Voxel-Based Morphometry in Tau-Positive and Tau-Negative Frontotemporal Lobar Degenerations

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Key Words
Tau \cdot Frontotemporal lobar degeneration \cdot Voxel-based morphometry \cdot Magnetic resonance imaging

Abstract

Background: The identification of specific, diagnostically useful predictors of protein dysfunction in the frontotemporal lobar degenerations (FTLD) is a problem of great clinical and biological interest. Correlations between regional patterns of tissue loss and specific proteinopathies have not been established. Objective: Specific brain imaging correlates of protein tau dysfunction were sought using voxel-based morphometry in FTLD subgroups with and without tau pathology. Methods: Seventeen patients with pathologically or genetically confirmed diagnoses of FTLD who had undergone volumetric brain magnetic resonance imaging (MRI) were identified retrospectively and tau-positive (n = 9) and tau-negative (n = 8) subgroups were defined. MRI data were compared with healthy age- and sex-matched controls using voxel-based morphometry implemented in a statistical parametric mapping software package. Results: Compared with controls, tau-positive and tau-negative subgroups had extensive common areas of regional brain atrophy predominantly affecting the frontal and anterior temporal lobes. No specific brain imaging features were identified for either subgroup. Conclusion: Patterns of frontotemporal atrophy do not predict the presence or absence of tau pathology; conversely, different immuno-histochemical profiles are associated with similar patterns of regional vulnerability to neuronal loss in FTLD.

As our understanding of the molecular basis of the degenerative dementias improves, the correlation of specific patterns of tissue loss with disordered protein function in these diseases holds considerable clinical and biological interest [1–5]. The advent of volumetric brain magnetic resonance imaging (MRI) has enabled the identification of distinctive patterns of regional atrophy in a number of clinical dementia syndromes and such patterns can reliably distinguish Alzheimer from non-Alzheimer dementias [6–11]. However, the substantial clinical and genetic heterogeneity within the non-Alzheimer group has so far precluded the identification of specific radiological correlates of protein dysfunction in these diseases [1–3, 11]. This problem is particularly relevant in the large...
group of frontotemporal lobar degenerations (FTLD), in which attempts to identify specific clinical and radiological phenotypes of protein pathophysiology have been largely unsuccessful [1–3]. Here we describe a detailed analysis of MRI features in a population of individuals with pathologically and genetically defined FTLD: we sought to identify specific patterns of regional brain atrophy in individuals with and without tau pathology. Tau is a microtubule-associated protein that has been implicated in the pathogenesis of many sporadic and hereditary neurodegenerative diseases and a number of mutations in the tau gene on chromosome 17 have been identified [1–5]. The tauopathies are a clinically, radiologically and pathologically diverse group of diseases. Although postmortem studies have not established macroscopic patterns of brain involvement that reliably distinguish diseases with and without tau pathology [12, 13], pathological case series are likely collectively to represent a more advanced stage of disease; brain imaging during life might detect specific macroscopic correlates of tau dysfunction at an earlier stage of disease. In the present study, the MRI phenotypes of tau-positive and tau-negative patients were compared using voxel-based morphometry: an unbiased statistical methodology that is emerging as a promising tool in the identification of characteristic patterns of brain atrophy in a number of the degenerative dementias [10, 14].

Methods

Subjects

Seventeen patients with pathologically or genetically confirmed diagnoses of FTLD who had undergone volumetric brain MRI were identified retrospectively from referrals to the Dementia Research Group at the Institute of Neurology, London. Patients were diagnosed clinically as having frontotemporal dementia (13 cases), semantic dementia (3 cases) or progressive non-fluent aphasia (PNFA; 1 case) according to current consensus criteria [15].

Pathological and Genetic Data

The pathological diagnosis was established either at postmortem or on antemortem brain biopsy; 1 patient had an identified tau mutation and a typical clinical presentation without pathological confirmation. After formalin fixation and paraffin embedding, tissue blocks were examined using routine techniques including haematoxylin and eosin, Luxol fast blue, cresyl violet and modified Bielschowsky’s silver stains. All specimens were immunostained with antibodies against tau (tau 12E8; Elan Pharmaceuticals) and ubiquitin (Dako, Carpinteria, Calif., USA). The pathological diagnosis was based on consensus criteria [16]. In patients with tau pathology and an autosomal dominant pedigree, the tau gene was sequenced and screened for known mutations to identify cases with frontotemporal dementia-parkinsonism linked to chromosome 17 (FTDP-17).

