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N-cadherin regulates GluA1-mediated depressive-like behavior in adolescent female rat offspring following prenatal stress

Shuya Shao a, Dan Yao a, Senya Li a, Jing Li a, Yufang Si b, Huiping Zhang c, Zhongliang Zhu b, Dongli Song d, Hui Li a*

a Department of Neonatology, The First Affiliated Hospital of Xi’an Jiaotong University, Xi’an, Shaanxi, China
b Key Laboratory of Resource Biology and Biotechnology in Western China, Maternal and Infant Health Research Institute and Medical College, Northwestern University, Xi’an, Shaanxi, China
c The Affiliated Children Hospital of Xi’an Jiaotong University, Xi’an, Shaanxi, China
d Division of Neonatology, Department of Pediatrics, Santa Clara Valley Medical Center, San Jose, California.

Short Title: Adolescent female depression

Corresponding Author:
Hui Li
Department of Neonatology
The First Affiliated Hospital of Xi’an Jiaotong University
277 Yanta West Road
Xi’an, Shaanxi, 710061, China
Tel: 13909210705
E-mail: huili@xjtu.edu.cn

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Abstract

Background: The incidence of depression is twice higher in women than in men, and gender differences in the prevalence rates first emerge around puberty. Prenatal stress (PS) induces gender-dependent depressive-like behavior in adolescent offspring, but the neuro-physiological mechanisms remain unclear. Our study aimed to investigate the possible neuro-physiological mechanisms of gender-dependent depressive-like behavior in PS adolescent offspring and further explored the possibility of treating depression in adolescent female rats.

Methods: The pregnancy rats were exposed to restraint stress in the third trimester for 7 days. The depressive-like behavior and the expression of N-cadherin, AMPARs in the hippocampus of adolescent offspring rats were assessed. 10 mg/kg AMPARs antagonist CNQX and 10 mg/kg N-cadherin antagonist ADH-1 were intraperitoneally injected into female adolescent offspring, respectively. 0.2 µg AMPARs agonist CX546 was administered to the dentate gyrus of male adolescent offspring to determine the role of N-cadherin-AMPARs in depressive-like behavior of the offspring following PS.

Results: We found that PS increased N-cadherin expression, which upregulated GluA1 expression in the dentate gyrus, mediating depressive-like behavior in adolescent female rat offspring by reducing PSD-95. In addition, ADH-1 and CNQX improved depressive-like behavior in adolescent female offspring following PS. Furthermore, injection of the CX546 into the dentate gyrus induced depressive-like behavior in PS male offspring.

Conclusion: The gender-dependent expression of N-cadherin-GluA1 pathway in adolescent offspring in the dentate gyrus was the key factor in gender differences of depressive-like behavior following PS.

Introduction

Major depression disorder is the most common neuropsychiatric disease and currently affects 350 million people worldwide [1]. Epidemiological investigation revealed that the incidence of depression is twice higher in women than in men, and gender differences in the prevalence rates first emerge around puberty [1]. Studies have shown that the occurrence of adolescent depression in offspring is related to prenatal stress (PS) [2, 3], and the occurrence of adolescent depression is gender-specific [4]. In addition, sexually dimorphic responses to PS in offspring cause an increased prevalence of internalizing disorders, such as depression, among females [5]. Our previous study also reported that only adolescent female offspring who experienced PS showed depressive-like behavior or decreased learning and memory abilities, while males were resistant [6, 7]. Nevertheless, the neurophysiological mechanism of depressive-like behavior in adolescent female rats remains unknown.

Van et al. found that the altered expression of cell adhesion molecules and glutamate receptors in the hippocampus were involved in the gender-specific effects of PS on offspring through genome-wide sequencing analysis [8]. Cadherin is a transmembrane cell adhesion molecule that allows cells to tightly bind to each other [9, 10]. There are many types of cadherin, among which N-cadherin is distributed in neurons. N-cadherin plays an important role in nerve migration and localization during brain development [11] and is highly expressed in the hippocampus [12] where it regulates α-amino-3-hydroxy-5-methyl-4-isoxazole propionic acid receptors (AMPARs)-mediated synaptic plasticity. The AMPARs are important ionic glutamate receptors, consisting of four subunits, including GluA1, GluA2, GluA3 and GluA4. AMPARs are widely distributed in the central nervous system, including in the hippocampus [13]. When long-term potentiation (LTP) occurs in the hippocampus, the distribution of N-cadherin in the synaptic membrane increases [14], the synapse becomes more stable [15, 16], and AMPARs in the synaptic membrane are more stable [17]. In the case of long-term depression, N-cadherin is internalized from the synaptic membrane, and then, AMPARs are removed from the postsynaptic membrane [18]. The interaction among N-cadherin and GluA1, GluA2 promotes the growth of dendritic spines [19].
Although the role of N-cadherin in depression is not clear, emerging evidence has found that AMPARs-mediated synaptic plasticity is critical in the pathogenesis of stress-related depression [20-22]. PSD-95 is an important marker of synaptic plasticity [23] that is decreased in the hippocampus of male rats following stress [24]. Male mice susceptible to chronic stress exhibit depressive-like behavior correlated with down-regulated expression of GluA1 and GluA2, which up-regulates expression of postsynaptic PSD-95 [25]. However, these studies only included adult male subjects; it is unclear whether N-cadherin-AMPARs mediate depressive-like behaviors in adolescent female rats by regulating PSD-95.

