, end of treatment, and washout. Data are not shown for 3 active subjects (2 subjects lacked a CBC differential at the end of washout, and 1 subject was administered AZD1981 during the washout period). Values are displayed as mean $\pm$ SEM.

ysis due to malfunction or loss of the eDiary device. Pill counts after each 2-week dispensation were performed by the research pharmacists, and they revealed an overall compliance level of 86.54 and 84.79% in the active and the placebo group, respectively, during the 4-week, randomized treatment period.

Effects on Eosinophil Shape Change

Eosinophil shape change after PGD₂ stimulation is shown as a mean increase in forward scatter above vehicle for subjects at randomization, the end of treatment, and the end of washout (Fig. 3). At baseline, the in vitro addition of AZD1981 significantly impaired PGD₂-mediated eosinophil shape change at a 0.1- μ M dose (not shown)

and a 1- μ M dose as calculated by area under the curve (AUC) (online suppl. Table S1). In vitro incubation with 1 μ M AZD1981 significantly decreased the AUC of the PGD₂-mediated eosinophil shape at baseline (prior to randomization) in both the active and the placebo arm ($p = 0.0005$ and $p = 0.002$, respectively).

At the end of the treatment period, ex vivo PGD₂-induced eosinophil shape change was significantly inhibited in the AZD1981-treated group but not in the placebo group (Fig. 3). Treatment with oral AZD1981 showed similar impairment in PGD₂-mediated eosinophil shape change to in vitro inhibition with 1 μ M AZD1981 performed at baseline. Furthermore, peak serum drug levels achieved at the end of AZD1981 therapy fell within or

Fig. 5. Expression of CRTh2 on blood basophils and eosinophils is shown at baseline, the end of treatment, and the end of washout. **a, b** Basophil CRTh2 levels in active (**a**) and placebo (**b**) recipients. **c, d** Eosinophil CRTh2 levels in active (**c**) and placebo (**d**) recipients. Values are shown as mean $\pm$ SEM. * $p < 0.05$. MFI, median fluorescent intensity.

above this range (online suppl. Table S2). This inhibitory effect of AZD1981 therapy trended toward baseline at the end of washout. In the placebo group, there were no significant changes observed at the end of treatment and washout relative to baseline (online suppl. Table S1B).

Effects on CRTh2⁺ Leukocyte Percentages in Blood

The functional inhibition of eosinophil shape change observed at the end of AZD1981 treatment (Fig. 3a) occurred in parallel with a rise in circulating eosinophils at the end of treatment (Fig. 4a) that did not reach statistical significance (from 2.49 ± 0.37 to $3.5 \pm 0.70\%$, $p = 0.08$). This rise in circulating eosinophils appeared to reverse at the end of washout ($2.9 \pm 0.46\%$, $p = 0.40$), supporting a transient impairment in eosinophil trafficking to the skin. Shifts in the percentage of blood basophils did not follow a similar pattern as eosinophils in the AZD1981 group, but assessments were limited by the presence of notable basopenia among subjects (Fig. 4b). No changes in these parameters were seen in the placebo group.

Effects on Basophil Surface Markers and Function

At the end of AZD1981 therapy, levels of CRTh2 surface expression rose significantly on blood basophils ($p =$

0.03) (Fig. 5a) and trended towards baseline at the end of washout ($p = 0.13$). Similar changes in basophil CRTh2 expression were not seen in the placebo group (Fig. 5b) or on blood eosinophils (Fig. 5c, d). We did not observe changes in the expression levels of other basophil chemotactant surface receptors, including CCR1, CCR3, or CCR5 (not shown). There was also no significant change in CD11b expression on basophils or eosinophils, or in CD203c expression on basophils, which are known to be enhanced after in vitro exposure to PGD₂ [24]. We did not observe significant changes in total blood leukocyte histamine content, which is a reflection of the presence of blood basophils, or of changes in basophil histamine release (online suppl. Fig. S2). Both of these measures for blood basophils were limited by the small numbers of subjects in this study.

Safety and Tolerability

During the treatment and follow-up periods, no significant safety AEs or complete blood count or liver function test abnormalities occurred. A total of 9 patients (64.2%) in the AZD1981 group experienced at least 1 AE, compared with 8 patients (66.6%) in the placebo group (Table 2). The majority of AEs were mild and did not lead

to any patients withdrawing from the study. The most frequently reported AE among the subjects was infection (5 in the active group and 5 in the placebo group).

