Case Report

The Potential and Limitations of Precision Oncology: Lessons Learned from Whole-Exome Sequencing in an Exceptional Response to Everolimus in Advanced Renal Cell Carcinoma

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Abstract

Through elucidating the genetic mechanisms of drug sensitivity, precision medicine aims to improve patient selection and response to therapy. Exceptional responders are patients that exhibit exquisite and durable responses to targeted therapy, providing a rare opportunity to identify the molecular basis of drug sensitivity. We identified an exceptional responder to everolimus, an oral inhibitor of the mammalian target of rapamycin (mTOR) pathway, in a patient with advanced renal cell carcinoma. Through whole-exome sequencing on pretreatment and metastatic tumor DNA, we identified alterations in several mTOR pathway genes, with several mutations implicated in mTOR activation. Importantly, these alterations are currently not included in commercially available next-generation sequencing panels, suggesting that precision medicine is still limited in its ability to predict responses to mTOR-targeted therapies. Further research to discover and validate predictive biomarkers of response to everolimus and other targeted therapies is urgently needed. Given the rarity of patients with exceptional responses to targeted agents, cooperative efforts to understand the molecular basis for these phenotypes are essential.

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Introduction

Advances in genomics and an expanded understanding of disease biology has transformed the therapeutic landscape of advanced renal cell carcinoma (RCC). Identification of mTOR signaling as a central element of RCC disease biology has led to the development and application of mTOR inhibitors for treatment of advanced disease [1–3]. Temsirolimus and everolimus, both analogs of rapamycin (rapalogs), allosterically inhibit mTOR complex 1 (mTORC1) and block mTOR activation. However, the clinical benefit of rapalogs is typically modest with reported progression-free survival <6 months. Despite limited benefit in unselected patients, there are multiple reports of outlier cases where patients demonstrate exceptional therapeutic response to these drugs.

Previous studies suggest that genomic alterations in mTOR pathway genes are potential determinants of mTOR inhibitor sensitivity in RCC [4–6]. Understanding the molecular basis of response is critical to establish predictive biomarkers for these therapies. In this study, we performed whole-exome sequencing (WES) of tumor DNA from a patient with advanced RCC who achieved durable disease control with everolimus.

Case Report

We identified a 60-year-old male patient with an exceptional response to everolimus. He presented initially with gross hematuria and flank pain. CT and MRI demonstrated a large 12-cm mass replacing the left kidney with invasion of the left psoas muscle. There was no evidence of metastatic disease. He underwent radical nephrectomy 4 weeks after presentation. On pathologic examination, he was confirmed to have RCC with clear cell and papillary features, Fuhrman grade 3, with gross involvement of the renal vein. Eight lymph nodes were examined with no evidence of nodal metastases. He was treated with adjuvant sunitinib on a clinical trial. The patient was unable to tolerate therapy, and sunitinib was discontinued after 1 month.

One year after nephrectomy, surveillance imaging demonstrated new pulmonary and hepatic lesions, as well as abnormal tissue in the left nephrectomy bed and left adrenal gland. Liver biopsy confirmed metastatic RCC. The patient was started on pazopanib as first-line therapy for metastatic disease. He had mild toxicity but did not require dose reduction or interruption. Ten weeks after starting pazopanib, a mixed response was seen on imaging. Although his pulmonary metastases were stable to slightly improved, his disease had clearly progressed in the liver (Fig. 1a, d). The patient was started on everolimus at this time.

Disease in the lung, liver, adrenal gland, and nephrectomy bed was significantly improved after 11 weeks of everolimus therapy. His lesions continued to regress at the 6- and 9-month time points and remained stable for 3 years (Fig. 1c–f). At that time, an MRI of the orbit was ordered to evaluate ptosis and proptosis of the right eye. A solitary site of metastatic progression was discovered in the right frontal sinus causing erosion of the calvarium with extension to the scalp, mass effect on the right frontal lobe, and extension to the right orbit and ethmoid sinus. The metastasis was surgically resected with pathologic examination confirming metastatic clear cell carcinoma. The resection was considered clinically complete, and no adjuvant radiation was recommended. Despite this recurrence, his other metastatic disease remained stable with only a sub-centimeter nodule in the lung and no disease detectable in the abdomen and pelvis. Given good tolerance and lack of progression elsewhere, everolimus therapy was continued.