Our previous study found that adolescent female offspring who experienced PS showed depressive-like behavior while males were resistant [6] and offspring exposed to PS exhibit gender-dependent alterations in dendrite length and fulcrum in the hippocampus [26]. In addition, some studies have investigated the effects of AMPARs on depressive-like behavior in the hippocampus after PS exposure, most of which only included male subjects [27-29]. However, whether N-cadherin regulates the AMPARs-dependent synaptic plasticity-mediated depressive-like behavior in adolescent female offspring following PS remains unclear. Therefore, our study aimed to investigate the role of N-cadherin-AMPARs pathway in mediating gender-dependent depressive-like behavior in PS adolescent offspring and further explored the possibility of treating depression in adolescent female rat offspring.

Materials and Methods

Animals

Sprague–Dawley rats were housed in an environment with controlled temperature (22 ± 1 °C), humidity (60%) and 12-h light/12-h dark cycles with free access to food and water. The lights were turned on at 8:00 am and turned off at 8:00 pm. Every effort was made to optimize comfort and to minimize the use of animals. All the experimental procedures were performed according to the institutional guidelines of the Animals Care and Use Committee of Xi’an Jiao tong University and were approved by Xi’an Jiao tong University Health Science Center.

Prenatal stress

Nulliparous female rats (250–270 g) were housed with male rats (280–350 g) for mating (3:1), and the vaginal smear was examined the following morning. The day on which the vaginal smear was positive was defined as day 0 of gestation. Each pregnant rat was subsequently housed individually. All the pregnant rats were randomly assigned to either the control group (n = 8) or the PS group (n = 8). The PS pregnant rats were subjected to restraint stress three times per day (at approximately 08:00–11:00, 12:00–15:00, and 16:00–19:00), for 45 min per session on gestational days 14–20 [30-32]. The pregnant dams returned into their home cages between each session of restraint and the rats did not have water and food available for the whole period of stress, which were free access to food and water for the rest of the time. The rats in the control group have water and food available for the whole period of investigation. The device was a transparent cylinder (6.8 cm in diameter) with an adjustable length to accommodate the size of the animals. Air holes in the cylinder were for breathing. The control group pregnant rats were undisturbed.

The birth of offspring was considered postnatal day 0 (PND 0). After birth, the offspring were left undisturbed with their mothers until weaning (21 days after birth). After weaning, male and female offspring were housed separately, each sex offspring from one litter were housed in a cage. For the first experiment, offspring rats were assigned to the CON (male, n = 13; female, n = 15) or PS (male, n = 16; female, n = 16) group for further behavioral experiments. Only litters of 6–12 pups were included in this study, and 5-week-old offspring (two pups per sex from each litter at most to avoid a ‘litter effect’) were used [6, 33].

Drugs

The AMPA receptor antagonist CNQX, the selective AMPARs agonist CX546 and the N-cadherin antagonist ADH-1 were obtained from Sigma-Aldrich. St. Louis, Missouri, USA. CNQX and ADH-1 were dissolved in 0.9% saline solution, CX546 was dissolved in 1% DMSO, 1% ethanol, and 98% saline. The
rats were intraperitoneally injected with CNQX or ADH-1 (10 mg/kg) or an equal volume of saline and were subjected to sucrose preference test 0.5 h after administration [34, 35]. The CX546 doses used were the same as previously reported [36], and drugs or the same volume of vector (1% DMSO, 1% ethanol, and 98% saline) were microinjected into the hippocampus 30 min before the sucrose preference test.

For the second experiment, female offspring rats were divided into 6 groups and 6 litters were used in each group: CON + saline (n = 6), PS + saline (n = 6), CON + CNQX (n = 6), PS + CNQX (n = 6), CON + ADH-1 (n = 6), PS + ADH-1 (n = 6), to avoid a 'litter effect', two female pups from each litter at most were used in 5-7-week-old. For the third experiment, male offspring rats were divided into 4 groups: CON + vector (n = 6), CON + CX546 (n = 6), PS + vector (n = 6), PS + CX546 (n = 6) and at most two male pups from each litter were used in 5-week-old.