Discussion and Conclusion

This is the first study to examine the safety and efficacy of a CRTh2 antagonist in CSU. For this study, we allowed the use of antihistamines, as a major goal of this our study was to evaluate the use of AZD1981 as an add-on therapy in antihistamine-refractory CSU patients. As such, we required all subjects to take a standard dose of H₁-antihistamines throughout the study. Regardless, all patients continued to have moderate disease despite antihistamine use, and we excluded subjects whose symptoms were controlled on antihistamines.

With AZD1981 treatment, the UAS7 decreased during the treatment phase, but it reached a significant decline from baseline at the end of drug washout. During the same time period, AZD1981-treated subjects experienced a 35.9% decline in itch component scores. This suggests the possibility of a carryover effect, as has been seen in other diseases targeted by a CRTh2 antagonist [39]. In the placebo group, decreases were observed in UAS7 and itch scores from baseline to the end of treatment which did not reach statistical significance. These findings were confirmed using one-way ANOVA, with significant differences ($p < 0.01$) being seen only in the active group for UAS7 and itch. Active treatment had a greater effect on weekly ISS than on hive scoring (Fig. 2). However, this study failed to meet the primary endpoint (UAS7 decrease ≥ 9.5). There was no significant change in DLQI, sleep interference, or in-clinic UAS as a result of this brief 4-week treatment period. Rescue medications were rarely taken by our subjects, and there was no significant impact of AZD1981 treatment on rescue medication use.

Consistent with the in vitro effects of AZD1981 on eosinophil migration, we found marked inhibition of ex vivo eosinophil PGD₂-induced shape change in CSU subjects on active therapy (Fig. 3) along with a transient rise in circulating eosinophils (Fig. 4). Both changes trended towards baseline during drug washout, suggesting a drug-induced effect on eosinophils. This suggests that PGD₂ may be a central pathway for eosinophil migration to the skin. Surprisingly, these changes did not correspond with a significant change in CRTh2 surface expression on blood eosinophils. Although in vitro PGD₂ binding to CRTh2 leads to receptor internalization [20, 21], our results suggest that the decrease in eosinophil CRTh2 observed in

Table 2. Reported adverse events

	Active (n = 14)	Placebo (n = 12)	p value
Headache	0	2 (16.6)	0.10
Musculoskeletal	3 (21.4)	1 (8.3)	0.36
Infection	5 (35.7)	5 (41.7)	0.76
Sleep impairment	1 (7.1)	1 (8.3)	0.91
Respiratory disorders (noninfectious)	0	0	–
Gastrointestinal disorders	5 (35.7)	2 (16.6)	0.34
Blood disorders	0	0	–
Eye and vision disorders	0	1 (8.3)	0.27
Cardiovascular disorders	1 (7.1)	1 (8.3)	0.91
Renal disorders	0	0	–
Reproductive disorders	0	0	–
Skin or hair disorders	0	2 (16.6)	0.10
Immunologic disorders	0	0	–

Values are expressed as n (%). p values are based on the χ^2 test for the categorical variables.

CSU subjects compared to healthy controls [19] may not solely reflect CRTh2 receptor activation by PGD₂. For instance, in vitro IL-5 stimulation significantly reduces human eosinophil CRTh2 expression [20], and it was shown that IL-5 m-RNA and protein expression are increased in chronic urticaria lesions [11, 46].

Although eosinophils are a consistent infiltrating cell in urticarial skin biopsies [11, 47], their role in CSU disease is unknown. The changes seen in eosinophil shape change and circulating eosinophils with CRTh2 antagonism suggest a role for mast cell-derived PGD₂ in the migration of eosinophils into skin lesions in CSU and contribution to itch. While eosinopenia is not a feature of CSU, there are data to suggest overall lower eosinophil counts in CSU patients than in healthy controls [48]. In contrast to this study, CRTh2 antagonism in subjects with persistent eosinophilic asthma led to reductions in sputum and tissue eosinophils and in asthma symptoms but did not shift peripheral eosinophil counts [43]. Thus, the effects of PGD₂ may be disease and tissue compartment specific.

In support of impairment of PGD₂ actions on basophils, levels of CRTh2 surface expression rose significantly on blood basophils at the end of AZD1981 therapy (Fig. 5a). Unlike what is observed in eosinophils, this finding in basophils supports the hypothesis that the reduced levels of basophil CRTh2 expression previously noted in active CSU subjects are due to ongoing PGD₂-mediated receptor downregulation [19]. We were limited in the performance

of additional functional assays of basophil CRTh2 inhibition such as basophil shape change due to the basopenia associated with CSU. However, the effects of AZD1981 on basophil receptor expression were specific to CRTh2, as we did not observe changes in the expression levels of other basophil chemoattractant surface receptors that may also have roles in basophil skin recruitment.