Two pathological subgroups were defined: patients with tau inclusions (4 cases of FTDP-17, 5 cases of sporadic Pick’s disease) constituted the ‘tau-positive’ subgroup; patients with ubiquitin-positive, tau-negative inclusions (8 cases) constituted the ‘tau-negative’ subgroup. Characteristics of the tau-positive and tau-negative subgroups are summarized in table 1. Overall, both subgroups were well-matched with respect to disease duration and clinical severity (as indexed by Mini-Mental State Examination score and Clinical Dementia Rating scale); the mean age of patients in the tau-negative subgroup was 10 years older than in the tau-positive subgroup.

MRI Acquisition

In all patients and in 20 healthy age- and sex-matched controls, T1-weighted volumetric MR scans were acquired using a 1.5-tesla Signa unit (General Electric Medical Systems, Milwaukee, Wisc., USA) with a spoiled gradient-echo technique. Imaging parameters were as follows: time to echo 5 ms, time to repeat 35 ms, flip angle 35°, and field of view 24 × 24 × 19.2 cm. This sequence yielded 124 contiguous 1.5-mm-thick coronal slices.

Image Analysis

Distributions of brain atrophy in the tau-positive and tau-negative groups were compared with controls using statistical parametric mapping software SPM99 (Wellcome Department of Imaging Neuroscience, London, UK; http://www.fil.ion.ucl.ac.uk/spm) under MATLAB® (Mathworks, Sherborn, Mass., USA); the various processing steps have been described in detail previously [10]. All images were first normalized to a customized template. This customized template was based on a mean brain image derived from a group of 10 normal controls and 10 patients with a neurodegenerative disease (5 Alzheimer’s disease, 5 FTLD), all age- and gender-matched to the study cohort and acquired on the same MR scanner; the mean image was normalized to a standard stereotactic space defined by the Montreal Neurological Institute (MNI) standard brain [17] and spatially smoothed using an isotropic Gaussian kernel of 8 mm full width at half maximum (FWHM). Each subject’s normalized brain image was segmented into grey matter, white matter and cerebrospinal fluid and each grey matter image was masked to exclude all non-brain voxels with a brain region performed using a semi-automated, iterative 3-dimensional morphologic technique implemented in MIDAS image analysis software [18]. Masked grey matter images were modulated

Table 1. Characteristics of tau-positive and tau-negative subgroups in this study

<table>
<thead>
<tr>
<th></th>
<th>Age years</th>
<th>Duration of symptoms years</th>
<th>MMSE score</th>
<th>CDR score</th>
</tr>
</thead>
<tbody>
<tr>
<td>Tau-positive (n = 9)</td>
<td>52 (8.7)</td>
<td>4 (2.1)</td>
<td>21 (4.5)</td>
<td>1.4 (0.2)</td>
</tr>
<tr>
<td>Tau-negative (n = 8)</td>
<td>62 (6.8)</td>
<td>4 (2.3)</td>
<td>22 (5.5)</td>
<td>1.2 (0.5)</td>
</tr>
</tbody>
</table>

Values refer to mean and standard deviation in parentheses at time of brain MRI. CDR = Clinical Dementia Rating scale; MMSE = Mini-Mental State Examination score.
Fig. 1. Group statistical parametric maps (SPMs) of significant volume loss in tau-positive and tau-negative subgroups compared with age-matched healthy controls. SPMs have been rendered on coronal and axial sections of the customized template MR brain volume (see Methods) and thresholded at an uncorrected voxel significance level of $p < 0.0001$ for display purposes; the Z score at each voxel is coded according to the colour scale (right). In both tau-positive and tau-negative cases, atrophy involves extensive bilateral frontal and anterior temporal areas. No regions are involved specifically in the tau-positive or in the tau-negative subgroups, although there are qualitative differences in the profile of temporal lobe atrophy between the two groups (see text).