Microinjection
A tube was implanted in the dentate gyrus of the hippocampus of rats 5 weeks after birth. Rats were anaesthetized with 10% ethyl carbamate (0.3–0.4 ml/100 g, i.p.), and their heads were fixed on a stereotaxis device (WPI, USA). According to the Paxinos and Watson rat brain stereotaxis map [37], a stainless steel casing with a core was inserted on each side of the dentate gyrus (AP: −3.5 mm, ML: ±2.1 mm; DV: −3.7 mm relative to bregma), and fixed with glass ionomer cement and dental powder. The rats were housed separately after the operation. After 7 days of recovery, microinjection into the dentate gyrus was performed using a single channel microinjection pump (KDS, USA). The CX546 (1 μl × 0.1 μg/μl, Sigma- Aldrich. St. Louis, Missouri, USA) was slowly injected into both sides of the dentate gyrus at a rate of 0.5 μl/min. The needle was kept in position for 5 min after injection to prevent drug efflux. According to different treatments, the rats were classified into three categories: CON + vector (n = 6), CON + CX546 (n = 6), PS + vector (n = 6), PS + CX546 (n = 6). The following experiments were performed 30 min after the operation.

Behavioral test
Sucrose Preference Test (SPT)
The Sucrose Preference Test was performed 36 days after birth. For the CX546 administration, it was performed 42 days after birth. All the rats were provided with 2% (w/v) sucrose solution and water at the same time for 24 h adaptation. At the same time, the total consumption of sucrose and water for 24 hours was recorded. The next day, the rats were subjected to 4 h of water deprivation before the behavioral test. To prevent the rats from habitually drinking from the water bottles, the daily placement of sucrose and drinking water was alternated. After deprivation, the consumption of sucrose and water of each group was recorded over 2 h. Three sucrose preference tests were performed for each rat. Sucrose preference was used as a measure of depressive-like behavior. Sucrose preference=Sucrose solution consumption/ (Drinking water consumption + Sucrose solution consumption) ×100%.

Forced Swim Test (FST)
The FST was performed 38 days after birth according to previous reports[38]. For the CX546 administration, it was performed 42 days after birth. Briefly, each rat was individually placed into a Perspex cylinder (diameter 18 cm, height 40 cm) filled with 25 °C water with a height of 30 cm for 15 min as pre-test under low light conditions. After 24 h later, the procedure was repeated, but on this occasion the duration that the rats remained immobile and the time of first immobility (latency immobility time) during a 5-min observation period was recorded. Rats spending more immobile time or less immobility latency time compared to the control group were characterized as increasing depressive-like behavior. The rats were sacrificed the day after FST.

Quantitative Real-Time PCR
The Quantitative Real-Time PCR was conducted according to previous reports [7]. Offspring rats anesthetized with intraperitoneal chloral hydrate (0.3–0.4 ml/100 g, i.p) were decapitated. The hippocampus tissues were dissected and frozen in liquid nitrogen and then kept in − 80 °C freezer immediately. Total RNA was extracted by RNA fast 200 RNA kit (Xianfeng Biotech, China), according
to the manufacturer’s instructions. The cDNA was synthesized using the PrimeScriptTM RT Master Mix kit (RR036A; Takara, Kyoto, Japan). Quantitative real-time PCR was performed with the SYBR® Premix Ex Taq™ II (Tli RNaseH Plus) (RR820l, TaKaRa) on Bio-Rad CFX96 system (Bio-Rad, Hercules, California, USA). The primer sequences were as Table 1. Every sample was performed in triplicates. Fold changes in the target gene relative to corresponding GAPDH endogenous control were calculated using the 2^−ΔΔCt method.

**Western Blot**

The Western Blot was conducted according to previous reports [7]. Total proteins were isolated from the hippocampus with RIPA buffer (Solarbio, China). The protein sample was separated with 10% SDS-PAGE, and then transferred to a PVDF film (Millipore, USA). After blocking with 5% skim milk at room temperature for 2 h, primary antibodies against GluA1 (1:5000, ab31232), GluA2 (1:5000, ab20673), GluA3 (1:5000, ab40845), GluA4 (1:5000, ab115322), N-cadherin (1:5000, ab18203), PSD-95(1:5000, ab18258), and β-actin (1:2000, ab8227) obtained from Abcam (USA) were added and incubated at 4 °C overnight. After washing, goat anti-rabbit secondary antibody (1:5000, Pioneer, China) was added and incubated at room temperature for 2 h. Then, the film was developed with ECL chemiluminescence (Millipore, China) and analyzed by imaging system (Bio-Rad, USA). The gray values of the target protein bands were scanned by Quantity One software. Results were standardized to β-actin.

**Immunofluorescence**

After the behavioral test, the rats were euthanized and transcardially perfused with 0.9% NaCl. Half of the brain was removed and fixed in 4% paraformaldehyde for 24 h at 4°C and immersed in 15%, 20%, and 30% sucrose concentrations overnight. Serial sections (10 μm/section) were cut through the entire anteroposterior extension of the hippocampus using a freezing microtome. The sections were permeabilized in 0.3% Triton X-100 for 30 min and then blocked in 10% normal goat serum for 1 h at 37°C before overnight incubation in primary antibodies against N-cadherin (PA5-19486, Invitrogen) and GluA1 (#13185, CST). Subsequently, the samples were incubated in secondary antibodies for an additional 2 h for fluorescence detection and covered with a coverslip. Images were acquired using a fluorescence microscope (Nikon Eclipse Ti).