Two years after resection, MRI was concerning for progression on the basis of several small areas of abnormal enhancement. He was treated with stereotactic radiosurgery to right nasal, right temporal, and right frontal dural lesions. There was no evidence of progression elsewhere, and he remained on everolimus. His disease remained stable for an additional 2 years. Imaging at

Fig. 1. Radiographic response to everolimus therapy. CT of the liver (**a**) and lung prior to starting everolimus (**d**); liver and lung after 6 months of everolimus (**b, e**); liver and lung after 80 months of everolimus (**c, f**). The metastatic lesions are indicated by white arrows.

that time demonstrated multiple lesions in the liver and spleen, as well as osseous metastatic disease. Everolimus therapy was discontinued, and the patient was started on nivolumab and denosumab. In total, the patient remained on everolimus for >7 years.

To identify potential molecular mechanisms for the extended benefit in response to everolimus, we performed WES on DNA from the primary (pretreatment) and metastatic tumor obtained at the time of surgical resection. The coding regions from approximately 25,000 genes were sequenced, with 90% of exons achieving 20x coverage or greater. Analysis of mTOR pathway genes revealed variants in several genes associated with mTOR signaling (Table 1). The variants identified are present in the dbSNP (Single Nucleotide Polymorphism Database); however, only the *PIK3C2G* (P146L) mutation was found in the COSMIC (Catalogue of Somatic Mutations in Cancer) database. The alterations identified include 2 phosphoinositide 3-kinases (PI3Ks) that are involved in regulating Akt activation and missense mutations in *PIK3R2* and *PIK3C2G* [7]. We also identified a missense mutation in diacylglycerol kinase zeta (*DGKZ*), encoding an enzyme that functions in mTORC1 activation [8]. In addition, we identified an alteration in protein kinase D (*PRKD3*) that activates Akt through diacylglycerol signaling [9]. A missense mutation was also detected in the insulin receptor (*INSR*) gene which encodes a receptor critical in activation of PI3K/Akt signaling and providing feedback regulation of mTORC1. Finally, we identified a missense mutation in eukaryotic translation initiation factor 3 (*EIF3B*) encoding an essential scaffolding component of the EIF3 complex activated by mTORC1 and functions in regulating protein translation [10].

To identify a potential genomic driver of disease recurrence in this patient, we compared alterations discovered in the primary tumor to those found in the metastatic lesion from the right frontal sinus obtained at the time of recurrence. Our analysis focused on alterations in mTOR pathway genes as a potential mechanism of everolimus resistance. Unique to the metastatic tumor are mutations in *INSR*, *PIK3R2*, and *DGKZ* genes, all of which are involved in either

Table 1. Gene variants identified in WES analysis

Gene	Tumor location	Mutation type	cDNA change	Protein change	Molecular function
Genes in the mTOR pathway					
EIF3B	P, M	Missense	c.190T>C	S64P	Protein biosynthesis; activated by mTORC1 to initiate translation
PIK3C2G	P	Missense	c.437C>T	P146L	PI3K catalytic subunit type II; promotes prolonged activation of Akt2
PRKD3	P	Missense	c.1333G>T	L445I	Mediates DAG signaling; involved in Akt activation and angiogenesis
DGKZ	M	Missense	c.29C>A	A10D	Transferase, lipid kinase; activates mTORC1 through DAG metabolism
INSR	M	Missense	c.5 G>C	A2G	Metabolism; activates PI3K/Akt signaling; negatively regulates mTORC1
PIK3R2	M	Missense	c.700A>C	S234R	PI3K regulatory subunit (p85β); promotes constitutive activation of PI3K/Akt
Genes frequently altered in RCC					
VHL	M	Missense	c.266T>A	L89P	Tumor suppressor; mutated form induces stabilization of HIF-1α
PBRM1	M	Missense	c.2626G>A	R876C	Likely tumor suppressor; subunit of SWI/SNF chromatin remodeling complex
KDM5C	M	Splice site	n/a	n/a	Histone demethylase; associated with genomic rearrangement in tumors
KDM3A	P,M	Missense	c.634A>G	I212V	Histone demethylase; involved in HIF-1α activation
ARID4A	P	Missense	c.2335A>G	T799A	DNA binding protein; regulates several transcription factors (i.e., RB, AR, E2F)
P, primary tumor; M, metastatic tumor; WES, whole-exome sequencing; RCC, renal cell carcinoma.					

PI3K/Akt signaling or mTORC1 activation. Common between both the primary and metastatic tumor was the *EIF3C* missense mutation, suggesting a clonal mutation, occurring early in the evolution of the tumor.

In addition to mTOR pathway gene analysis, we also examined the sequencing data for genes commonly altered in RCC. In both the primary and metastatic tumors, we identified nonsynonymous alterations in *VHL*, *PBRM1*, *KDM3A*, *KDM5C*, and *ARID4A* (Table 1). The alterations identified in *VHL* and *PBRM1* have been previously characterized as inactivating, loss-of-function mutations in RCC [11]. *KDM3A*, *KDM5C*, and *ARID4A* are involved in chromatin remodeling and are frequently altered in RCC [12].