and smoothed with an isotropic Gaussian kernel of 8 mm FWHM [19]. Regionally specific differences in grey matter volume between the tau-positive and tau-negative subgroups and between each pathological subgroup and controls were assessed using a single subject condition and covariate model implemented in statistical parametric mapping (SPM99) software (http://www.fil.ion.ucl.ac.uk/spm); age and gender were included in the model as nuisance variables. Volume changes in the tau-positive and tau-negative subgroups relative to the control group and between the two pathological subgroups were assessed at each brain voxel by estimating the t score at a significance threshold of $p < 0.05$ corrected for multiple comparisons across the entire brain volume according to Gaussian random field theory [19].

In every subject, the whole-brain volume and total intracranial volume (TIV) were estimated from the raw T1-weighted MRI volume using MIDAS image analysis software as previously described [18, 20]. To estimate whole-brain volume, every axial slice between the lowest point of the cerebellum and the superior point of the cortex was measured using a fixed CSF-brain intensity threshold (60% of mean brain signal intensity). To estimate TIV, each volume was registered to the standard MNI template and every tenth axial slice was measured using a fixed threshold (30% of mean brain signal intensity) to outline the outer border of dura with linear interpolation between slices. Segmentations were manually checked and edited to ensure accuracy. Whole-brain volumes were normalized to TIV and the mean normalized whole-brain volumes in the tau-positive and tau-negative groups were compared in a one-way analysis of variance.

Results

Both the pathological subgroups had significant and extensive cortical volume loss compared with healthy controls (fig. 1): in both the tau-positive and tau-negative cases, atrophy involved predominantly frontal and temporal cortices and was somewhat more pronounced in the right hemisphere. Areas of atrophy that were common to both tau-positive and tau-negative cases clustered in the anterior temporal lobes, frontal opercula and frontal convexities bilaterally. The locations of maximal volume loss within each of these brain regions in the tau-positive and tau-negative subgroups are presented in table 2. No brain regions were involved exclusively in the tau-positive or tau-negative subgroup. However, inspection of figure 1 and table 2 suggests differences in the distribution of atrophy within the common affected regions between the tau-positive and tau-negative subgroups: for example, atrophy was relatively more severe in the right anterior temporal lobe and bilateral prefrontal regions in the tau-positive cases, and relatively more severe in the left posterior temporal lobe in the tau-negative cases. A direct comparison of the tau-positive and tau-negative cases yielded no
Table 2. Locations of maximal volume loss in tau-positive and tau-negative cases

<table>
<thead>
<tr>
<th>Region</th>
<th>Side</th>
<th>Coordinates of maximal volume loss, mm</th>
<th>Z score</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>x</td>
<td>y</td>
</tr>
<tr>
<td>Tau-positive</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Premotor</td>
<td>R</td>
<td>57</td>
<td>−9</td>
</tr>
<tr>
<td>Inferior anterior temporal</td>
<td>R</td>
<td>44</td>
<td>7</td>
</tr>
<tr>
<td>Orbitofrontal</td>
<td>R</td>
<td>6</td>
<td>4</td>
</tr>
<tr>
<td>Frontal operculum</td>
<td>L</td>
<td>−42</td>
<td>3</td>
</tr>
<tr>
<td>Frontal operculum</td>
<td>R</td>
<td>37</td>
<td>−2</td>
</tr>
<tr>
<td>Medial frontal</td>
<td>R</td>
<td>5</td>
<td>25</td>
</tr>
<tr>
<td>Lateral anterior temporal</td>
<td>R</td>
<td>54</td>
<td>8</td>
</tr>
<tr>
<td>Hippocampus</td>
<td>R</td>
<td>34</td>
<td>−13</td>
</tr>
<tr>
<td>Postcentral gyrus</td>
<td>L</td>
<td>−60</td>
<td>−24</td>
</tr>
<tr>
<td>Orbitofrontal</td>
<td>L</td>
<td>−39</td>
<td>34</td>
</tr>
<tr>
<td>Tau-negative</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Frontal operculum</td>
<td>R</td>
<td>36</td>
<td>−1</td>
</tr>
<tr>
<td>Posterior superior temporal</td>
<td>L</td>
<td>−63</td>
<td>−37</td>
</tr>
<tr>
<td>Postcentral</td>
<td>L</td>
<td>−58</td>
<td>−55</td>
</tr>
</tbody>
</table>

Data are derived from comparisons of each of the two FTLD subgroups with the healthy control subjects. Local maxima are shown for both the tau-positive and tau-negative subgroups contrasted with controls. Voxel coordinates are in millimetres after transformation into standard MNI stereotactic space [17]. Only those local maxima exceeding a stringent statistical threshold of \( p < 0.001 \) corrected for multiple comparisons across the entire brain volume are shown.