**Statistical Analysis**

SPSS PASW Statistics v22.0 was used for data analysis. All data was represented as mean± standard error of measurement (S.E.M). For data with variance homogeneity and normal distribution, the two-way ANOVA was performed. After the influencing factor was determined, the one-way ANOVA stratified analysis was used. Mean differences with more than two groups were compared using one-way ANOVA, followed by Fisher’s LSD test to multiple comparisons if the main effect was significant at p < 0.05. A P < 0.05 was considered statistically significant.

**Results**

**Adolescent female offspring exhibited depressive-like behavior following PS**

After exposure to 7 days of restraint stress during the third trimester of pregnancy, the depressive-like behavior of the offspring was tested 5-6 weeks after birth [39] (shown in Fig. 1A). Based on the two-way ANOVA, the overall interaction (sex × PS) for sucrose consumption was observed (F(1,57)=8.652, p=0.005). The results showed that the sucrose consumption in the females of PS group decreased 23.80% compared to that in the CON group (F(1,30)=6.901, p=0.013). However, there was no significant increase in male offspring in sucrose consumption (shown in Fig. 1C). Both sex and PS had no effect on total water consumption of the offspring rats (shown in Fig. 1B). In the CON group, gender had no effect on sucrose consumption.

In FST, the overall interaction (sex × PS) was observed (F(1,57)=9.288, p=0.003) for the immobility time (F(1,57)=9.288, p=0.003) and the latent immobility time (F(1,57)=11.769, p =0.001). The immobility time of the females in the PS group, but not the males, was significantly longer compared to that in the CON group (F(1,30)=18.052, p<0.001, shown in Fig. 1E). In addition, the latent immobility time was reduced in the PS female offspring (F(1,30)=26.255, p<0.001) but not in the PS male offspring (shown
in Fig. 1F). In the CON group, gender did not affect the immobility time and latent immobility time in FST. Moreover, PS had no effect on baseline activity (shown in Fig. 1D). In general, adolescent female offspring exhibited depressive-like behavior following PS, while male offspring were resistant.

**PS upregulated N-cadherin, GluA1 in the hippocampus**

To explore the impacts of PS on the expression of N-cadherin in adolescent offspring, we performed an analysis of N-cadherin expression in the hippocampus (shown in Fig. 2A). Based on the two-way ANOVA, we observed the overall interaction (sex × PS) for both the relative expression of N-cadherin mRNA (*F*(_1,20_)=13.512, *p*=0.001) and protein (*F*(_1,20_)=4.638, *p*=0.044). The one-way ANOVA stratified analysis showed that PS increased N-cadherin expression in the hippocampus of the female offspring (N-cadherin mRNA, *F*(_1,10_)=22.389, *p*=0.001; N-cadherin protein, *F*(_1,10_)=6.643, *p*=0.028, shown in Fig. 2B, E). However, there was no significant change in expression of N-cadherin in the male offspring. This finding suggested that N-cadherin probably mediated a gender-specific behavioral phenotype in adolescent offspring after PS.

With respect to the N-cadherin-regulated expression of AMPARs, we next explored whether AMPARs isoymes in the hippocampus are required for depressive-like behavior following PS. Based on the two-way ANOVA, the overall interaction (sex × PS) was observed for both the relative expression of GluA1 mRNA (*F*(_1,20_)=12.583, *p*=0.002) and protein (*F*(_1,20_)=7.236, *p*=0.014), as well as the relative expression of GluA2 mRNA (*F*(_1,20_)=15.051, *p*=0.001) and protein (*F*(_1,20_)=5.974, *p*=0.024). The one-way ANOVA stratified analysis showed that the PS female offspring exhibited increased GluA1 and GluA2 mRNA expression (GluA1, *F*(_1,10_)=30.484, *p*<0.001; GluA2, *F*(_1,10_)=19.419, *p*=0.001, shown in Fig. 2C-D) and protein expression (GluA1, *F*(_1,10_)=5.039, *p*=0.049; GluA2, *F*(_1,10_)=5.951, *p*=0.035, shown in Fig. 2F-G) in the hippocampus compared to the CON females, while no significant differences were observed in the PS male offspring for the four tested isoforms. PS did not exert significant effects on either the GluA3 or GluA4 expression in the hippocampus (Supplement 1).

We also found that PS increased N-cadherin and GluA1 expression in the entire hippocampus of the female offspring by immunofluorescence (Supplement 2). Furthermore, the upregulation of N-cadherin and GluA1 in PS group was more obvious in dentate gyrus and N-cadherin and GluA1 were co-localized (*F*(_1,10_)=8.163, *p*=0.046; *F*(_1,10_)=23.352, *p*=0.008, shown in Fig. 2H). Therefore, these results suggested that the depressive-like behavior in female adolescent offspring exposed to PS may be mainly caused by the increased expression of N-cadherin-GluA1 pathway in the dentate gyrus.