Discussion

Emerging data on the role of mTOR signaling revealed that mTOR-activating genetic alterations occur frequently in cancer, suggesting that these tumors would be sensitive to mTOR inhibitors. Specifically, several studies have identified activating mutations in the *MTOR* gene, inactivating mutations in *TSC1* or *TSC2*, and alterations in upstream effectors (i.e., PI3K and PTEN),

Table 2. mTOR pathway-associated genes included in analysis

PI3K/Akt signaling “upstream”	<i>PIK3CA, PIK3CB, PIK3CD, PIK3CG, PIK3C2, PIK3R1, PIK3R2, AKT1/2/3, PTEN, NF1, NF2</i>	mTOR signaling “upstream”	<i>MTOR, RPTOR, RICTOR, mLST8, FKBP12, DEPDC5, RHEB, TSC1, TSC2</i>	mTOR signaling “downstream”	<i>RPS6K1, EIF3 complex, EIF4 complex, SREBP, 4EBP, HIF1, AMPK, STK11</i>
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all support high-level mTOR activation [4–6]. In RCC, alterations in *MTOR*, *TSC1*, or *TSC2* have been reported in up to 10% of tumors, suggesting a potential link to mTOR inhibitor sensitivity. However, there is conflicting evidence in recent studies showing minimal association between mTOR pathway variants and response to mTOR inhibitor therapy [13, 14]. Evaluating cases where patients demonstrate an extraordinary response to everolimus has the potential to identify the molecular mechanisms of sensitivity and advance the development of a predictive biomarker for advanced RCC.

In this case study, WES analysis was performed on tumor DNA from an RCC patient with extraordinary response to everolimus. This analysis included focused evaluation of mTOR pathway genes frequently altered in cancer and previously reported to be associated with response to mTOR inhibition (Table 2). The variants identified in our analysis have not been reported to be associated with sensitivity to mTOR inhibitors; however, several of them have been shown to activate oncogenic mTOR signaling. Specifically, *PIK3R2* and *PIK3C2G* are frequently mutated in several malignancies where they have been shown to promote PTEN instability and constitutive PI3K/Akt activation [15, 16]. We also identified mutations in *INSR* and *PRKD3*, both regulating mTORC1 activity through PI3K/Akt signaling, and it is suggested that *PRKD3* genetic alterations can promote proliferation and survival signaling through prolonged Akt activation [17]. Finally, *EIF3B* expression has been linked to increased mTORC1 activation and was recently identified as a prognostic marker for adverse outcomes in ccRCC [10]. Importantly, the *EIF3B* variant was found in both the primary and metastatic tumors, suggesting a truncal alteration that occurred early in tumorigenesis.

Study of exceptional responders provides the opportunity to discover the molecular basis for sensitivity to anticancer therapies. Advances in DNA sequencing technology with ultrahigh-throughput next-generation sequencing (NGS) enables insight into the tumor-wide mutational landscape and has rapidly advanced the field of precision oncology. Employing the WES approach in super-responders facilitates the discovery of novel molecular mechanisms that can be used in developing a predictive marker for prospectively identifying patients who may similarly respond to the specific therapy. Commercially available NGS panels are often limited to known pathologic variants and those that predict response to targeted therapies. Importantly, most diagnostic and research NGS panels have a limited number of genes involved in mTOR signaling, therefore limiting applicability for discovery of a predictive marker for everolimus therapy. Through a WES approach, we identified several novel variants in mTOR pathway genes as potential molecular markers of everolimus sensitivity. Additional investigation is needed to characterize the functional significance of these alterations in everolimus sensitivity and apply these findings to future biomarker studies.

Conclusion

This case highlights the importance of molecular characterization in patients with exceptional response to targeted agents. WES has the capability to detect potential therapeutic targets not included in precision medicine panels that can be employed as a clinical tool in

guiding therapy selection. The discovery of novel variants in a patient with an exceptional response to everolimus provides the evidence to warrant further investigation as potential predictive markers.

Statement of Ethics

Written informed consent was obtained from the patient for the publication of this case report and any accompanying images.

Conflict of Interest Statement

The authors declare no conflicts of interest.

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Author Contributions

C.W. and E.B.-M. compiled clinical data and imaging studies. C.W. and O.H. prepared study material for WES analysis. N.G. and D.C. performed pathological analysis and data analysis. D.C., G.D., A.P., and C.H. performed data analysis and interpretation. A.P. was involved in manuscript, table, and figure preparation. C.H. and G.D. were involved in conception and study design. All authors read and approved the final version of this manuscript.

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