Discussion

This study suggests that patterns of frontotemporal atrophy do not predict the presence or absence of tau pathology, and conversely, that different immunohistochemical profiles are associated with similar patterns of regional vulnerability to neuronal loss in FTLD. These findings support the view that the topography of atrophy rather than immunohistochemistry determines phenotype in FTLD [3], and emphasize the pathological and genetic heterogeneity of these diseases. The results are consistent with pathological case series that have failed to identify specific macroscopic correlates of tau pathology [12, 13]. However, the apparent lack of correlation between tau pathology and the distribution of atrophy on brain imaging should be interpreted cautiously. The number of patients in each of the pathological subgroups here was relatively small, and the study may, therefore, have lacked sensitivity to detect subtle differences in the cross-sectional profile of atrophy that might emerge in larger populations. The present data (fig. 1, table 2) do suggest differences in the profile of frontotemporal atrophy between tau-positive and tau-negative subgroups; however, these differences did not attain statistical significance. Even if such differences are substantiated in larger cohorts, they are unlikely to be useful in predicting the pathological diagnosis in the individual patient with an established clinical FTLD syndrome. All of the patients in this study fulfilled consensus clinical criteria for the diagnosis of FTLD [15]; it remains possible that pathologically determined differences in the profile of frontotemporal atrophy may be present earlier in the course of these diseases (possibly preceding the onset of clinical symptoms), with convergent involvement of cortical regions as the pathological process becomes more widely distributed. In Alzheimer’s disease, it is known that the pattern of regional atrophy may shift with advancing disease [14, 21]; however, little information is presently available concerning the evolution of atrophy in FTLD. The prospect...
of disease-modifying therapies in the neurodegenerative diseases [1] means that the detection of clinico-radiological correlations would have both maximal specificity and maximal clinical value early in the course of the disease.

It is important to keep in mind that FTLD is highly heterogeneous clinically as well as pathologically, and certain clinical phenotypes such as PNFA are underrepresented in the present sample: specific radiological and pathological correlations may exist for clinically defined subpopulations within the FTLD spectrum, though not for the group considered collectively. In addition, an uncertain proportion of cases classified as tau-negative using currently available pathological and genetic criteria may nevertheless have tau dysfunction, which might produce a similar pattern of tissue destruction [1]. The rightward asymmetric distribution of anterior temporal lobe involvement in both subgroups here might appear somewhat at variance with previous studies describing a leftward asymmetry of temporal atrophy [6, 8, 22]: this difference may reflect the relative preponderance of behavioural and personality disturbance (the frontotemporal dementia syndrome) in the majority (13/17) of cases in the present cohort. In contrast to primary impairments of language (PNFA) or semantic memory (semantic dementia), which are associated with selective involvement of frontotemporal regions in the dominant hemisphere [6, 11], frontal behavioural presentations are frequently associated with non-dominant frontotemporal atrophy [23–25].

It remains possible that differences between pathological subgroups may be detected using imaging parameters other than the cross-sectional extent or severity of volume loss: for example, different rates of global or regional brain atrophy [26]. The present cross-sectional study detected no significant differences in whole-brain volume between tau-positive and tau-negative cases after a similar duration of clinical disease. However, the direct demonstration of differential rates of atrophy would require the prospective analysis of serial volumetric MR images registered into a common stereotactic space; moreover, differential rates of atrophy might manifest at the level of particular brain regions rather than globally [14]. We therefore suggest that the present findings underline the need for further prospective studies of genetically defined ‘at risk’ FTLD populations.

Acknowledgments

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References


VBM and FTLD Immunohistochemistry