**AMPA receptors antagonist CNQX improved depressive-like behavior in adolescent female offspring after PS**

To confirm that GluA1 is required for the depressive-like behavior in adolescent female offspring exposed to PS conditions, we injected the AMPARs antagonist CNQX into PS female offspring 30 min before behavioral testing (shown in Fig. 3A). Based on the two-way ANOVA, the overall interaction (CNQX × PS) for sucrose consumption was observed (*F*(_1,40_)=6.962, *p*=0.012), as well as the effect of PS (*F*(_1,40_)=7.721, *p*=0.009). The CNQX administration had no effect on the behavior of offspring in the control group. However, we found that CNQX significantly improved the depressive-like behavior of adolescent female offspring after PS, as evidenced by the fact that the sucrose consumption of the females in the PS group significantly increased 78.55% after injection of CNQX (*F*(_1,24_)=36.839, *p*<0.001, shown in Fig. 3C). Both CNQX and PS had no effect on the total water consumption (shown in Fig. 3B). Furthermore, the overall interaction (CNQX × PS) for both immobility time and latent immobility time were observed (*F*(_1,40_)=15.122, *p*<0.001; *F*(_1,40_)=5.379, *p*=0.026), as well as the effect of PS (*F*(_1,40_)=40.820, *p*<0.001; *F*(_1,40_)=4.735, *p*=0.036). The one-way ANOVA stratified analysis showed that the immobility time decreased 65.55% (*F*(_1,24_)=40.656, *p*<0.001, shown in Fig. 3E), while latent immobility time increased 207.65% (*F*(_1,24_)=13.067, *p*=0.001, shown in Fig. 3F) in the PS female offspring after CNQX injection.

We also found the interaction of CNQX and PS on the relative mRNA and protein expression of GluA1 (*F*(_1,20_)=4.886, *p*=0.039; *F*(_1,20_)=4.965, *p*=0.037), as well as the effect of PS (*F*(_1,20_)=103.787, *p*<0.001; *F*(_1,20_)=21.284, *p*<0.001) and CNQX administration (*F*(_1,20_)=229.398, *p*<0.001; *F*(_1,20_)=68.317,
p<0.001). CNQX administration reduced GluA1 expression in the hippocampus of CON group (GluA1 mRNA, F(1,10)=65.223, p<0.001; GluA1 protein, t =2.617, df=10, p =0.026, shown in Fig. 4A-B). In addition, PS significantly increased GluA1 in the hippocampus (GluA1 mRNA, F(1,10)=51.615, p<0.001; GluA1 protein, F(1,10)=67.070, p<0.001, shown in Fig. 4A-B), which was rescued by CNQX administration (GluA1 mRNA, F(1,10)=209.991, p<0.001; GluA1 protein, F(1,10)=79.247, p<0.001, shown in Fig. 4A-B). However, CNQX had no effect on N-cadherin expression in the hippocampus of CON and PS group (shown in Fig. 4C-D). This further demonstrated that GluA1 in the hippocampus was required for depressive-like behavior in female offspring exposed to PS, but altered expression of GluA1 in the hippocampus had no effect on N-cadherin.

N-cadherin antagonist ADH-1 improved the depressive-like behavior of adolescent female offspring by regulating the levels of GluA1 after PS
To confirm that N-cadherin regulates the depressive-like behavior of adolescent female offspring under PS conditions, we injected the N-cadherin antagonist ADH-1 into PS female offspring 30 min before behavioral testing (shown in Fig. 5A). Based on the two-way ANOVA, the overall interaction (ADH-1 × PS) for sucrose consumption was observed (F(1,35)=10.002, p=0.003), as well as the effect of PS (F(1,35)=3.456, p=0.037). We found PS induced depressive-like behavior in female offspring (shown in Fig. 5C, E, and F). In addition, the ADH-1 significantly improved depressive-like behavior in the adolescent female offspring after PS while had no effect on the behavior of offspring in the control group. After injecting ADH-1, the sucrose consumption by the females offspring in the PS group increased 55.01% (F(1,17)=15.517, p=0.001, shown in Fig. 5C). Furthermore, the overall interaction (ADH-1 × PS) for both immobility time and latent immobility time were observed (F(1,35)=16.457, p<0.001; F(1,35)=7.979, p=0.008), as well as the effect of PS (F(1,35)=24.919, p<0.001; F(1,35)=10.582, p=0.003). The one-way ANOVA stratified analysis showed that the immobility time decreased 68.15% (F(1,17)=26.842, p<0.001, shown in Fig. 5E), and the latent immobility time increased 171.07% (F(1,17)=54.780, p<0.001, shown in Fig. 5F) in the PS female offspring after ADH-1 injection.