Interval improvement in clinical symptoms in CSU patients is reported to occur with rising blood basophil numbers [49]. After 4 weeks of AZD1981 treatment, we did not find significant changes in the percentage of basophils or a rise in total blood leukocyte histamine content. Given the wide variance of the latter measure in CSU disease, we were underpowered to detect increases in basophil number based on past experience [50]. In addition, the lack of change in other basophil chemoattractant receptors may also explain the lack of change in measures of circulating basophils.

The reason for the decrease in itch with AZD1981 therapy is currently unknown. Intradermal PGD₂ injections trigger wheal and flare responses but do not produce immediate itching [35]. We did not alter basophil histamine release mediated through the IgE receptor or spontaneous histamine release, consistent with past studies that showed that PGD₂ enhances antigen-mediated basophil histamine release but not that from polyclonal IgE receptor crosslinking [51]. Therefore, the improvements in CSU subjects' itch in our AZD1981 trial may be through indirect actions on mast cells and/or basophils.

The late onset of itch reduction in the AZD1981 recipients could be the result of a reduction in eosinophil recruitment to the skin and/or eosinophil-derived granule proteins (EDGPs) that contribute to itch pathways. Previous studies have shown that PGD₂ activation of eosinophils triggers ECP release [22]. Thus, it is possible that PGD₂ may trigger the release of other EDGPs. Prior studies have revealed evidence of eosinophil activation in CSU, specifically the presence of extracellular major basic protein (MBP) and ECP in CSU skin lesions and elevated levels of ECP and MBP in the serum of CSU patients despite normal peripheral eosinophil counts [52, 53]. MBP produces an immediate erythematous wheal when injected intradermally in humans and guinea pigs [54, 55]. Eosinophil MBP and eosinophil peroxidase were recently shown to trigger histamine release from skin-derived mast cells via MrgX2 receptors, which are increased in CSU skin mast cells compared to those of healthy controls [56]. Thus, there is a need for additional studies to fully understand the consequences of CRTh2 activation on eosinophil EDGP release.

Among the limitations of this study are the small number of subjects, the short duration of therapy and washout periods, and the lack of skin tissue biopsies to correlate with the peripheral blood leukocyte findings. In addition, the optimal dose of AZD1981 in CSU is unknown, and we were limited to the use of a single dose (40 mg TID) in this study. AZD1981 was previously studied in allergic asthma at doses ranging from 10 to 400 mg BID but failed to produce a clinically relevant improvement in lung function in patients with allergic asthma on inhaled corticosteroid and long-acting β_2 -agonist therapy [57]. Our dose was selected based on preclinical pharmacokinetic and pharmacodynamic studies comparing BID to TID dosing and safety considerations. While it is possible that the use of a higher dose may have achieved the primary endpoint, a higher frequency of liver function abnormalities had previously been noted with higher doses (400 mg BID) [57]. Despite this, we did observe a delayed yet modest reduction in disease activity (particularly in itch), an impairment in eosinophil shape change and potentially migration, and a rise in basophil CRTh2 expression, which would suggest that this pathway was inhibited by AZD1981.

In summary, this is the first study to examine the safety and efficacy of a CRTh2 antagonist in antihistamine-refractory CSU. Our findings for AZD1981 support the hypothesis that the CRTh2 pathway contributes to the formation of CSU skin lesions and symptoms, particularly for eosinophils. AZD1981 was well tolerated, with no significant safety issues. AZD1981 appeared to have a delayed positive effect on itch more so than on wheals, although the study failed to meet the primary endpoint in UAS7 reduction at the dose studied. Given these findings, further studies are needed with longer treatment periods and higher dose ranges to determine whether inhibition of the PGD₂/CRTh2 pathway will be an effective treatment for CSU.

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Statement of Ethics

This study was conducted using the International Conference on Harmonization E6 Guidelines for Good Clinical Practice. All subjects gave their written informed consent. Before study initiation, the protocol and the written informed consent documents were approved by The Johns Hopkins Hospital Institutional Review Board (NA_00089252).

Disclosure Statement

C.W. was in full-time employment with AstraZeneca during the design, initiation, and conduct of this study. He had no role in data collection or analysis. The remaining authors have no other conflicts of interest to declare.

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Author Contributions

All authors have read and approved all versions of the manuscript, its content, and its submission.

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