We analyzed whether intraperitoneal ADH-1 improved depressive-like behavior in female offspring after PS by regulating GluA1. Based on the two-way ANOVA, the interaction (ADH-1 × PS) on the relative mRNA and protein expression of N-Cadherin (F(1,20)=5.665, p=0.027; F(1,20)=5.559, p=0.029) were observed, as well as the effect of PS(F(1,20)=51.228, p<0.001; F(1,20)=4.762, p=0.041) and ADH-1 administration(F(1,20)=20.454, p<0.001; F(1,20)=47.937, p<0.001). We found that ADH-1 administration reduced N-Cadherin expression in the hippocampus of CON group (N-Cadherin mRNA, F(1,20)=10.996, p=0.008; N-Cadherin protein, F(1,20)=12.627, p=0.005, shown in Fig. 6A-B). Moreover, ADH-1 rescued the up-regulated N-Cadherin expression in hippocampus induced by PS (N-Cad mRNA, F(1,20)=13.300, p=0.004; N-Cad protein, F(1,20)=36.672, p<0.001; shown in Fig. 6A, B).

The interaction (ADH-1 × PS) on the relative mRNA and protein expression of GluA1 (F(1,20)=6.987, p=0.016; F(1,20)=4.867, p=0.039) were also observed, as well as the effect of PS(F(1,20)=97.783, p<0.001; F(1,20)=19.100, p<0.001) and ADH-1 administration(F(1,20)=39.116, p<0.001; F(1,20)=31.484, p<0.001). ADH-1 also reduced GluA1 expression in hippocampus of female offspring in both CON (GluA1 mRNA, F(1,20)=11.820, p=0.006; GluA1 protein, F(1,20)=7.892, p=0.018) and PS group (GluA1 mRNA, F(1,20)=27.328, p<0.001; GluA1 protein, F(1,20)=24.144, p<0.001, shown in Fig. 6C, D). This indicated that the increase of GluA1 may be due to the increase of N-Cadherin caused by PS, which mediated depressive-like behavior in adolescent female offspring.

Hippocampal postsynaptic density mediates the depressive-like behavior of female offspring following PS
To elucidate the mechanism of GluA1-mediated the depressive-like behavior of female offspring following PS, we performed an analysis of PSD-95 expression in the hippocampus. Based on the two-way ANOVA, the interaction (sex × PS) on the relative mRNA and protein expression of PSD-95 (F(1,20)=7.375, p=0.013; F(1,20)=4.488, p=0.047) were observed, as well as the effect of PS(F(1,20)=7.832, p=0.011; F(1,20)=7.560, p=0.012). We found that PS induced depressive-like behavior in female offspring (shown in Fig. 2) by down-regulating PSD-95 (PSD-95 mRNA, F(1,20)=30.948, p<0.001; PSD-95 protein, F(1,20)=86.248, p<0.001). However, N-cadherin antagonist ADH-1 did not rescue the effect of PS on the PSD-95 expression.
adolescent offspring is a key factor in mediating gender differences in depressive response to PS.

Altered in the dentate gyrus of male offspring, and they did not exhibit depressive PS only increased in AMPARs.

Although the role of AMPARs in depression has been mentioned before, pregnancy, while male offspring were resistant, which is consistent with clinical research. The differential expression of N-cadherin in the dentate gyrus of female offspring, which in turn increased GluA1, mediating depressive-like behavior. However, expression of N-cadherin was not altered in the dentate gyrus of male offspring, and they did not exhibit depressive-like behavior in response to PS. The differential expression of N-cadherin-GluA1 in the dentate gyrus induced by PS in adolescent offspring is a key factor in mediating gender differences in depressive-like behavior.

**Discussion/Conclusion**

Numerous studies have shown that PS leads to depression in adolescent offspring [2, 3] with gender specificity [4]. Female offspring are more likely to develop emotional disorders, and sexually dimorphic responses to PS in offspring are one reason for the increased prevalence of depression among females [5]. In this study, we found that only female offspring showed depressive-like behavior during adolescence when the dams were exposed to restraint stress during the late stage of pregnancy, while male offspring were resistant, which is consistent with clinical research [5]. Although the role of AMPARs in depression has been mentioned before [40, 41], gender differences in AMPARs-mediated depressive-like behavior following PS are unclear. In this study, we found that PS only increased the expression of N-cadherin in the dentate gyrus of female offspring, which in turn increased GluA1, mediating depressive-like behavior. However, expression of N-cadherin was not altered in the dentate gyrus of male offspring, and they did not exhibit depressive-like behavior in response to PS. The differential expression of N-cadherin-GluA1 in the dentate gyrus induced by PS in adolescent offspring is a key factor in mediating gender differences in depressive-like behavior.
There are different explanations for gender differences in depression. First, Salk et al. found that gender differences in depression mainly emerge during adolescence, while there is no gender difference or even a somewhat higher prevalence of depression among boys than girls in childhood. In addition, observations in adulthood are inconsistent with the gender difference in depression [1]. Furthermore, stress duration may mediate gender differences in depression. Studies have proved that females develop depressive-like behavior following shorter stress duration; on the other hand, males require longer durations of stress to develop depressive-like behavior [42-44]. In addition, Muir et al. found that after 4 days of chronic variable stress, females developed depressive-like behaviors, while it takes 21 days for males to develop these behaviors [45]. Finally, differences in female and male sex hormones are also important for gender differences with respect to depression. Women are more likely to suffer from depression during periods of marked hormonal fluctuations, including adolescence, postpartum and perimenopause periods [46]. Rodent studies have also shown that ovariectomized female rats exhibit depressive-like behavior [47, 48].

In our study, depressive-like behavior in adolescent female offspring might be due to prenatal restraint stress for 7 days, while this stress duration did not induce depressive-like behavior in male offspring. Moreover, estrogen fluctuations in adolescent female offspring may alter N-cadherin expression in the dentate gyrus, which mediated GluA1-dependent depressive-like behavior in female offspring.

In our previous findings, PS decreased the expression of GluA1-3 in the hippocampus and induced depressive-like behavior in male offspring [32], which is inconsistent with the results of this study. The possible reasons are as follows. First, our previous study only focused on male offspring, while depressive-like behavior and AMPARs expression in female offspring were not evaluated. Second, adolescent offspring were observed in the present study, while previous results occurred in childhood (21 days after birth) in male offspring. In addition, Miranda et al. found that GluA1 expression decreased and GluA2 expression increased in the dentate gyrus of female adult offspring following PS [49]. This difference may also be due to the different ages of the offspring.

The development of the rat hippocampus mainly occurs during the third trimester, which is also an active period of accelerated development of synapse formation. Therefore, stress during late pregnancy is more likely to cause an imbalance in hippocampal functions, leading to depression [50]. This is one of the important reasons that we selected the hippocampus for analysis. In this study, we also found that the effects of PS on N-cadherin and GluA1 mainly occurred in the dentate gyrus, and N-cadherin and GluA1 were co-localized. These findings suggested that N-cadherin regulated GluA1 in the dentate gyrus, mediating depressive-like behavior in adolescent female offspring following PS. Studies have confirmed that AMPARs regulates synaptic plasticity in the hippocampus. Our previous study shows that the dendrites at the top of pyramidal neurons were shorter, the dendrite fulcrum was decreased, and expression of syn-1 was decreased in the hippocampus of the offspring exposed to PS [9]. This suggests that the altered synaptic plasticity in the hippocampus might be important for depressive-like behavior following PS. PSD-95 is critical for AMPARs-dependent synaptic plasticity [51]. We analyzed its expression in the hippocampus of female offspring with depressive-like behavior. We found that the expression of PSD-95 was decreased in the hippocampus of PS female offspring with depressive-like behavior. Furthermore, systemic administration of AMPARs antagonists rescued the depressive-like behavior and PSD-95 expression. This further confirmed that PS caused the up-regulation of GluA1 expression in the dentate gyrus of female offspring, affecting synaptic plasticity by reducing PSD-95, which mediated the depressive-like behavior in adolescent female offspring.

ADH-1 is a cyclic pentapeptide that disrupts N-cadherin interactions and has been shown to inhibit cell growth and tumor progression both in vitro and in vivo [52]. In our study, we found an antidepressant effect of ADH-1, wherein intraperitoneal ADH-1 resulted in a significant reduction in N-cadherin and GluA1 in the dentate gyrus, improving depressive-like behavior in female offspring.
induced by PS. Notably, this is the first study to target N-cadherin as a potential therapeutic modality in depression.

The present study has limitations. First, direct evidence that up-regulated N-cadherin-GluA1 in the dentate gyrus induced depressive-like behavior in PS adolescent female rat offspring is lacking. Second, we did not examine the AMPARs and N-cadherin expressions in the control group treated with CNQX and ADH-1. Third, to conclude that “Hippocampal postsynaptic density mediates the depressive-like behavior of female offspring following PS” we should analyze the levels of PSD-95 not only in total lysates but also in PSD-enriched preparations.

In conclusion, our study found that the high incidence rate of depression in adolescent women probably results from the susceptibility of female offspring to PS. We further elucidated that PS increased expression of N-cadherin, which up-regulated GluA1 expression in the dentate gyrus, mediating depressive-like behavior in adolescent female offspring by reducing PSD-95. We also found that ADH-1, an antagonist of N-cadherin, exerted potential antidepressant effects. This study provides insight for elucidating the underlying mechanisms of adolescent female depression and antidepressant drugs.

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Statement of Ethics
All studies involving animals have been approved by the Institutional Animal Care and Use Committee of Xi'an Jiaotong University (No.2018012) and follow the ARRIVE guidelines.

Conflict of Interest Statement
The authors have no conflicts of interest to declare.

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Author Contributions
Shao Shuya propose the idea of the research. Shuya Shao, Hui Li, and Zhongliang Zhu designed the study. Shuya Shao performed behavioral and molecular biological experiments, analyzed data, and wrote and revised the manuscript, during which Yufang Si helped a lot. Dan Yao, Senya Li and Jing Li managed some behavioral experiments. Huiping Zhang undertook some PS model experiments. Corresponding authors Hui Li also supervised the study, acquired the funding, and revised the manuscript.

Data Availability Statement:
All data generated or analysed during this study are included in this article and its supplementary material files. Further enquiries can be directed to the corresponding author.
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Figure legends

**Fig. 1. Effects of PS on the behavior of rat offspring.** The SPT and FST were used to study depressive-like behavior of offspring rats. (A) Study design of PS model establishment and behavioral test. (B) 24-h water consumption in the SPT. (C) Sucrose preference. (D) Total activity in the FST. (E) Immobility time in the FST. (F) Latent immobility time in the FST. *P < 0.05 vs. CON-female. ***P < 0.001 vs. CON-female.

**Fig. 2. Effects of PS on the expression of N-cadherin, GluA1-2 in rat offspring.** (A) Experimental protocol. (B-D) Relative mRNA expression of N-cadherin, GluA1-2 in the hippocampus. (E-G) Protein expression of N-cadherin, GluA1-2 in the hippocampus. (H) Representative images of immunofluorescence staining showing N-cadherin and GluA1 in the dentate gyrus. N = 6 rats/group, and the number of experiments = 3. Magnification: 40X. Scale bar represents 50 μm. (I) Immunoﬂuorescence intensity analysis. *P < 0.05 vs. CON-female, ***P < 0.01 vs. CON-female,****P < 0.001 vs. CON-female.

**Fig. 3. Effects of the AMPA receptor antagonist CNQX on the behavior of female rat offspring.** (A) Experimental protocol. (B) Study design of PS model establishment, CNQX administration and behavioral test. (C) Sucrose
preference. (D) Total activity in the forced swim test. (E) Immobility time in the forced swim test. (F) Latent immobility time in the forced swim test. **P < 0.01 vs. CON + saline, ***P < 0.001 vs. PS + saline.

Fig. 4. Effects of the AMPARs antagonist CNQX on N-cadherin expression in female rat offspring. (A-B) Relative mRNA expression and protein expression of GluA1-2 in the hippocampus. (C-D) Relative mRNA expression and protein expression of N-cadherin in the hippocampus. **P < 0.01 vs. CON + saline, ***P < 0.001 vs. CON + saline, ****P < 0.001 vs. PS + saline.

Fig. 5. Effects of the N-cadherin antagonist ADH-1 on the behavior of female rat offspring. (A) Study design of PS model establishment, ADH-1 administration and behavioral test. (B) 24-h water consumption in the sucrose preference test. (C) Sucrose preference. (D) Total activity in the forced swim test. (E) Immobility time in the forced swim test. ***P < 0.001 vs. CON + saline, **P < 0.01 vs. PS + saline.

Fig. 6. Effects of the N-cadherin antagonist ADH-1 on GluA1 and GluA2 expression of female rat offspring. (A) Relative mRNA expression of N-cadherin in the hippocampus. (B) Protein expression of N-cadherin in the hippocampus. (C-D) Relative mRNA expression of GluA1-2 in the hippocampus. (E) Protein expression of GluA1-2 in the hippocampus. *P < 0.05 vs. CON + saline, **P < 0.01 vs. CON + saline, ***P < 0.001 vs. CON + saline, ****P < 0.001 vs. PS + saline.

Fig. 7. The effects of PS, CNQX and ADH-1 on PSD-95 expression. (A-B) Effects of PS on PSD-95 expression. (C-D) Effects of CNQX on PSD-95 expression. (E-F) Effects of ADH-1 on PSD-95 expression. *P < 0.05 vs. CON-female, **P < 0.01 vs. CON + saline, ***P < 0.001 vs. CON + saline, &P < 0.01 vs. PS + saline.

Fig. 8. Effects of CX546 administration in the dentate gyrus on the expression of GluA1. (A) Study design of PS model establishment, CX546 administration and behavioral test. (B) Location of CX546 administration in the hippocampus. (C) Protein expression of GluA1 in the hippocampus. *P < 0.05 vs. CON + saline, ****P < 0.001 vs. PS + saline.

Fig. 9. Effects of CX546 administration in the hippocampus dentate gyrus on the expression of GluA1 and the behavior. (A) Representative images of immunofluorescence staining showing GluA1 in the dentate gyrus. N = 6–7 mice/group, and the number of experiments = 3. Magnification 40X. Scale bar represents 50 μm. (B) Immunofluorescence intensity analysis. (C) 24-h water consumption in the sucrose preference test. (D) Sucrose preference. (E) Total activity in the forced swim test. (F) Immobility time in the forced swim test. (G) Latent immobility time in the forced swim test. **P < 0.01 vs. PS + saline, ***P < 0.001 vs. PS + saline.
A

Pregnancy

Offspring

30 min before behavioral test

Sacrifice

Prenatal stress

FST

SPT

Administration

B

24h water consumption (ml)

CON+saline
CON+CNQX
PS+saline
PS+CNQX

C

Sucrose preference (%)

CON+saline
CON+CNQX
PS+saline
PS+CNQX

D

Total activity (m)

CON+saline
CON+CNQX
PS+saline
PS+CNQX

E

Immobile time (s)

CON+saline
CON+CNQX
PS+saline
PS+CNQX

F

Latency immobility time (s)

CON+saline
CON+CNQX
PS+saline
PS+CNQX